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# RAPID COMMUNICATION

# Construction of a novel predictive model with seven metabolism-related genes for hepatocellular carcinoma by machine learning



The gene signature from the high-risk group may be used to conduct a risk assessment, and to enhance early cancer screening, early diagnosis, and treatment.<sup>1</sup> Models for predicting risk may also be able to help in the decisionmaking process for the clinical management of HCC patients.<sup>2</sup> ScRNA-seq developed rapidly which allows the study of transcriptional activity within an individual cell and enables the discovery of the gene expression of small

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but clinically significant tumor subpopulations.<sup>3,4</sup> In this study, scRNA-seq data of HCC from GSE149614 were used.<sup>5</sup> A UMAP algorithm was employed to visualize scRNA-seq data (Fig. 1A). With a cutoff of |logFC| > 0.5 and adjusted *P* value < 0.05, genes that were differentially expressed between the primary and metastatic tumor samples were identified. MRGs were obtained from the Reactome Pathway Database (https://reactome.org/). Using Venn diagrams, 78 differentially expressed genes were found to be metabolism-related (Fig. 1B and Table S1). The volcano plot of MRGs in HCC samples is shown in Figure S1A. The heatmap shows the top 50 MRG (Fig. S1B).

Subsequently, 26 genes related to the overall survival (OS) of 78 MRGs were selected via uni-Cox regression analysis (Table S2). The LASSO results indicated that eight MRGs were important which were then used as the input genes in the multi-Cox regression analysis (Fig. S1C, D). Multi-Cox regression analysis showed that CYP27A1, CYP2C9, HMGCS2, NQO1, GLB1, PLPP1, and PGAM1 were hub MRGs. Kaplan-Meier analysis showed that high expression of CYP27A1, CYP2C9, HMGCS2, and PLPP1 was correlated with better OS outcomes, while high expression of NQO1 was correlated with worse OS outcomes (Fig. S2). T-SNE results of the expression of CYP27A1, CYP2C9, HMGCS2, NQO1, GLB1, PLPP1, and PGAM1 in single-cell data (GSE149614) were shown in Figure S3. The MRG signatures were calculated based on the relative coefficient and expression of each gene as follows: risk score  $(RS) = -0.00140 \times CYP27A1 - 0.00195 \times CYP2C9$ 0.00072  $\times$  HMGCS2 + 0.00120  $\times$  NQ01 + 0.01625  $\times$  $GLB1 - 0.01114 \times PLPP1 + 0.03327 \times PGAM1$  (Table S3). Each patient in the TCGA-LIHC and ICGC-LIRI-JP cohorts was then classified into MRG high- and low-risk groups by median risk score: TCGA-LIHC is 0.957 and ICGC-LIRI-JP is 0.967 (Table S4). Next, the MRG signature was evaluated

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**Figure 1** Development of a metabolism-related predictive model. (A) Characterization of single-cell RNA sequencing from primary and metastatic HCC cells and screening of marker genes. UMAP analyses were performed to analyze the single-cell RNA sequencing data GSE149614.<sup>5</sup> Group 1, metastatic HCC patients. Group 2, primary HCC patients. (B) Venn diagram showed the 78 metabolism-related genes (MRGs) which were differentially expressed between primary and metastatic HCC patients. (C) ROC analysis of seven core MRG genes, and their combined predictive efficiency in the TCGA-LIHC cohort. (D) Kaplan–Meier analysis of the possibility of overall survival in the TCGA-LIHC cohort with the MRG high- and low-risk scores. (E) Violin plot shows the difference in immune infiltration between the MRG high- and low-risk groups by xCell algorithm.

by ROC curves in the TCGA-LIHC cohort (Fig. 1C) and the ICGC-LIRI-JP cohort (Fig. S4A). In comparison to the patients in the MRG high-risk group, the OS rate of patients in the MRG low-risk group was significantly higher in both the TCGA-LIHC cohort (Fig. 1D) and the ICGC-LIRI-JP cohort (Fig. S4B). Survival analysis of the TCGA-LIHC cohort also indicated that the low-risk group had a higher OS value (Fig. S4C, D).

The nomogram prediction model was established by combining OS-related clinical parameters and RS. The OS-related clinical parameter was analyzed by univariate and multivariate Cox regression analyses (Fig. S5A, B). A clinically applicable nomogram forecasting individual survival probability at 1, 3, and 5 years is shown in Figure S5C. Next, calibration curves, ROC curves, and decision curve analyses were utilized for model validation. The calibration curves (Fig. S6A), ROC curves (Fig. S6B), and decision curve (Fig. S6C-E) analyses implied that the predictive power and clinical practicability of the nomogram prediction model were strong.

Furthermore, we discovered the different characteristics of the tumor mutation burden, gene set enrichment, and immune microenvironment between the MRG high- and low-risk group patients. As shown in Figure S7A and B, the genes with the most frequent mutations were TP53 (42%) and CTNNB1 (33%) in the MRG high- and low-risk groups, respectively. Comparing the waterfall plots of the MRG high-risk group and low-risk group, it was observed that TP53, TTN, and MUC16 had higher frequencies in the MRG high-risk group (Fig. S7A, B). It can also be concluded that the MRG high-risk group suffered high tumor mutation burdens. The significant pathways in the different MRG risk groups were explored by Gene Set Enrichment Analysis. The top six pathways enriched in the MRG high- and low-risk groups were filtered (Fig. S8). The top six pathways that were significantly related to the progression of HCC in the MRG high-risk group were endocytosis, vasopressin-regulated water reabsorption, progesterone-mediated oocyte maturation, RNA degradation, oocyte meiosis, and pancreatic cancer (Fig. S8). In contrast, retinol metabolism, drug metabolism-cytochrome P450, fatty acid metabolism, primary bile acid biosynthesis, glycine, serine and threonine metabolism, and complement and coagulation cascades were primarily screened in the MRG low-risk group (Fig. S8 and Table S5). The immune microenvironment of the MRG high- and low-risk groups was analyzed by the xCell algorithm, and the results are shown in the violin plot in Figure 1E and Table S6. For patients in the MRG highrisk group, naïve CD8+ T cells, common myeloid progenitors, endothelial cells, granulocyte-monocyte progenitors, hematopoietic stem cells, M2 macrophages, and plasmacytoid dendritic cells demonstrated high levels of infiltration (Fig. 1E). However, activated myeloid dendritic cells, B cells, memory CD4<sup>+</sup> T cells, class-switched memory B cells, common lymphoid progenitors, M1 macrophages, mast cells, monocytes, NKT cells, and Th2 CD4<sup>+</sup> T cells were positively correlated with a low RS (Fig. 1E).

In summary, through single-cell RNA sequencing and machine learning, we constructed a novel MRG prognostic model for HCC patients and evaluated its clinical application for the first time. Our prognostic model showed highly accurate risk stratification and accurate identification of HCC patients with poor subtypes, which could predict the prognosis of HCC and guide personalized treatment for HCC patients. We also discovered the different characteristics of tumor mutation burden, gene set enrichment, and immune infiltration in MRG high- and low-risk group patients. These results indicated that the underlying molecular mechanisms were markedly different, which provides a solid basis for the identification and treatment of high-risk HCC patients by the MRG signature.

## Author contributions

Zuhui Pu and Lisha Mou designed the study and revised the manuscript. Lin Liu, Yumiao Qiu, and Yingying Liang performed the analysis and wrote the manuscript.

## **Conflict of interests**

The authors declare no competing interests.

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## Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.gendis.2022.12.014.

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