



REVIEW ARTICLE

Cancer chemotherapy and beyond: Current status, drug candidates, associated risks and progress in targeted therapeutics

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Abstract Cancer is an abnormal state of cells where they undergo uncontrolled proliferation and produce aggressive malignancies that causes millions of deaths every year. With the new understanding of the molecular mechanism(s) of disease progression, our knowledge about the disease is snowballing, leading to the evolution of many new therapeutic regimes and their successive trials. In the past few decades, various combinations of therapies have been proposed and are presently employed in the treatment of diverse cancers. Targeted drug therapy, immunotherapy, and personalized medicines are now largely being employed, which were not common a few years back. The field of cancer discoveries and therapeutics are evolving fast as cancer type-specific biomarkers are progressively being identified and several types of cancers are nowadays undergoing systematic therapies, extending patients' disease-free survival thereafter. Although growing evidence shows that a systematic and targeted approach could be the future of cancer medicine, chemotherapy remains a largely opted therapeutic option despite its known side effects on the patient's physical and psychological health. Chemotherapeutic agents/pharmaceuticals served a great purpose over the past few decades and have remained the frontline choice for advanced-stage malignancies where surgery and/or radiation therapy cannot be prescribed due to specific reasons. The present report succinctly reviews the existing and contemporary advancements in chemotherapy and assesses the status of the enrolled drugs/pharmaceuticals; it also comprehensively discusses the emerging role of specific/targeted therapeutic strategies that are presently being employed to achieve better clinical success/survival rate in cancer patients.

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Introduction

Cancer is a heterogeneous and multifactorial disease in which a series of genomic/molecular alterations cause the uncontrolled growth and proliferation of the cells, causing a rapid increase in tissue mass in the affected parts of the body. Under normal conditions, a cell gets signals to die and to supplant the organism with a young and healthier cell. Cancer cells grow using the body's oxygen and supplements, depriving other cells of regular supplements and growth factors. These cells can turn the microenvironment in their favor, deceive the immune system of the body, and can exploit the physiology of other cells to accommodate their needs.^{1–6} Some of the biomarkers which are currently been used to detect cancer including human epididymis protein 4 (HE4), carcinoembryonic antigen (CEA), legumain, mesothelin, osteopontin, and vitamin E-binding plasma protein. A plethora of anti-cancer drugs and natural medicinal compounds have been devised over the years, which can suppress tumor growth through diverse mechanisms. Some of the drugs/compounds work on crucial

cellular enzymes, while others may alter cell metabolism. They have also shown the potential to interfere with some critical cellular processes, e.g., programmed cell death/apoptosis, drug resistance, DNA damage, DNA replication, or immune reactions. These drugs have distinct modes of action and selectivity for multiple cancer types^{7–12}; however, subtle alteration to their chemical structure could abolish their anti-cancer potency.^{13,14}

The idea of chemotherapy (utilizing toxic compounds and drugs to destroy cancerous cells) came into existence after the reports of mustard gas killing lymphatic tissues and bone marrow. The effects were later validated in mice using an effective derivative of the gas (nitrogen mustard) that showed effective regression of lymphoma tissues.^{15–17} The first patient receiving this nitrogen mustard was forty-eight years old lymphosarcoma patient whose cancer softened and cleared initially. Unfortunately, later he died after relapsing, but this secret military trial at Yale University paved the way for chemicals in treating cancers and developing the field of cancer chemotherapy for treating a variety of cancers.^{18–21}

Chemotherapy primarily works by circumventing the cancer cells from further growth and division. Cancer cells usually divide and grow at a much-accelerated rate than normal cells and are physiologically possess very high endogenous stress. Therefore, the drugs can destroy them rapidly and more effectively in comparison to other surrounding cells. Some of the promising inhibitor therapies which have been used to treat solid cancers are polyadenosine diphosphate-ribose polymerase inhibitors, angiogenesis inhibitors, histone deacetylase (HDAC) inhibitors, mechanistic target of rapamycin (mTOR) inhibitors, poly (adenosine di-phosphate-ribose) polymerase (PARP) inhibitors, p53/mouse double minute 2 homolog (MDM2) inhibitors, hedgehog pathway blockers, tyrosine kinase inhibitors and proteasome inhibitors.^{22,23} Chemotherapy may have different variations, which can affect the target cells in distinct manners.²⁴ Some of the treatments may directly alter the quality of cellular proteins, rendering them non-functional and affecting major cellular physiological pathways. Major chaperone repressors, autophagy suppressors, or proteasome inhibitors belong to these classes of molecules.^{25–31} Other drugs may target some essential hormones and interfere with the body's overall metabolism.^{32,33} The 2018 Nobel Prize attributed to immune checkpoint therapy research has highlighted the importance of immunotherapy, which underlies the possibilities of modulating our immune system in our favor to cope up with the cancer-like conditions and fight back against the disease.^{24,34}

The selection of chemo-preventive drugs or a combination of drugs is mostly based on the type and stage of cancer. The primary objective of these drugs is to neutralize the cancerous cells and ameliorate the stress caused by the tumor's growth. The dosage and timespan of the treatment are two important points of consideration. It has been observed that the medications are given at very high doses causing numerous side effects and harm to other healthy cells.³⁵ One major challenge is the relapse of the disease. During reoccurrence, it has been widely observed that the cancer cells attain drug resistance following longer exposure to the drugs. Despite enormous applications, most chemotherapies are given to prolong and ease the patient's survival, hence called palliative chemotherapy.²¹ However, prolonged exposure to these drugs may have excessively adverse effects on a patient's physical and mental health, making it difficult to continue the ongoing treatment. Oncologists, for the most part, provide these medications with possible time intervals in between, which also gives non-transformed cells a chance to mend.³⁶ In most cases, a team of specialists, including radiologists, nutritionists, psychologists, and the primary consultant collectively decide the treatment plans that include drug combinations, dosage, duration of cycles, and additional supplements.

This report mainly reviews various aspects of cancer chemotherapy, including various applications of the drugs/chemotherapeutic agents that are already in practice. We also succinctly discussed various side effects and drawbacks of these treatment plans and further provided an overview of recent advancements in interventional strategies by focusing on targeted drug therapy, immunotherapy,

personalized and integrative medicines. [Figure 1](#) depicts the significant intrinsic and extrinsic cellular and molecular events causing the transformation of a normal cell to a cancer cell and its distinct hallmarks.

Cancer chemotherapeutics: what is known so far?

The idea of cancer as a disease was known to Hippocrates, the father of medicine, around 400 BC. He later coined the term 'carcinos' due to the similarity of cancer tissue growths with crabs. The term was later replaced with the Latin word cancer by Celsus. Many ancient pieces of literature have described several kinds of malignancies in humans, and multiple treatment approaches were also in use in different parts of the ancient world, as found in the literature of that era.^{37,38} However, the progress of strategies to treat the disease was more or less static until the beginning of the twentieth century.³⁹ The extract of the plant *Colchicum autumnale* was used by Greek physicians to dissolve the tumor mass. Interestingly, in the 1930s, the active compound in the extract, colchicine, was reported that can interfere with microtubule assembly and may work as a potential drug.⁴⁰ Vincristine and vinblastine are other similar compounds that were medicinally recognized as possible anti-tumor agents a long time ago.⁴¹ The ancient pieces of the literature suggest that the Greek and Roman physicians have contributed enormously to the identification and characterization of disease. Arabic, Indian, and Chinese medicinal archives have also paved the way for multiple medieval and modern approaches to understand and treat the disease. Interestingly, for centuries, most of the approaches remain limited to cauterization, bloodletting, surgery, herbal medicine, etc.⁴² In modern onco-medicine, which was evolved during the Second World War, many active compounds were reported following the detailed characterization of the disease at the genetic and molecular level. The initial decades have seen enormous debate on the use of cytotoxic agents as drug candidates; however, advancements were made, and success came in the form of Eli Lilly introducing *Vinca* alkaloids in the 1960s, which was then followed by procarbazine.⁴³ In the following years, the idea of combination therapy was introduced for acute lymphocytic leukemia and non-Hodgkin's lymphoma patients that were also supplemented with supportive medical care to enhance the quality of life.²¹

In the second half of the twentieth century, a series of reports have identified numerous mutations, external stimuli (toxic chemicals, viruses, etc.), metabolic alterations, etc. as the underlying causes of cancer.^{44,45} Identification of a plethora of cellular pathways that are affected in various cancer types has given different molecular alternatives to target and develop effective drugs ([Fig. 2](#)).^{46–48} Several modes of medications have also been introduced and tried for drug administration based on the types and stages of the diagnosed cancer. Here, we discuss the available drug medications, following which we have

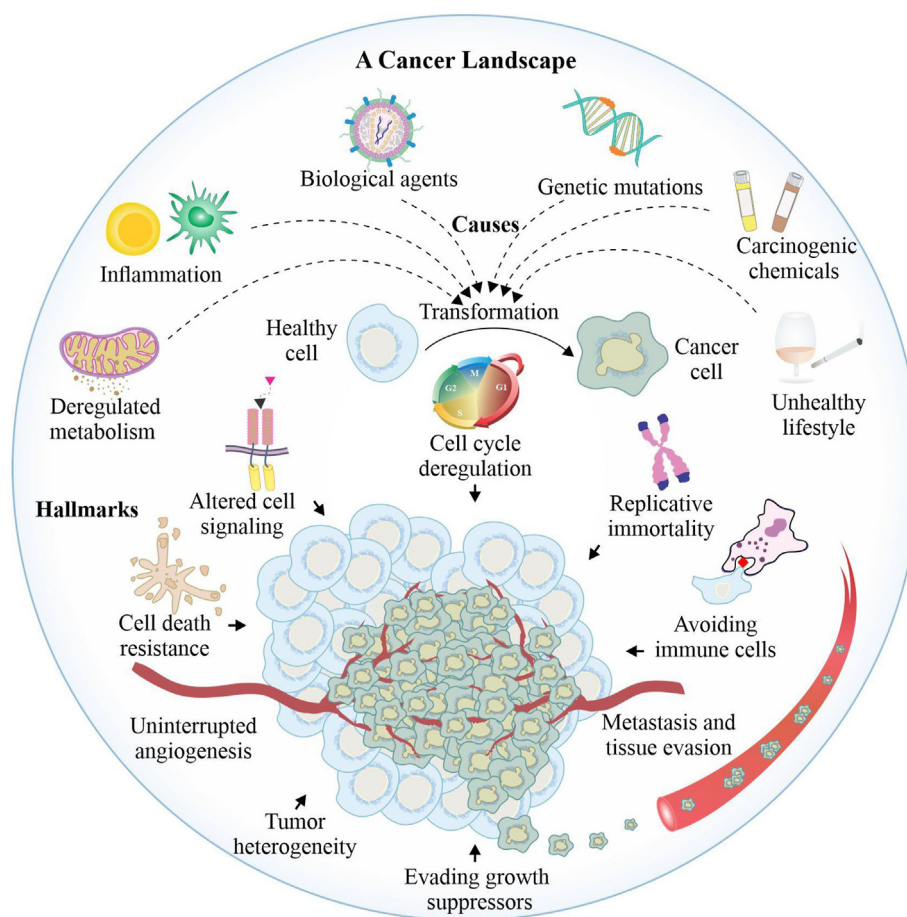


Figure 1 Schematic diagram showing different intrinsic and extrinsic biological/molecular events potentiating the transformation of a normal cell to the cancer cell, while the lower part of this diagram showing different hallmarks of a transformed cancer cell.

revisited the chemotherapeutic agents, which have substantially pushed the field forward and extended the lives of millions in the past several decades.

Intravenous (IV) chemotherapy requires drug infusion straightforwardly into the patient's veins that may take from minutes to a few hours. Some medications can also be given either as pills or fluid regularly or at some intervals.⁴⁹ Intramuscular shots and intra-abdominal administration are other methods of giving chemotherapeutic medications to target the specific locations of tumor mass.⁵⁰ In addition, a new technology called pressurized intraperitoneal aerosol chemotherapy has been developed to efficiently deliver intraperitoneal chemotherapy for end-stage peritoneal metastases patients.⁵¹ Topical treatment methods imply providing drugs through the skin surface. The anti-cancer compounds derived from either natural sources or their synthetic chemical analogs can kill cancer cells or stop their development.^{52–54} Notably, various other approaches are emerging to treat the cancer cells, including small molecule inhibitors,⁵⁵ a combination of anti-cancer agents and immunotherapy,⁵⁶ nanotechnology-based drugs,^{57–59} and organo-seleno compounds (Fig. 2).³¹

Nevertheless, the selectivity of most drug particles is restricted, and such medications are considered one of the most harmful medications utilized in the treatment.^{60,61} Since the initial findings of modern anticancer drug discoveries in the 1940s, continuous efforts are being made to treat this life-threatening disease. Advances in scientific methods and techniques have brought new treatment and diagnostic tools and advanced patient care measures (Fig. 2).^{62–69} Treatment plans may have palliation as one strategy that may lead to shrinkage of the obvious tumor, easing side effects, and increasing the life quality of patients. Figure 2 describes various ways of providing anti-cancer medications to patients depending upon their type and stage of the disease.

Along with primary treatment, adjuvant therapy is also given to the patients for additionally combating the disease through supportive strategies.^{70,71} Surgery and radiation can be given to patients to supplement a chemotherapeutic plan to curb the disease by destroying shrouded cancer cells that could not be targeted otherwise.⁷² Sometimes adjuvant chemotherapy is given to patients after surgery or radiation or both to lessen the chances of recurrence.^{73–75}

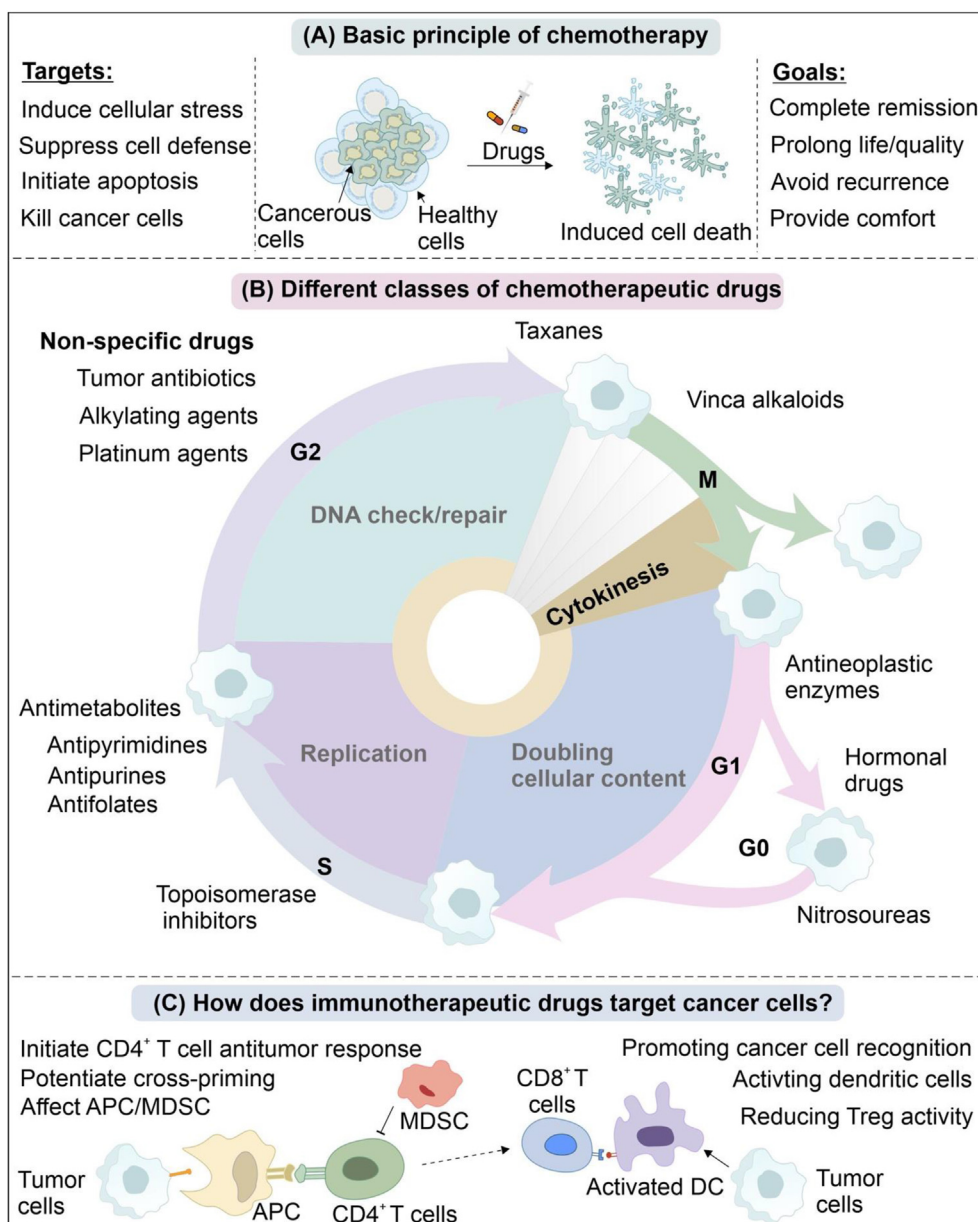


Figure 2 Schematic diagram showing the basic principles of chemotherapy (A), different classes of chemotherapeutic agents/drugs (B), and immunotherapeutic mechanism(s) of targeting cancer cells (C).

Adjuvant chemotherapy starts typically within three to five weeks of the surgical removal of a tumor and has distinctive treatment lengths relying upon the disease. Although chemotherapy has significantly improved overall survival, patients experience a wide range of physical and psychological symptoms that impact their quality of life. Symptoms seldom occur in isolation. Chemotherapy-induced hair loss remains greatly feared, with a negative impact on the well-being of many cancer patients. Cancer- and chemotherapy-induced poor appetite is usually the result of taste changes, mouth sores, nausea and vomiting, increased satiety, medication side effects, pain, fatigue, depressed mood, and anxiety.

Basic principles of chemotherapy: years of research, miles to go

The cancer cell metabolic mechanisms functionally overlap with the host cells, so cancer treatment is very challenging.^{76,77} Designing drugs or therapeutics mainly focuses on selectivity, which can specifically kill the cancerous cells without affecting the non-cancerous cells (Fig. 2). In a way, the anti-cancer therapy may resemble antimicrobials, but the cancerous cells and the bacterial cells have differences in terms of the metabolic and physiological characters.^{78–80} For our immune cells, bacterial cell recognition is straightforward, while our transformed cells can hide cell surface

receptors to evade the immune system.⁸¹ Early-stage detection of cancers increases the chances of treatment and survival. Therefore, recognizing and diagnosing those transformed cells remains one of the most challenging tasks in oncological research. At early stages, most known biomarkers remain undetectable, and cancer cells' ability to hide from immune cells freezes our immune responses.^{82,83}

In a tumor subpopulation, transformed cells differ drastically from the other cells in terms of physiology, metabolism, proliferation rate, etc.^{84–86} Besides that, the genetic diversity may pose additional challenges to target all kinds of cells in a particular tissue mass owing to the tumor heterogeneity.^{87,88} A combination of drug therapy is applied to address this challenge, and the regime is devised according to the cancer phase and type (Fig. 2).^{89,90}

In the subsequent section, we provide a comprehensive overview of various chemotherapeutic agents which have served the purpose for several decades in different cancer types. These drugs could be classified in different ways based on their chemical nature/source, molecular target, mechanism/mode of action, or effectiveness in model systems against various malignancies/cancer types (Table 1).

Alkylating agents

One major class of the current drugs belongs to alkylating agents, which can interfere with the formation/linkage of DNA double strands.⁹¹ These drugs can cause it by transferring one alkyl group to the guanidine base in DNA (Fig. 2). Cross-linking of nucleic acids with proteins or peptides can likewise affect the DNA geometry, cause erroneous base pairing and DNA strand breakage, and eventually preventing the cell division and causing irreversible senescence.⁹² Mechanistically, alkylating agents in their electrophilic forms interact with cellular DNA and form covalent adducts, underlying their broader utility. Therefore, alkylating drugs are the frontline chemotherapeutic agents given against most of the cancer types, yet the greatest therapeutic value of their inhibition is known for slow-growing cancers. In Table 1, we described the common chemotherapeutic alkylating agents including the altretamine, chlorambucil, platinum drugs (cisplatin, carboplatin, oxaliplatin), doxorubicin, epirubicin, etoposide, mitoxantrone, cyclophosphamide, thiotepa, and busulfan that are in clinical practice and also provided details of their molecular targets, mode of action, and studied model systems in various malignancies/cancer-types.

Antimetabolites

Antimetabolites are the next class of substances that may compete, replace, or inhibit some specific metabolites inside the cells and thus meddle with the cellular metabolism (Fig. 2).⁹³ These molecules mostly possess a structure similar to a cellular metabolite or enzyme–substrate, usually identified and processed by an enzyme to meet the cellular needs.⁹⁴ Tetrahydrofolates are formed from

unreduced dietary folates by the enzyme dihydrofolate reductase. For example, various medications, such as aminopterin, methotrexate (amethopterin), pyrimethamine, trimethoprim, and triamterene, act directly on the production of these metabolites, hindering the enzyme from producing folate insufficiency.⁹⁵ Methotrexate, an antifolate drug, is one of the exceptionally successful anticancer medications that represses dihydrofolate reductase (DHFR) and obstructs the transformation of dihydrofolic acid (DHFA) into tetrahydrofolic acid (THFA).⁹⁶ This coenzyme activity is indispensable to the cells as they are essential for the nucleotide synthesis pathways. Methotrexate is an anti-inflammatory molecule and is hence used as a medication for many other inflammatory diseases, like rheumatoid arthritis and extreme psoriasis, apart from various cancer types. A detailed description of common antimetabolites is comprehensively provided in Table 1.

Pyrimidine-derived antimetabolites (pyrimidine antagonists) and *Vinca* alkaloids

Pyrimidine-derived antimetabolites are a group of chemotherapeutic agents that interrupt DNA synthesis and exert their cytotoxicity to cancer cells (Fig. 2). For instance, cytarabine also known as cytosine arabinoside or Ara C interrupts DNA as well as RNA synthesis by displacing the cytosine with itself (that differs merely by its base sugar arabinose). Cytarabine/cytosine arabinoside is enrolled frequently in clinics to treat leukemia or non-Hodgkin's lymphoma.⁹⁷ Some other notably pyrimidine-derived antimetabolites include fluorouracil, 5-fluorouracil (5-FU), capecitabine, floxuridine, gemcitabine, decitabine, raltitrexed, and tegafur (Table 1).^{98–103} Being as pyrimidine or purine analogs having altered chemical groups, these drugs/compounds induce apoptosis while progressing through the S phase of the cell cycle by their misincorporation to RNA and DNA or by inhibiting the key enzymes required for nucleic acid synthesis. Among these enzymes, DNA polymerases, ribonucleotide reductase, and thymidylate synthetase are the main intracellular targets of the pyrimidine antimetabolites. 5-FU has been a promising pyrimidine analog that largely interrupts DNA and RNA synthesis. 5-FU that depends on its incorporation to DNA inhibits mitosis and stimulates cell death in the dividing cells.

Vinca alkaloids, primarily extracted from several species under the *Vinca* genus, are mainly cell division inhibitors, which can interact with microtubular protein tubulin to obstruct its polymerization and meddle with the cytoskeleton, leading to stalling of mitotic division.¹⁰⁴ The chromosomes do not separate efficiently during cell division, leading to metaphase capture. Vincristine and vinblastine are primary examples of this drug class with different degrees and ranges of anti-tumor activity (Table 1).^{105,106} Cancers treated with these drugs include acute leukemia, Hodgkin's and non-Hodgkin's lymphoma, rhabdomyosarcoma, Ewing's sarcoma, neuroblastoma, Wilms' tumor, miscellaneous myeloma, chronic leukemia, thyroid cancer, brain tumors, and several blood-related disorders.¹⁰⁷

Table 1 Table listing chemotherapeutic drugs/agents used in current clinical practice along with details of their molecular targets, mode of action, and investigated model systems across diverse malignancies.

Drugs/Compound	Molecular targets	Mechanism of action	Studied model system	Reference/Source
Antineoplastic alkylating agents				
Altretamine	DNA polymerase alpha catalytic subunit Ribonucleoside-diphosphate reductase large subunit DNA	Inhibitor Inhibitor Other/unknow	B-cell chronic lymphocytic, leukemia/prolymphocytic leukemia/small lymphocytic lymphoma refractory, and refractory non-Hodgkin's lymphoma	278
Bendamustine	DNA	Intra- and inter-strand crosslinking	Chronic lymphocytic leukaemia (CLL), follicular non-Hodgkin's lymphoma refractory, refractory Hodgkin lymphoma, refractory Mantle cell lymphoma, Waldenström's macroglobulinemia (WM), recurrent multiple myeloma, and refractory indolent B cell non-Hodgkin lymphoma	279
Busulfan	DNA	Cross-linking/alkylation	Chronic myelogenous leukemia	279
Carboplatin	DNA	Cross-linking/alkylation	Advanced cervical cancer, advanced Endometrial Cancer, advanced esophageal cancers, advanced head and neck Cancer, advanced melanoma, advanced non-small cell lung cancer, advanced ovarian carcinoma, advanced Sarcoma, metastatic breast cancer, neuroendocrine carcinoma of the skin, pleural mesothelioma, refractory Hodgkin lymphoma, retinoblastoma, advanced bladder cancer, advanced small cell lung cancer, advanced testicular cancer, advanced thymoma, advanced thymic carcinoma, and refractory non-Hodgkin's lymphoma	279
Carmustine	DNA Glutathione reductase, mitochondrial RNA	Other/unknown inhibitor Other/unknown	Glioblastoma, brainstem glioma, medulloblastoma, astrocytoma, ependymoma and metastatic brain tumors, multiple myeloma, Hodgkin's disease, non-Hodgkin's	280

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Table 1 (continued)

Drugs/Compound	Molecular targets	Mechanism of action	Studied model system	Reference/Source
Chlorambucil	DNA	Cross-linking/alkylation	lymphomas, and may be used on the skin (topically) for cutaneous T-cell lymphoma	279
Cisplatin	DNA DNA-3-methyladenine glycosylase Alpha-2-macroglobulin Serotransferrin Copper transport protein ATOX1	Cross-linking/alkylation Not Available Not Available Not Available Not Available	Chronic lymphocytic leukemia, lymphomas, and Hodgkin's disease Testicular cancer, ovarian cancer, bladder cancer, head and neck cancer, esophagus cancer, small cell and non-small cell lung cancer, non-Hodgkin's lymphoma and trophoblastic neoplasms	279
Cyclophosphamide	DNA Nuclear receptor subfamily 1 group I member 2	Cross-linking/alkylation Not Available	Lymphoma, multiple myeloma, leukemia, ovarian cancer, breast cancer, small cell lung cancer, neuroblastoma, and sarcoma.	279
Dacarbazine	DNA DNA polymerase alpha subunit B 6-phosphogluconate dehydrogenase, decarboxylating	Cross-linking/alkylation Other/unknown Inhibitor	Melanoma skin cancer, soft tissue sarcoma, Hodgkin lymphoma	279
Daunorubicin	DNA DNA topoisomerase 2-alpha DNA topoisomerase 2-beta	Intercalation Inhibitor Inhibitor	Acute nonlymphocytic leukemia in adults and acute lymphocytic leukemia in adults and children	279
Decitabine	DNA DNA (cytosine-5)-methyltransferase 1 DNA (cytosine-5)-methyltransferase 3A DNA (cytosine-5)-methyltransferase 3B	Other/unknown Inhibitor Inhibitor Inhibitor	Myelodysplastic syndromes, and acute myeloid leukemia (AML)	279,281
Doxorubicin	DNA DNA topoisomerase 2-alpha Nucleolar and coiled-body phosphoprotein 1	Intercalation Inhibitor Not available		279
Epirubicin	DNA topoisomerase 2-alpha DNA	Inhibitor Intercalation	Breast cancer	279,282
Etoposide	DNA topoisomerase 2-alpha DNA topoisomerase 2-beta	inhibitor Inhibitor	Testicular cancer, lung cancer, lymphoma, nonlymphocytic leukemia, neuroblastoma, and ovarian cancer, Kaposi's sarcoma, Ewing's sarcoma and glioblastoma	279

Idarubicin	DNA DNA topoisomerase 2-alpha	Intercalation Inhibitor	multiforme Acute myelogenous, acute lymphoblastic and chronic myelogenous	279
Ifosfamide	DNA Nuclear receptor subfamily 1 group I member 2	Other/unknown Not available	Germ cell testicular cancer, soft tissue sarcoma, acute lymphocytic leukemia, bladder cancer, bone cancer, pancreatic cancer, stomach cancer, breast cancer, cervical cancer, endometrial cancer, lung cancer, ovarian cancer, neuroblastoma, lymphoma, and Wilms' tumor	279
Irinotecan	DNA topoisomerase 1, DNA topoisomerase I, mitochondrial	Inhibitor Inhibitor	Metastatic colorectal cancer, other colorectal cancers and non-small/small lung cancers	279
Lomustine	DNA Stathmin-4	Cross-linking/alkylation Antagonist	Metastatic brain tumors, Hodgkin's and non-Hodgkin's lymphoma	279
Mechlorethamine	DNA	Intercalation	Hodgkin's disease, chronic lymphocytic leukemia, chronic myelogenous leukemia, small cell lung cancer, medulloblastoma, and non-Hodgkin's lymphoma	283
Melphalan	DNA	Cross-linking/alkylation	Multiple myeloma, ovarian cancer, breast cancer, melanoma, and amyloidosis	284
Mitomycin C	DNA	Cross-linking/alkylation	Gastric cancer, anal and colon cancer, breast cancer, non-small cell lung cancer, head and neck cancer, small bladder papillomas, pancreatic cancer, cervical cancer	279
Mitoxantrone	DNA DNA topoisomerase 2-alpha	Intercalation Inhibitor	Advanced, hormone-refractory prostate cancer, acute nonlymphocytic leukemia (ANLL), breast cancer and non-Hodgkin's lymphoma.	285
Oxaliplatin	DNA	Cross-linking/alkylation	Metastatic or recurrent colorectal cancer, metastatic pancreatic cancer and metastatic gastric cancer.	286
Temozolomide	DNA	Cross-linking/alkylation	Brain tumors (astrocytomas) in adult patients, metastatic melanoma and	287

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Table 1 (continued)

Drugs/Compound	Molecular targets	Mechanism of action	Studied model system	Reference/Source
Topotecan	DNA topoisomerase 1	Inhibitor	glioblastoma multiforme	288
Thiotepa	DNA topoisomerase I, mitochondrial DNA	Inhibitor Intercalation Cross-linking/alkylation	Ovarian cancer, small cell lung cancer, and certain types of cervical cancer Adenocarcinoma of the breast, adenocarcinoma of the ovary, papillary thyroid cancer and bladder cancer, Hodgkin's and non-Hodgkin's lymphomas	
Trabectedin	DNA	Binder	Soft tissue sarcoma, and ovarian cancer	290
Antineoplastic antimetabolites				
Azathioprine	Ras-related C3 botulinum toxin substrate 1	Modulator	Leukemia and lymphomas	291
Cladribine	Ribonucleoside-diphosphate reductase large subunit	Inhibitor	Drug-resistant T-cell prolymphocytic leukemia	292
	Ribonucleoside-diphosphate reductase subunit M2	Inhibitor		
	Ribonucleoside-diphosphate reductase subunit M2 B	Other/unknown		
	DNA	Inhibitor		
	DNA polymerase alpha catalytic subunit	Inhibitor		
	DNA polymerase epsilon catalytic subunit A	Inhibitor		
	DNA polymerase epsilon subunit 2	Inducer		
	DNA polymerase epsilon subunit 3			
	DNA polymerase epsilon subunit 4			
	Purine nucleoside phosphorylase			
Clofarabine	DNA polymerase alpha catalytic subunit	Inhibitor	Relapsed or refractory acute lymphoblastic leukaemia, acute myeloid leukaemia, juvenile myelomonocytic leukaemia, and myelodysplastic syndrome	293
	Ribonucleoside-diphosphate reductase large subunit	Inhibitor		
	DNA	Other/unknown		
Fludarabine	Ribonucleoside-diphosphate reductase large subunit	Inhibitor	Chronic lymphocytic leukemia, non-Hodgkin's lymphoma, acute myeloid leukemia, and acute lymphocytic leukemia	294
	DNA polymerase alpha catalytic subunit	Incorporation into and destabilization		
	DNA	Agonist		
	Deoxycytidine kinase			
Gemcitabine	DNA	Cross-linking/alkylation	Breast cancer, ovarian cancer,	295

	Ribonucleoside-diphosphate reductase large subunit	Inhibitor	non-small cell lung cancer, pancreatic cancer and bladder cancer	
	Thymidylate synthase	Inhibitor		
	UMP-CMP kinase			
Mercaptopurine	Hypoxanthine-guanine phosphoribosyltransferase	Inhibitor	Acute lymphocytic leukemia (ALL), chronic myeloid leukemia (CML), Crohn's disease and ulcerative colitis.	296
	Amidophosphoribosyltransferase	Inhibitor		
	Inosine-5'-monophosphate dehydrogenase			
Nelarabine	DNA	Incorporation into and destabilization	Acute T-cell lymphoblastic leukemia	297
	DNA polymerase alpha catalytic subunit	Inhibitor		
Pentostatin	Adenosine deaminase	inhibitor	Hairy cell leukaemia refractory to alpha interferon	298
Tioguanine	DNA	Intercalation	Acute leukemia	299
Pyrimidine-derived antimetabolites (pyrimidine antagonists)				
Capecitabine	Thymidylate synthase DNA RNA	Inhibitor	Metastatic breast, gastric and colorectal cancers	99
		Incorporation into and destabilization		
		Incorporation into and destabilization		
Cytarabine	DNA polymerase beta	Inhibitor	Acute myeloid leukemia (AML), acute lymphocytic leukemia (ALL), chronic myelogenous leukemia (CML) and non-Hodgkin's lymphoma	97
	DNA	Cross-linking/alkylation		
Decitabine	DNA	Other/unknown	Myelodysplastic syndromes (MDS)	101
	DNA (cytosine-5)-methyltransferase 1	Inhibitor		
	DNA (cytosine-5)-methyltransferase 3A	Inhibitor		
	DNA (cytosine-5)-methyltransferase 3B			
Gemcitabine	DNA	Cross-linking/alkylation	Metastatic ovarian cancer, metastatic non-small cell lung cancer, and metastatic pancreatic adenocarcinoma	100
	Ribonucleoside-diphosphate reductase large subunit	Inhibitor		
	Thymidylate synthase	Inhibitor		
	UMP-CMP kinase			
Fluorouracil	Thymidylate synthase	Other/unknown	Actinic keratosis (AK), breast cancer, malignant neoplasm of colon, malignant neoplasm of pancreas, malignant neoplasm of stomach, rectal carcinoma, superficial basal cell carcinoma,	98
	DNA	Incorporation into and destabilization		
	RNA	Incorporation into and destabilization		

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Table 1 (continued)

Drugs/Compound	Molecular targets	Mechanism of action	Studied model system	Reference/Source	
			and hyperkeratotic actinic keratosis		
Raltitrexed	Thymidylate synthase	Inhibitor	Advanced Colorectal Cancer,	102	
	Folylpolyglutamate synthase, mitochondrial	Antagonist	Pleural Mesotheliomas		
Tegafur	Thymidylate synthase	Inhibitor	Advanced Gastric Cancer	103	
Other antineoplastic agents (belong to one or multiple drug classes)					
Arsenic trioxide (Trisenox®)	Inhibitor of nuclear factor Kappa-B kinase subunit beta	Inducer	Refractory Acute Promyelocytic Leukemia	300	
	Thioredoxin reductase 1, cytoplasmic	Inhibitor			
	Transcription factor AP-1	Inducer			
	G1/S-specific cyclin-D1	Inducer			
	Mitogen-activated protein kinase 3	Inducer			
	Mitogen-activated protein kinase 1 inducer	Not available			
	RAC-alpha serine/threonine-protein kinase	Not available			
	Cyclin-dependent kinase inhibitor 1	Not available			
	Histone deacetylase 1				
	Protein PML				
Hydroxyurea	Ribonucleoside-diphosphate reductase large subunit	Inhibitor	Melanoma, resistant chronic myelocytic leukemia, and recurrent, metastatic, or inoperable carcinoma of the ovary and Sickle-cell anemia		301
Paclitaxel (Taxol®)	Tubulin beta-1 chain	Inhibitor	Kaposi's sarcoma and cancer of the lung, ovarian, and breast		302
	Apoptosis regulator Bcl-2	Inhibitor			
	Microtubule-associated protein 4	Not Available			
	Microtubule-associated protein 2	Not Available			
	Microtubule-associated protein tau	Not Available			
	Nuclear receptor subfamily 1 group I member 2	Inducer			
Docetaxel (Taxotere®)	Tubulin beta-1 chain	Not available	Locally advanced or metastatic breast cancer, locally advanced or metastatic non-small cell lung cancer, hormone refractory metastatic prostate cancer, gastric adenocarcinoma and head	303	
	Apoptosis regulator Bcl-2	Not available			
	Microtubule-associated protein 2	Not available			
	Microtubule-associated protein 4	Not available			
	Microtubule-associated protein tau	Not available			
		Binder			

Ixabepilone (Ixemptra®)	Nuclear receptor subfamily 1 group I member 2 Tubulin beta-3 chain	Inhibitor	and neck cancer Breast cancer, head and neck cancer, melanoma, lung cancer, non-Hodgkin's lymphoma, prostate cancer, renal cell carcinoma	279
Methotrexate	Dihydrofolate reductase Humans Thymidylate synthase Bifunctional purine biosynthesis protein PURH	Inhibitor Inhibitor Inhibitor	Pediatric acute lymphoblastic leukemia, gestational choriocarcinoma, choriadenoma destruens, breast cancer, epidermoid cancer of the head and neck, lung cancer, and advanced non-Hodgkin's lymphoma	304
Pemetrexed (Alimta®)	Thymidylate synthase Bifunctional purine biosynthesis protein PURH Dihydrofolate reductase Trifunctional purine biosynthetic protein adenosine-3	Inhibitor Inhibitor Inhibitor Inhibitor	Malignant pleural mesothelioma, and metastatic non-small cell lung cancer (NSCLC)	305
Streptozocin	DNA Solute carrier family 2, facilitated glucose transporter member 2 O-GlcNAcase BT_4395 Bifunctional protein NCOAT	Cross-linking/alkylation Ligand Antagonist Not available	Malignant neoplasms of pancreas	306
Vinblastine	Tubulin alpha-1A chain Tubulin beta chain Tubulin delta chain Tubulin gamma-1 chain Tubulin epsilon Transcription factor AP-1	Adduct Adduct Adduct Adduct Adduct Other/unknown	Breast cancer, testicular cancer, lymphomas, neuroblastoma, Hodgkin's and non-Hodgkin's lymphomas and Kaposi's sarcoma	307
Vincristine	Tubulin beta-1 chain	Inhibitor	Acute lymphocytic leukemia (ALL), Hodgkin lymphoma, non-Hodgkin's lymphomas, Wilms' tumor, neuroblastoma, rhabdomyosarcoma, relapsed Philadelphia chromosome-negative (Ph-) acute lymphoblastic leukemia (ALL)	308
Vindesine	Tubulin beta-1 chain	Inhibitor	Acute leukaemia, malignant lymphoma, Hodgkin's disease	309
Vinorelbine	Tubulin beta chain	Antagonist inhibitor	Advanced non-small cell lung cancer (NSCLC), relapsed or	310

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Table 1 (continued)

Drugs/Compound	Molecular targets	Mechanism of action	Studied model system	Reference/Source
			refractory Hodgkin lymphoma, metastatic squamous cell head and neck cancer, recurrent ovarian cancer, metastatic breast cancer	

Other antineoplastic drugs/agents

Besides these three established classes, several other antineoplastic agents are also enrolled in the clinical practice of cancer chemotherapy (Table 1). Given their clinical promises, in the subsequent section, we described the biological activities, targets, and mechanisms of key chemotherapeutic drugs that may belong to one or multiple drug classes and can affect more than one cellular pathway in the cancer cells and induce apoptosis by different mechanisms (Fig. 2). A succinct review of well-known anti-cancer drugs that are in routine clinical practice is provided as follows.

Paclitaxel/Taxol

It is a complex diterpene taxane ring-containing drug acquired from the bark of the Western yew tree (*Taxus brevifolia*), which induces cytotoxic activity by a novel system.¹⁰⁸ It increases the polymerization of tubulin, thereby inducing a mechanism inverse to that of the aforementioned alkaloids.¹⁰⁹ It forestalls the microtubule's structural adjustment and depolymerization, bringing an inherent cytotoxic activity to the paclitaxel due to the significance of tubulin-microtubule dynamics.¹¹⁰ The confirmed antagonistic impacts of paclitaxel are reported on the metastatic ovarian and bosom carcinomas after the disappointment from first-line chemotherapy and backslide cases.¹¹¹ It has also shown efficacy against the advanced stage tumors of the head and neck disease, small cell lung cancer, oesophageal adenocarcinoma, and prostate cancer (Table 1).^{112,113} Docetaxel is a more potent relative of paclitaxel with a comparable activity. It has been shown to provide benefits against the bosom and ovarian cancers, which get resistant to first-line chemotherapeutic drugs.¹¹⁴ Unfortunately, many side effects have been reported from the long exposure to these drugs, which mainly include neutropenia, arrhythmia, neuropathy, and cardiovascular breakdown (Table 2).^{115,116}

Etoposide

Etoposide is a semi-synthetic derivative of a plant glycoside called podophyllotoxin.^{117,118} It causes DNA damage by altering DNA topoisomerase-II function, and posing G2 cell cycle arrest.¹¹⁹ Etoposide creates a ternary complex with DNA and the topoisomerase-II complex, preventing the latter's binding to the DNA strands, while in doing so, it causes damage to DNA strands.¹²⁰ Cancer cells depend on this enzyme more than healthy cells, as they divide more quickly; therefore, it causes defects in DNA synthesis that promotes apoptosis of the cancer cells.¹²¹ Etoposide is effective against several aggressive malignancies including Kaposi's sarcoma, Ewing's sarcoma, testicular disease, lung disease, lymphoma, non-lymphocytic leukemia, and glioblastoma multiforme (Table 1).¹²²

Table 2 Table listing the adverse effects of various chemotherapy drugs, affected organs, their reported toxicities/side effects and current stage of their clinical progresses.

Drugs/Compound	Affected organ	Reported toxicity/side effects (frequency in >30% cases)	Current status	Reference/Sources
Alkylating agents/drugs				
Altretamine	Blood	Nausea and vomiting, Low blood counts, Peripheral neuropathy	Approved	278
Bendamustine	Blood	Low blood counts, Increase in bilirubin levels	Approved, Investigational	311,312
Busulfan	Blood	Low blood counts, Nausea and vomiting (usually mild with standard doses), Loss of fertility, Diarrhea (usually mild with standard doses), Poor appetite, Mouth sores	Approved, Investigational	313,314
Carboplatin	Cervix, skin, endometrium, esophagus, head and neck, lung, bladder, thymus, mesothelium, testicle	Low blood counts, Nausea and vomiting, Taste changes, Hair loss, Weakness, Blood test abnormalities, abnormal magnesium level	Approved	315
Carmustine	Brain, skin	Nausea and vomiting, Facial flushing, Pain and burning at the injection site, Low blood counts	Approved, Investigational	280
Chlorambucil	Blood	Low blood counts	Approved	316,317
Cisplatin	Testicle, ovary, head-neck, lung, blood, esophagus, trophoblast	Nausea and vomiting, Low blood counts, Kidney toxicity, Ototoxicity, Blood test abnormalities (low magnesium, low calcium, low potassium)	Approved	318,319
Cyclophosphamide	Blood, ovarian, breast, lung, brain	Low blood counts, Hair loss, Nausea and vomiting, Poor appetite, Discoloration of the skin or nails	Approved, Investigational	320
Dacarbazine	Skin, blood	Local pain, burning sensation and irritation at the needle site during the infusion, Low blood counts, Nausea and vomiting, Poor appetite, Elevation of blood liver enzymes	Approved, Investigational	321,322
Daunorubicin	Blood	Pain along the site where the medication was given, Urine may appear red, red-brown, orange or pink from the color of the medication for one to two days after you receive a dose, Low blood counts, Nausea or vomiting, Hair loss on the scalp or elsewhere on the body	Approved	323,324
Decitabine	Blood	Low blood counts, Fatigue, Fever, Nausea, Cough, Petechiae, Diarrhea, Constipation, Hyperglycemia	Approved, Investigational	325

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Table 2 (continued)

Drugs/Compound	Affected organ	Reported toxicity/side effects (frequency in >30% cases)	Current status	Reference/Sources
Doxorubicin	Blood, breast, ovarian, bladder, stomach, bronchus	Pain along the site where the medication was given, Nausea or vomiting, Low blood counts, Hair loss on the scalp or elsewhere on the body	Approved, Investigational	326,327
Epirubicin	Breast	Low blood counts, Mouth sores, Hair loss on the scalp or elsewhere on the body, Nausea and vomiting, Fatigue, Amenorrhea	Approved	321,328
Etoposide	Testicle, lung, blood, brain, ovarian	Low white blood cell count, Low platelet, Hair loss, Menopause, Loss of fertility, Nausea and vomiting	Approved	122
Idarubicin	Blood	Pain along the site where the medication was given, Low blood counts, Urine may appear red, red-brown, orange or pink from the color of the medication for one to two days after you receive a dose, Nausea or vomiting, Mouth sores (in the first week after therapy), Hair loss on the scalp or elsewhere on the body, Diarrhea/abdominal cramps	Approved	329
Ifosfamide	Testicle, blood, bladder, bone, pancreas, cervix, breast, stomach, breast, endometrium, lung, ovary	Low white blood cell count, Low platelet count, Hair loss, Nausea and vomiting, Poor appetite, Blood in the urine	Approved	330
Irinotecan	Colon, lung	Diarrhea; two types early and late forms, Early diarrhea accompanied by symptoms runny nose, increased salivation, watery eyes, sweating, Fever, flushing, abdominal cramping, Late diarrhea, Nausea and vomiting, Weakness, Low white blood cell count, Low red blood cell count, Hair loss, Poor appetite, Weight loss	Approved, Investigational	331
Lomustine	Brain, blood	Low blood counts (bone marrow suppression), Nausea and vomiting	Approved, Investigational	332
Mechlorethamine or Chlormethine	Blood, lung, brain	Low blood counts, Nausea and vomiting, Hair loss, Mouth sores, Redness, dryness, irritation with topical use, Loss of fertility	Approved, Investigational	15
Melphalan	Blood, ovary, breast, skin	Low blood counts, Nausea and vomiting	Approved	333

Mitomycin C	Stomach, intestine, breast, lung, bladder, pancreas, crevice	Low blood counts, Mouth sores, Poor appetite, Fatigue	Approved	137
Mitoxantrone	Prostate, blood, breast	Low blood counts, Nausea and vomiting, Fever, increases in blood tests measuring liver function	Approved, Investigational	334
Oxaliplatin	Intestine, pancreas, stomach	Peripheral neuropathy, Nausea and vomiting, Diarrhea, Mouth sores, Low blood counts, Fatigue, Loss of appetite	Approved, Investigational	335,336
Temozolomide	Brain, skin	Nausea and vomiting, Constipation, Headache, Fatigue	Approved, Investigational	337
Topotecan	Ovary, lung, cervix	Low blood counts, Nausea and vomiting, Hair loss, Diarrhea	Approved, Investigational	338–340
Thiotepa	Breast, ovary, thyroid, bladder	Low blood counts, Hair loss	Approved, Investigational	341
Trabectedin	Ovary	Anemia, Increased liver enzymes, Low white blood cell count, Nausea, Low platelets, Vomiting, Fatigue, Low potassium, Low phosphorous, Decreased appetite, Constipation, Increased CPK	Approved, Investigational	342,290
Purine antimetabolites (purine antagonists)				
Azathioprine	Blood	Black, tarry stools, Bleeding gums, Blood in the urine or stools, Chest pain, Chills, Cough, Fever, Hoarseness, Lower back or side pain, Painful or difficult urination, Pinpoint red spots on the skin, Sore throat, Sores, ulcers, or white spots on the lips or in the mouth, Swollen glands, unusual bleeding or bruising, unusual tiredness or weakness	Approved	291
Cladribine	Blood	Fever, Fatigue, Low white blood cell count, Low red blood cell count, Neutropenic fever, Myelosuppression	Approved, Investigational	343
Clofarabine	Blood	Leukopenia, Anemia, Infection, Limb pain, Lymphopenia, Low platelet count, Elevated liver enzymes, Vomiting, Nausea, Itching, Neutropenia, Diarrhea, Neutropenic fever, Creatinine increased, Bilirubin increased, Chills, Headache, Fever, Rash, High heart rate, Abdominal pain, Fatigue, Decreased appetite	Approved, Investigational	293,344
Fludarabine	Blood	Low blood counts, Fever, Infection, Weakness, Cough, Nausea and vomiting, Poor appetite	Approved	294
Gemcitabine	Breast, ovary, lung, pancreas,	Flu-like symptoms (muscle pain, fever,	Approved, Investigational	295

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Table 2 (continued)

Drugs/Compound	Affected organ	Reported toxicity/side effects (frequency in >30% cases)	Current status	Reference/Sources
	bladder	headache, chills, fatigue), Fever (within 6–12 h of first dose), Fatigue, Nausea (mild), Vomiting, Poor appetite, Skin rash, Low blood counts, Temporary increases in liver enzymes, Blood or protein in the urine		
Mercaptopurine	Blood, intestine	Low blood counts, Liver toxicity, Increased bilirubin, Increased liver enzymes, jaundice, abdominal swelling (ascites)	Approved	345
Nelarabine	Blood	Low blood counts, also known as bone marrow suppression (anemia, Fatigue, neutropenia, thrombocytopenia), Nausea	Approved, Investigational	297
Pentostatin	Blood	Cough or hoarseness, Fever or chills, Lower back or side pain, Painful or difficult urination, Skin rash or itching (sudden), Unusual tiredness or weakness	Approved, Investigational	346
Thioguanine	Blood	Low blood counts	Approved	347
Pyrimidine antimetabolites (pyrimidine antagonists)				
Capecitabine	Breast, stomach, intestine	Low white blood cell count, Low red blood cell count, Hand-foot syndrome, Diarrhea, Elevated liver enzymes, Fatigue, Nausea and vomiting, Rash and itching, Abdominal pain	Approved, Investigational	99
Cytarabine	Blood	Headache, Low blood counts, nausea and vomiting, Mouth sores, increases in blood tests measuring liver function	Approved, Investigational	97
Gemcitabine	Breast, ovarian, lung, pancreas, bladder	Flu-like, Fever, Fatigue, Nausea, Vomiting, Poor appetite, Skin rash, Low blood counts, Temporary increases in liver enzymes, Blood or protein in the urine	Approved	348,349
Fluorouracil	Breast, colon, pancreas, skin, stomach, rectal	Diarrhea, Nausea and possible occasional vomiting, Mouth sores, Poor appetite, Watery eyes, sensitivity to light, Taste changes, metallic taste in mouth during infusion, Low blood counts, white and red blood cells and platelets may temporarily decrease, Skin reactions,	Approved	98

Raltitrexed	Colon, mesothelium	Hair thinning, Hand -foot syndrome Increased risk of infection, Breathlessness and looking pale, Tiredness and weakness during and after treatment, Diarrhoea or constipation, Feeling or being sick, Skin problems, Mouth sores and ulcers, Tummy (abdominal) pain, Liver changes, Loss of appetite and weight loss, Dehydration	Approved, Investigational	102
Tegafur	Stomach	Myelosuppression, Central neurotoxicity, Gastrointestinal toxicity	Approved, Investigational	350
Other antineoplastic agents (belong to one or multiple drug classes)				
Arsenic trioxide (Trisenox®)	Blood	Nausea and vomiting, Cough, Fatigue, Fever, Headache, Rapid heartbeat, Sore throat, Abdominal pain, Diarrhea, Blood test abnormalities (low potassium and magnesium), Shortness of breath, Blurred vision and eye irritation, elevated blood sugar level, swelling of the face, hands, feet or legs, Difficulty sleeping, Rash, Heart rhythm changes, Joint pain, Itching, Numbness or tingling of hands or feet	Approved	351,352
Hydroxyurea	Skin, blood, ovary, breast	Low blood counts	Approved	301
Paclitaxel (Taxol®)	Bone, lung, ovary, breast	Low blood counts, Hair loss, Arthralgias and myalgias, pain in the joints and muscles, Peripheral neuropathy, Nausea and vomiting, Diarrhea, Mouth sores, Hypersensitivity reaction	Approved, Vet Approved	353
Docetaxel (Taxotere®)	Breast, lung, prostate, stomach	Low white blood cell count, Low red blood cell count, Fluid retention with weight gain, swelling of the ankles or abdominal area, Peripheral neuropathy, Nausea, Diarrhea, Mouth sores, Hair loss, Fatigue and weakness, Infection, Nail changes	Approved, Investigational	296
Ixabepilone (Ixempra®)	Breast, skin, lung, blood, prostate, kidney	Peripheral neuropathy, Weakness, Muscle and joint pains, Hair loss, Nausea and Vomiting, Low white blood cell count	Approved, Investigational	354
Methotrexate	Blood, breast chromium	Dizziness, Drowsiness, Headache, Hair Loss, Swollen, Tender Gums Decreased Appetite, Reddened Eyes	Approved	355
Pemetrexed	Pleura, lung	Fatigue, Nausea	Approved, Investigational	305

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Table 2 (continued)

Drugs/Compound	Affected organ	Reported toxicity/side effects (frequency in >30% cases)	Current status	Reference/Sources
(Alimta®) Streptozocin or Streptozotocin	Pancreas	Nausea and vomiting, Nephrotoxicity	Approved, Investigational	356
Vinblastine	Breast, blood, testicle, bone	Low blood counts, Injection site reactions, Fatigue and weakness	Approved	307
Vincristine	Blood	Hair loss, neurotoxicity, alopecia, myelosuppression	Approved, Investigational	357
Vindesine or Eldisine	Blood	Myelosuppression, Neurotoxicity	Approved, Investigational	358
Vinorelbine	Lung, blood, ovary, breast	Low blood counts, Nausea or vomiting, Muscle weakness, Constipation	Approved, Investigational	359, 360

Camptothecin and its derivatives

Camptothecin is a long-known topoisomerase inhibitory compound of natural origin. It was initially extracted from the bark of the tree *Camptotheca acuminata*.^{123,124} Camptothecin is a cytotoxic alkaloid comprised of a pentacyclic ring structure containing a pyrrole (3,4- β) quinoline moiety, an S-configured alpha-hydroxy lactone form, and a carboxylate form.¹²⁵ In later years, many more effective derivatives of this topoisomerase inhibitor have been synthesized and are effectively being applied in cancer chemotherapy.¹²⁶ The most recognized drug of this class, topotecan, is used against the advanced cancers of the ovary and lung. It targets topoisomerase-I, an enzyme responsible for easing the torsional strain in DNA by inducing single-strand breaks during replication.¹²⁰ The inhibition of topoisomerase-I by the active compounds may lead to sudden inhibition of DNA replication and transcription, causing growth arrest (Table 1).¹²⁷ In the physiological conditions, topotecan remains in equilibrium with its carboxylate structure.¹²⁸ Its active lactone can intercalate between DNA bases in the topoisomerase-I cleavage complex.¹²⁹ The coupling site of topotecan in the cleavage complex interferes with the topoisomerase-I while reiterating the scratched DNA strand in the wake of releasing the strain. Such intercalation traps the topoisomerase-I in the cleavage complex bound to the DNA.¹³⁰ Another well-known drug of this class is irinotecan that has shown efficacy against late-stage colorectal carcinoma, aggressive growths in cancers of the lung, cervix, ovary, colon, etc.^{131,132}

Actinomycin D (dactinomycin)

A transcription inhibitor chromopeptide molecule actinomycin D (also called dactinomycin) binds to DNA while initiating transcription at initiation complex and inhibits RNA elongation steps.¹³³ It can intercalate, preferably between guanidine and cytosine, or bind the DNA strand's minor grooves to form a highly stable drug-DNA complex, preventing DNA unwinding and thus stalling the progress of the RNA chain.¹³⁴ Dactinomycin was initially obtained from *Streptomyces* sp., identified as an antibiotic agent, but later its anti-tumor potential was recognized, and thus it was inducted in the chemotherapy as an anticancer antibiotic to treat Ewing's sarcoma, Wilms' tumor, rhabdomyosarcoma, trophoblastic neoplasm, testicular and ovarian cancers, etc (Table 1).¹³⁵

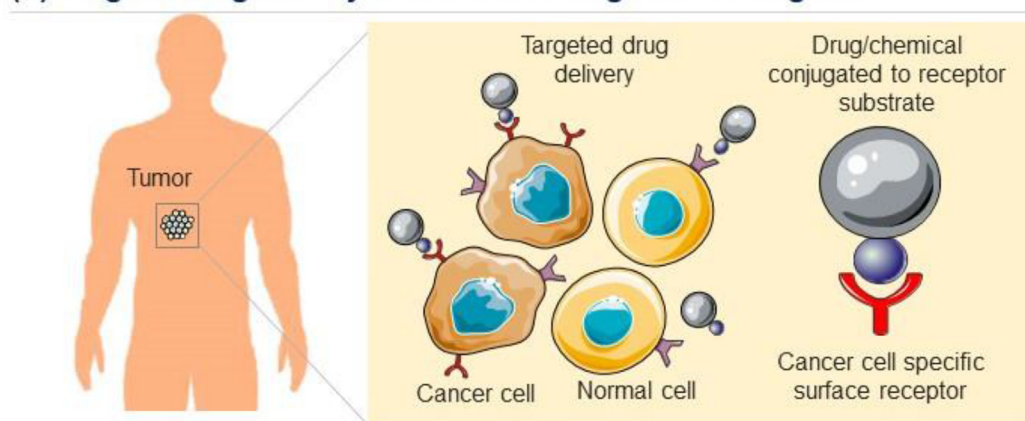
Mitomycin C

Mitomycins, a class of natural quinones, are also derived from *Streptomyces caespitosus* and are known for their transformation and competence, enhancing potential in bacterial cells. It is also a known antineoplastic agent as it slows down fibroblast cell growth in glaucoma.^{136,137} These bioreductive alkylating agents (especially mitomycin C derivatives) are also utilized as anti-cancer therapeutics because their structural modification impedes the cell cycle during G1 and S phase transitions. Its

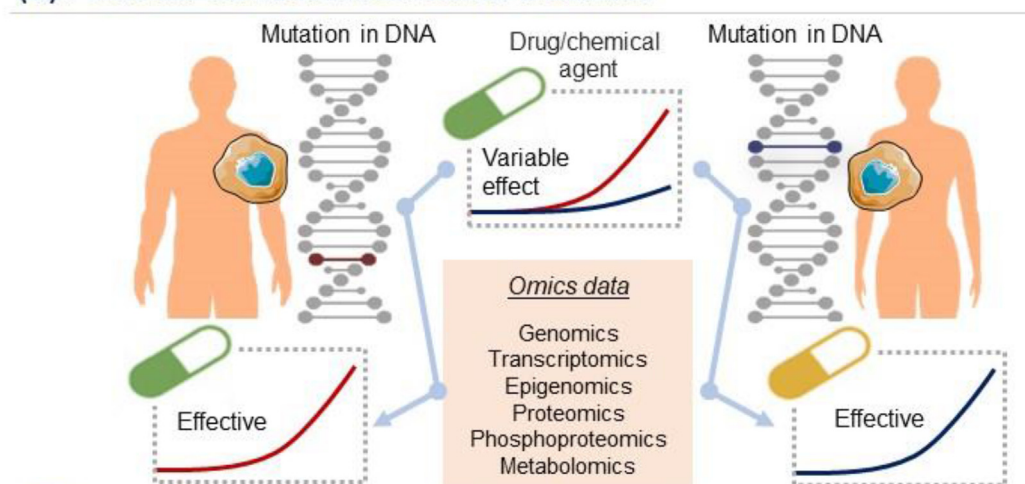
reduction to form mitosene can lead to the N-alkylation of DNA bases.¹³⁸ It can lead to inhibition of replication by cross-linking at adenine N6 and guanine O6 and N2.¹³⁹ It is effectively used in the palliative treatment of non-small cell lung carcinoma and other solid tumors of breast,

colorectal, oesophageal, pancreatic, and cervical origins. Porfiromycin is an N-methyl derivative of mitomycin C that is found effective against cancer of head and neck regions (Table 1).¹⁴⁰

(A) Targeted drug delivery of anti-cancer drugs/chemical agents



(B) Personalized medicine for cancer treatment



(C) Immunotherapy for cancer treatment

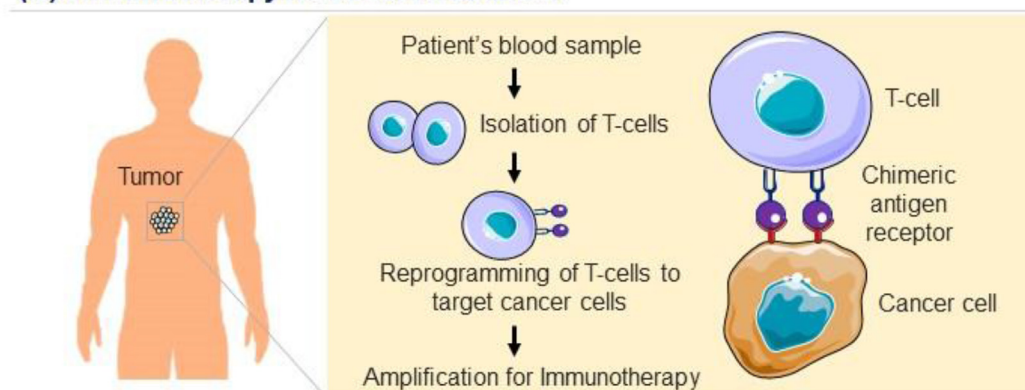


Figure 3 Schematic diagram showing targeted cancer therapeutic approaches. (A) Model showing targeted drug delivery approach of anti-cancer drugs/chemical agents conjugated with receptor substrate specific to the cancer cell receptor. (B) Model showing the building of a personalized medicine approach based on omics data and drug response study targeting specific mutation/signature in individual patients for precision cancer therapeutics. (C) Model showing immunotherapeutic regime involving reprogramming of T-cells to target cancer cells by a chimeric antigen receptor for their application in cancer immunotherapy.

Daunorubicin (rubidomycin)

Daunorubicin causes a break in DNA strands by actuating topoisomerase-II and producing quinone-type free radicals. Daunorubicin, otherwise called daunomycin, is a chemotherapy medicine used to treat cancer growth in acute myeloid leukemia (AML), acute lymphocytic leukemia (ALL), chronic myelogenous leukemia (CML), and Kaposi's sarcoma, etc.^{141,142} It is applied by infusion into a vein, and following its intercalation with DNA, it hinders macromolecular biosynthesis. This predominantly obstructs the movement of the enzyme topoisomerase-II, which loosens up the DNA supercoiling.¹⁴³ Daunorubicin impedes the topoisomerase-II complex after it unties the DNA chain for replication and thereby it represses the DNA double helix from resealing and therefore interferes with the replication to proceed (Table 1).^{144,145}

Bleomycin

Another *Streptomyces* compound with anti-tumor activity bleomycin is mostly used against Hodgkin's lymphoma, ovarian and testicular cancers. It chelates copper or iron, produces superoxide radicals, and intercalates between DNA strands — causes chain scission.¹⁴⁶ Emerging evidence proposes that bleomycin also hinders the consolidation of thymidine into DNA strands, and the cleavage of DNA relies upon oxygen and metal particles *in vitro*.¹⁴⁷ The specific mechanism of DNA strand scission is uncertain, yet it has been proposed that bleomycin chelates metal particles (principally iron), delivering a pseudoenzyme that responds with oxygen to create superoxide and hydroxide free radicals that divide DNA; however, little myelosuppression is the unique highlight.¹⁴⁸ The drug has also been effective against squamous cell carcinoma of the skin, and cancers of the mouth, head, neck, genitourinary tract, and throat (Table 1).

Lateral damage: Putting multiple organs at risk

Aggressive tumors are primarily marked with uncontrolled and extensive cell division that may compromise the normal physiology of other non-transformed cells in the vicinity of affected organs/tissues. In many ways, cancer cells achieve super-specialized metabolic and survival potential with the capacity to tolerate extreme stresses like low oxygen and nutrient supplies. Cancers might start from mutation of one type or the other, but as the disease progresses,^{149,150} multiple mutations and genetic abnormalities accumulate that lead to increased tumor heterogeneity.⁸⁷ The heterogeneous nature of tumor mass plays a vital role in developing resistance against most of the drugs that we have described here. This leads to a challenge of its kind, leaving doctors with the only option of changing the medication after certain intervals as one type of drug stops acting against tumors. In many cases, the combination therapy that includes more than one drug at a time could be an effective strategy against some form/type of cancers, such as cancers affecting bone marrow (leukemia) and lymph nodes (lymphoma). Clinical trials help a lot in understanding the effects of new combinations and thus have rapidly improved the cancer chemotherapy paradigm.

The sustainability of chemotherapy relies upon its continuous effect on cell division and tumor growth. As discussed above, the majority of the treatments work by compromising the stability or integrity of nucleic acids that tend to stall the cell cycle or activate programmed cell death pathways. Other drugs may act by generating extreme proteotoxic stress inside the cells by interfering with the protein folding or degradation machinery, including chaperone, proteasome, and autophagy.^{151–154} The idea is to generate cellular stresses so that the cancer cells that are already under compromised physiology may die while other cells can survive. However, it is evident from the above descriptions that most of these drugs act by interfering with one or multiple cellular pathways of extreme importance.¹⁵⁵ Therefore, chemotherapeutic

Advantages of cationic antimicrobial peptides as anti-tumor agents

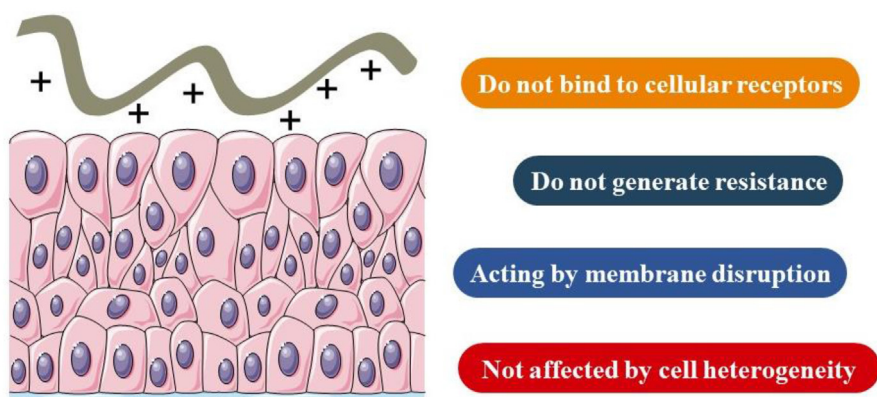


Figure 4 Advantages of antimicrobial peptides as anti-tumor agents.

drugs are mostly referred to as cytotoxic agents, causing a plethora of side effects. In most cases, long-term treatments may lead to enormous toxicity causing severe damage or failure of multiple organs. This may lead to enormous physiological and psychological stress to the patients under chemotherapeutic medication. Table 2 summarizes the adverse effects of various chemotherapeutic agents and the current stage of their clinical progress. The side effects of these drugs may depend on many factors and variables, such as type, stage, and location of the tumor, along with age, health, and other clinical aspects of the patients. The most common and readily visible side effects originate from the gut that may lead to nausea, vomiting, constipation, and diarrhea.^{156,157} Most of the anticancer drugs can directly interrupt the cell division pathways; therefore, the first line of affected organs may also include those which are formed with continuously dividing cells, such as skin, hair, and intestinal linings.^{158,148} Skin rashes, alopecia, and mucositis are the most visible side effects that one can observe in patients undergoing chemotherapy.^{157,159} Radiation also contributes significantly to these debilitating changes. A general problem observed in many patients is fatigue that can occur due to multiple factors, including depression, anemia, high doses of radiation therapy, etc.¹⁶⁰

Chemotherapeutic drugs may also directly injure the bone marrow and cause a steep decline in blood cell count, exposing patients to several other types of possible infections.¹⁶¹ Thrombocytopenia, a condition of excessive bleeding because of lower platelet numbers, is observed more frequently in patients taking alkylating and/or antimetabolites agents.¹⁶² Alkylating agents may further trigger conditions, such as hemorrhagic cystitis leading to microscopic haematuria, nocturia, dysuria, etc. along with mild or intense suprapubic pain.¹⁶³ Whereas, antimetabolites can generate enormous toxicity in vital organs, like the liver and kidneys.^{164,165} The most devastating of all is their direct impact on cardiovascular health. Different classes of chemotherapeutic drugs have varying impacts on cardiac cells, leading to multiple types of systemic conditions adversely affecting patients' survival.¹⁶⁶ Loss of libido and sexual dysfunctions are other conditions caused due to long-term exposure to these drugs, which affects not only physical well-being but also the patient's psychological health (Table 2).^{167,168}

These drugs and radiation therapies may have several direct or indirect impacts on cognitive functioning leading to a condition, which is now commonly referred to as chemo brain.^{169,170} This is a condition of the inability of someone undergoing chemo-drug therapies to do their routine cognitive functions, which may be related to their memory, cognition, and behavior.¹⁷¹ The drugs may adversely impact communication, reasoning, learning, problem-solving, memory storage, concentration, following commands, etc. Damage of optic and the ocular motor nerves is also observed in several patients taking antineoplastic drugs that may lead to the weakening of vision as well.^{172,173} The majority of chemotherapeutic agents affect these tissues in a dose-dependent manner; though, there are differences in their sensitivity to individuals due to enormous factors such as metabolism efficiency, genetic makeup, other medication, and environmental factors

(Table 2).¹⁷⁴ Despite enormous toxicities and side effects prevalent in the public domain, the chemotherapy medicines remained the most immediate and viable therapeutic option in most cases, where we can see the cancer cells but cannot remove them through surgery or source of radiation. To address various above-described detrimental physical and psychological changes, modern-day clinical practices involve a comprehensive team of doctors and specialists for the patients who are undergoing long-term treatments and are more prone to develop severe physiological complications.

Recent advancements and next-generation therapeutic approaches

Since the early 19th century, rigorous investigations and clinical studies were carried out around the world. Several research groups have made landmark discoveries during and after the Second World War. Following the development of a fundamental description of the disease and associated pathways, the focus shifted majorly towards advancing the diagnosis and treatment methods of the disease. Despite enormous progress being made in the second half of the last century, not much could have been achieved in successfully curing most of the cancers. One primary reason behind the failures is our inability to diagnose the oncogenic changes inside the body at the early stages of cancer development. In recent years, the concept of liquid biopsies has emerged that primarily relies on the screening of mutated DNAs, proteins, RNAs, and other genetic markers in the patient's blood samples.^{175–177} This practice can help diagnose the disease at a much earlier stage than we do now, using the traditional tissue biopsies and imaging methods.¹⁷⁸ New technological tools, like advanced software and robotic platforms, have also speeded cancer diagnosis and treatment. In this subsequent section taking chemotherapy into account, we comprehensively reviewed the recent progress made in the targeted drug delivery, personalized medication, and immunotherapy (Fig. 3).

Targeted drug delivery

The major drawback of most chemotherapeutic treatment plans is their non-specificity or inability to ascertain and target cancerous cells directly. Therefore, this lack of specificity poses a major therapeutic limit reflected in the failure of delivering drugs locally to the cancer tissue mass. In the past few decades, many new approaches have been constituted and tested, and have given hopes for future drug delivery (Fig. 3A). Nanoparticle-based delivery methods, targeted antibodies, aptamer functionalization, and specific drugs like Herceptin (in breast cancer) offered the promise that can harness great success in the upcoming years.^{179–181} Cancer cell-directed antibodies are at the forefront of targeted drug delivery methods and have accelerated the hunt for more effective or potentially improvised approaches.^{182,183} Different types of nano-carriers and formulations of nano-drugs have also improved the delivery of drug compounds to the cancer cells.^{184–186}

The most significant advantage of targeted drug deliveries is that the enormous drug-mediated toxicities on other adjacent tissues and organs can be minimized. We can also avoid most of the collateral damage leading to stress and organ failures. However, the effectiveness of many of these delivery methods in patients is still under clinical evaluation. Further details regarding these delivery systems can be accessed in the previously published reports.^{187,188}

Personalized medication

Given the heterogeneity of tumor cells, the cancer of one patient is different from that of another in terms of accumulated genetic abnormalities, its pace and progression, the response to the treatment, and the chances of developing resistance to the administered drugs.⁴⁵ The current technologies have made it possible to get insights into this disease heterogeneity or explore molecular variations at the individual level, thereby tailoring the treatment regime as per the personalized needs of every cancer patient.^{149,189} This helps in choosing the most appropriate drugs and their dosages for a better outcome in cancer management.¹⁹⁰ For instance, dabrafenib and vemurafenib are directed against *BRAF* gene mutation, while trastuzumab works well against HER-2 mutated cells.^{47,191,192} The idea here is to dive deep into the omics (genomic, transcriptomic, proteomic, metabolomic, etc.) data of the patient to identify the major molecular players fostering the transformed cells, and design the treatments against these clinically actionable targets (Fig. 3B).^{155,193,194} Different types of mutations have been observed to be frequent in specific subtypes of cancer. Therefore, measuring and manipulating these driver mutations based on an individual patient is expected to yield better results. Following are some examples of drugs that are potentially effective in patients harbouring some specific type of genomic alteration. A mutation in anaplastic lymphoma kinase (ALK) was reported to drive tumor formation in 5% of the non-small-cell lung cancer population and led to the discovery of ALK blockers such as crizotinib and ceritinib.¹⁹⁵ An oral *BRAF* inhibitor, namely vemurafenib has been shown to possess a greater affinity for the mutant *BRAF*^{V600E} than the wildtype *BRAF*, and therefore was recommended to the patients carrying the above mutation in the tumors.¹⁹⁶ Several potential targets for novel drugs have been identified using high-throughput screening technologies, and various compounds against them have been recently approved or are under investigation. Similarly, alterations in phosphoinositide 3-kinases (PI3K)/protein kinase B (AKT)/mTOR pathway have also been frequently found in many solid tumors. Among these, activation of phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha (PIK3CA) and loss of phosphatase and tensin homolog (PTEN) mutations are the most common. Basket trials have been conducted to evaluate the effectiveness of adding alpelisib, a specific p110 α PIK3CA inhibitor to the hormonal treatment for PIK3CA mutant luminal breast cancer patients. The results emphasize the importance of checking for this molecular alteration in patients and following the tested treatment regime for better anti-tumor response.¹⁹³ Next, aberrant changes in the fibroblast growth factors

(FGF) signaling pathway have also been associated with the initiation and progression of oncogenic events. Different types of genetic alterations in FGF receptor amplification, mutations, and gene-fusion have been shown to respond variably to certain drugs, and hence demand molecular profiling for the selection of effective drugs.^{46,197} Another promising example is of using poly ADP ribose polymerase (PARP) inhibitor olaparib for the treatment of breast cancer susceptibility gene (*BRCA*)-mutant cancers for better targeting of cancer cells, without much effect on the normal cells.^{198,199} On the cytoprotective side, recently a natural alkylated withanolide viz. 2,3-Dihydro-3beta-methoxy withaferin-A was shown to protect normal cells against a variety of stresses enrolled in cancer therapeutics.²⁰⁰ The idea of precision or personalized medicine is new but has been beneficial for lots of patients; however, the cost of these new generation approaches for genome profiling is still high, making it unaffordable for a large population of the world.²⁰¹

Antimicrobial peptides

Advances in cancer therapy have prompted scientists to look for natural tumoricidal chemicals, such as antimicrobial peptides (AMPs), that could be used for cancer treatment.^{202–204} In addition to antibacterial action, several AMPs have been shown to have intrinsic anticancer potential.^{205,206} These molecules constitute an intriguing new resource for anticancer drugs, and they provide several distinct therapeutic advantages over other anticancer treatments since they do not bind directly to any particular receptor in cancer cells.²⁰⁷ The polycationic and amphipathic character of AMPs has been postulated as a possible explanation for the preferential interaction of AMPs with cancer cells, as compared to normal cells.^{208,209} Membrane disruption has been identified as a critical stage in the microbial killing mechanism of several AMPs. These anticancer effects of AMPs are unaffected by molecular heterogeneity inside a tumor or across tumors, nor are they counteracted by cancer cells' flexibility and alternate survival pathways, as is commonly found in cancer resistance (Fig. 4).^{204,209,210}

Cathelicidins are members of the antimicrobial peptide family found in granules of vertebrate neutrophils.²¹¹ The only human cathelicidin is the cationic peptide LL-37. Cathelicidins are composed of a highly conserved N-terminal signal peptide including a cathelin domain and a C-terminal cationic antimicrobial peptide. Apart from their antimicrobial activity, cathelicidin-derived antimicrobial peptides exhibit a range of biological roles, including antiviral, antifungal, anticancer, and immunomodulatory properties.^{212,213} LL-37 exhibits anticancer activity against oral cancer cells and has been identified as a molecular target for colon cancer.²¹⁴ Immunohistochemical staining of colon cancer tissue showed that LL-37 is abundantly expressed in normal colon mucosa but downregulated in colon cancer tissues due to DNA methylation in the LL-37 promoter.²¹⁵

In human oral squamous cell carcinoma SAS-H1 cells, the active domain of LL-37 induces mitochondrial depolarization, which results in caspase-independent apoptosis.

Furthermore, mice lacking cathelicidin are more susceptible to azoxymethane-induced colon carcinogenesis.²¹⁶

Many important advancements have been achieved using antimicrobial peptides against colorectal cancer.²¹⁷ Using LL-37 on the surface of magnetic nanoparticles led to a significant reduction in cell viability of colon cancer cell lines, inducing apoptosis at a much higher rate than in the case of LL-37 peptide alone.²¹⁸

Fan et al created a biodegradable and injectable thermosensitive hydrogel containing docetaxel and LL-37 polymeric nanoparticles. Intraperitoneal injection of this hydrogel substantially inhibited the development of HCT116 carcinomatosis in a mouse model of colorectal carcinoma.²¹⁹

In comparison to existing chemotherapeutic agents, the flexible nature of peptide structures, along with non-receptor activity, might confer strategic benefits on AMPs for the treatment of colon cancer in the future.²²⁰ New treatment approaches for patients with advanced-stage, unresectable melanomas continue to be of significant clinical interest, as these patients cannot be treated surgically and have only a modest or temporary response to chemotherapy and radiation. In phase 1 clinical trial NCT02225366, intra-tumoral injections of LL-37 are being explored for melanoma patients with cutaneous metastases to determine the most effective dose of LL-37 that could be administered. Patients will receive weekly intra-tumoral injections of LL-37 (starting dose, 250 µg/tumor) every 7 days for up to 8 weeks. Researchers are also interested in understanding whether LL37 can activate the immune system, which could help in the control of the disease.²²¹

Although AMPs have the potential to develop into a more effective and safer alternative to conventional cancer therapy, future research and development of anticancer peptide therapeutics will most likely focus on the poor pharmacokinetics, the control of selectivity, toxicity, and routes of administration through the establishment of appropriate *in vivo* models.²²²

Immunotherapy

Supplementing our immune system by devising immunotherapy strategies is another highly-specific and largely promising approach to address the problem of late-stage cancer therapeutics.^{223,224} Several new approaches have been developed in the last decade to modify and train a patient's immune system to search, identify and target the transformed cells inside the body by recognizing multiple cell surface markers.²²⁵ For instance, supplementing the treatment regime with immunotherapy has significantly improved the success rate of the chemotherapeutic drugs in many cancer types, especially the triple-negative breast cancers.^{226–228} The immunotherapy drugs are given alone or in combination with chemotherapy drugs to achieve better clinical outcomes.²²⁹ Prophylactic and therapeutic vaccines have been developed to help patients' immune systems towards learning to tackle the transformed cells.²³⁰ Atezolizumab is an example of an anti-cancer immune drug that helps to dissolve hard-to-treat breast cancers.^{231–233} Tisagenlecleucel, an FDA-approved drug, treats B-cell lymphoblastic leukemia by effectively engineering

the chimeric T cell receptor (CAR-T) of the patient (Fig. 3C).²³⁴ Some treatment plans also include interferons and interleukins to jump-start the fight against cancer cells by boosting the patient's immune system. Moreover, the field of immunotherapy is gradually evolving and is beyond the scope of our present article.

Combination therapy

Combination therapy may offer the advantages of the fast tumor regression of targeted therapies with the long-lasting responses induced by immunotherapy.²³⁵ Immunotherapy may be able to consolidate the dramatic tumor responses achieved with targeted therapy into durable, long-lasting remissions, in which sustained host responses targeting multiple antigens may reduce the risk of developing potentially lethal drug-resistant clones.²³⁶ Combining cancer vaccines with immune checkpoint inhibitors (ICIs) such as those targeting cytotoxic T lymphocyte-associated protein 4 (CTLA-4) and programmed cell death protein 1/programmed cell death ligand 1 (PD-1/PD-L1) is one approach to avoid T cell anergy.^{237,238} Moreover, T-cell and NK-cell invasion may be facilitated by targeted drugs that modify the tumor endothelium.²³⁹ Gemcitabine, for example, is a deoxycytidine analog that induces tumor cell death, which results in antigen cross-presentation and cross-priming.^{240,241} Carboplatin, pemetrexed, and pembrolizumab are now commonly used in the treatment of non-small cell lung cancer as first-line agents.²⁴² For triple-negative breast cancer, nab-paclitaxel and atezolizumab (anti-PD-L1) were recently authorized based on an improvement in progression-free survival (PFS) over chemotherapy.²⁴³

Adenosine receptor antagonists (ARs) are involved in regulating tumor progression.²⁴⁴ The four ARs discovered so far, A₁, A_{2A}, A_{2B} and A₃, belong to the class A family of G protein-coupled receptors (GPCRs).²⁴⁵ The A_{2B}R seems more promising as a target in chemotherapeutic interventions,²⁴⁶ whereas the A_{2A}R is the main target in the immunotherapeutic approach.^{247,248} The existence of dual compounds, i.e., molecules acting on the A_{2A}R and on the A_{2B}R, has prompted their use in both immunotherapeutic and chemotherapeutic interventions.^{249,250}

Epigenetic modulators may have a dual function in the tumor microenvironment, boosting both antigen expression and T cell effector activity.^{251,252} ICI-induced reactivation of exhausted T cells is linked with considerable chromatin remodeling, indicating that epigenetic modification may relieve certain patients with exhausted T cell function.^{253,254}

While many strategies focus on modifying the native tumor microenvironment, an alternative strategy involves the direct infusion of engineered immune cells or cellular receptors into the patient via adoptive cellular therapy with the goal of increasing the number of effector cells that recognize tumor-expressed antigens.²⁵⁵ Upon reinfusion, lymphocytes grown *ex vivo* may respond more potently to tumor antigen since they have not been continuously exposed to tolerogenic microenvironmental signals.²⁵⁶ Tumor infiltrating lymphocytes (TIL) treatment,²⁵⁷ chimeric antigen receptor (CAR) engineered T-cells,^{258,259} and TCR-engineered cell therapy^{260,261} are the three main forms of

adoptive T cell therapy currently being investigated.^{257,262,263}

Injecting oncolytic viruses directly into the tumor microenvironment is an alternate technique for improving tumor antigen recognition and strengthening T cell responses.²⁶⁴ Talimogene laherparepvec (T-VEC) is a modified herpes simplex virus that is injected intra-lesionally into the melanoma tumor in individuals who have failed to get their tumor removed. It induces immediate lysis of tumor cells and the production of granulocyte-macrophage colony-stimulating factor (GM-CSF), which serves as a cytokine to stimulate the recruitment, maturation, and activity of antigen presenting cells (APCs).²⁶⁵

Several studies combining immunotherapy with molecularly targeted treatment, either concurrently or sequentially, have shown hepatotoxicity as a major concern.²⁶⁶ Combinations of dabrafenib, trametinib, and anti-PD-1 treatment have been associated with a greater risk of grade 3/4 adverse events in melanoma than would be expected with targeted therapy alone.²⁵⁶

Emerging drugs are beginning to take a more comprehensive perspective of the tumor microenvironment (TME) and are revealing complementary techniques to simultaneously restore and enhance immune activity and promote therapeutic synergy.^{267,268} Future research of combined therapy should continue to concentrate on exploring the intricate interplay between targeted drugs and immunotherapy, as well as optimizing parameters such as administration time, dose, and sequencing that may enhance the therapeutic index.²⁴⁰

Conclusions and future perspectives

Chemotherapy is a largely investigated and clinically assessed approach in cancer therapeutics. In this report, we summarized the present status of chemotherapy and several drugs/agents presently enrolled in the clinical practice to provide an overview of how the field has evolved over the years and has traced the possibilities and druggability of diverse cancers in the clinic. New trends are evolving in the drug discovery paradigm that includes designing new derivatives of old drugs with high efficacy and lower toxicities.^{269,270} Several natural compounds isolated from microbial, plants, or natural sources were suggested to hold enormous potential to target stress response pathways by mounting a hyperactive response instigating cancer cell death.^{271–274} Natural compounds/agents as chemotherapeutic drugs demonstrated several advantages over synthetic ones, owing to their less cytotoxicity, cost, and straightforward production/extraction process.^{273,275,276} Therefore, recognition and attenuation of treatment toxicity in cancer chemotherapy will be a key factor in determining the success of its regimes in the future.

Revised clinical strategies further underlined that combination therapy could emerge as a promising therapeutic approach to treat diverse cancers in the future. Combination strategies comprise an obvious overlap of the chemotherapeutic regimes with the targeted drug delivery, personalized medicine, and immunotherapy. It may not only improve the efficacy of enrolled treatment but can

also elicit the responsiveness of certain tumors towards their effective therapeutic targeting in the clinic. Among these, personalized combination therapies that target individual tumor types based on their molecular signatures may offer a great promise. With acquiring more personalized insights in the chemotherapeutic treatments, a greater emphasis will be on individual regimes that could meet a high clinical success.²⁷⁷ Therefore, chemotherapy beyond its conventional clinical efficacy offers to improve the clinical outcome in combination therapies. Translation of these combined approaches in the clinics is yet to see a complete success and thus warrants more clinical investigations.

Conclusively, clinical practice of cancer chemotherapy has achieved considerable success, however, it still holds scope to improve, particularly in terms of efficacy and safety of enrolled chemotherapeutic regimes. Therefore, it further requires concerted efforts and careful investigations assessing the efficacy of combined therapeutic regimes. In summary, more investigative focus on drug design in individual or combined regimes and rigor on their clinical trials involving careful patient selection and stratification are warranted to achieve the greater therapeutic success of chemotherapeutic regimens in the clinic.

Conflict of interests

The authors declare no conflict of interests.

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References

1. Janssen LME, Ramsay EE, Logsdon CD, Overwijk WW. The immune system in cancer metastasis: friend or foe? *J Immunother Cancer*. 2017;5(1):79.
2. Laplagne C, Domagala M, le Naour A, et al. Latest advances in targeting the tumor microenvironment for tumor suppression. *Int J Mol Sci*. 2019;20(19):4719.
3. Emon B, Bauer J, Jain Y, Jung B, Saif T. Biophysics of tumor microenvironment and cancer metastasis - a mini review. *Comput Struct Biotechnol J*. 2018;16:279–287.
4. Costa A, Scholer-Dahirel A, Mechta-Grigoriou F. The role of reactive oxygen species and metabolism on cancer cells and their microenvironment. *Semin Cancer Biol*. 2014;25:23–32.

5. Gilbert S, Oliphant C, Hassan S, et al. ATP in the tumour microenvironment drives expression of nfP2X7, a key mediator of cancer cell survival. *Oncogene*. 2019;38(2):194–208.
6. Pfirschke C, Siwicki M, Liao HW, Pittet MJ. Tumor microenvironment: no effector T cells without dendritic cells. *Cancer Cell*. 2017;31(5):614–615.
7. Bedoui S, Herold MJ, Strasser A. Emerging connectivity of programmed cell death pathways and its physiological implications. *Nat Rev Mol Cell Biol*. 2020;21(11):678–695.
8. Rodrigues D, Souza T, Jennen DGJ, Lemmens L, Kleinjans JCS, de Kok TM. Drug-induced gene expression profile changes in relation to intestinal toxicity: state-of-the-art and new approaches. *Cancer Treat Rev*. 2019;77:57–66.
9. Delou JMA, Souza ASO, Souza LCM, Borges HL. Highlights in resistance mechanism pathways for combination therapy. *Cells*. 2019;8(9):1013.
10. Kalra RS, Bapat SA. Enhanced levels of double-strand DNA break repair proteins protect ovarian cancer cells against genotoxic stress-induced apoptosis. *J Ovarian Res*. 2013;6(1):66.
11. Caccuri F, Sommariva M, Marsico S, et al. Inhibition of DNA repair mechanisms and induction of apoptosis in triple negative breast cancer cells expressing the human herpesvirus 6 U94. *Cancers*. 2019;11(7):1006.
12. Kalra RS, Bapat SA. Expression proteomics predicts loss of RXR- γ during progression of epithelial ovarian cancer. *PLoS One*. 2013;8(8):e70398.
13. Huang C, Vaishnavi K, Kalra R. 3 β -methoxy derivation of withaferin-a attenuates its anticancer potency: bioinformatics and molecular evidences. *Med Aromatic Plants*. 2015;4(219):2167-0412.
14. Chaudhary A, Kalra RS, Malik V, et al. 2, 3-dihydro-3 β -methoxy withaferin-A lacks anti-metastasis potency: bioinformatics and experimental evidences. *Sci Rep*. 2019;9(1):17344.
15. Chen Y, Jia Y, Song W, Zhang L. Therapeutic potential of nitrogen mustard based hybrid molecules. *Front Pharmacol*. 2018;9:1453.
16. Needham DM, Cohen JA, Barrett AM. The mechanism of damage to the bone marrow in systemic poisoning with mustard gas. *Biochem J*. 1947;41(4):631–639.
17. Ghanei M. Delayed haematological complications of mustard gas. *J Appl Toxicol*. 2004;24(6):493–495.
18. Cairns J. The treatment of diseases and the war against cancer. *Sci Am*. 1985;253(5):51–59.
19. Duffy MJ. The war on cancer: are we winning? *Tumour Biol*. 2013;34(3):1275–1284.
20. Christakis P. The birth of chemotherapy at Yale. Bicentennial lecture series: surgery Grand Round. *Yale J Biol Med*. 2011;84(2):169–172.
21. DeVita Jr VT, Chu E. A history of cancer chemotherapy. *Cancer Res*. 2008;68(21):8643–8653.
22. Sawyers C. Targeted cancer therapy. *Nature*. 2004;432(7015):294–297.
23. Dewanjee S, Vallamkondu J, Kalra RS, et al. The emerging role of HDACs: pathology and therapeutic targets in diabetes mellitus. *Cells*. 2021;10(6):1340.
24. Mattheolabakis G, Rigas B, Constantinides PP. Nanodelivery strategies in cancer chemotherapy: biological rationale and pharmaceutical perspectives. *Nanomedicine*. 2012;7(10):1577–1590.
25. Leu JIJ, Pimkina J, Pandey P, Murphy ME, George DL. HSP70 inhibition by the small-molecule 2-phenylethanesulfonamide impairs protein clearance pathways in tumor cells. *Mol Cancer Res*. 2011;9(7):936–947.
26. Wu WK, Wu YC, Yu L, Li ZJ, Sung JJ, Cho CH. Induction of autophagy by proteasome inhibitor is associated with proliferative arrest in colon cancer cells. *Biochem Biophys Res Commun*. 2008;374(2):258–263.
27. Selimovic D, Porzig BBOW, El-Khattouti A, et al. Bortezomib/proteasome inhibitor triggers both apoptosis and autophagy-dependent pathways in melanoma cells. *Cell Signal*. 2013;25(1):308–318.
28. Hasima N, Ozpolat B. Regulation of autophagy by polyphenolic compounds as a potential therapeutic strategy for cancer. *Cell Death Dis*. 2014;5:e1509.
29. Sannino S, Guerriero CJ, Sabnis AJ, et al. Compensatory increases of select proteostasis networks after Hsp70 inhibition in cancer cells. *J Cell Sci*. 2018;131(17):jcs217760.
30. Zhu K, Dunner Jr K, McConkey DJ. Proteasome inhibitors activate autophagy as a cytoprotective response in human prostate cancer cells. *Oncogene*. 2010;29(3):451–462.
31. Chen J, Chen X, Xu D, et al. Autophagy induced by proteasomal DUB inhibitor NipT restricts NipT-mediated cancer cell death. *Front Oncol*. 2020;10:348.
32. Stewart DA, Winnike JH, McRitchie SL, Clark RF, Pathmasiri WW, Sumner SJ. Metabolomics analysis of hormone-responsive and triple-negative breast cancer cell responses to paclitaxel identify key metabolic differences. *J Proteome Res*. 2016;15(9):3225–3240.
33. Sun H, Zhang AH, Liu SB, et al. Cell metabolomics identify regulatory pathways and targets of magnoline against prostate cancer. *J Chromatogr B Analyt Technol Biomed Life Sci*. 2018;1102–1103:143–151.
34. Kroemer G, Zitvogel L. Immune checkpoint inhibitors. *J Exp Med*. 2021;218(3):e20201979.
35. Torino F, Barnabei A, Paragliola R, Baldelli R, Appetecchia M, Corsello SM. Thyroid dysfunction as an unintended side effect of anticancer drugs. *Thyroid*. 2013;23(11):1345–1366.
36. Mahner S, Eulenburger C, Staehle A, et al. Prognostic impact of the time interval between surgery and chemotherapy in advanced ovarian cancer: analysis of prospective randomised phase III trials. *Eur J Cancer*. 2013;49(1):142–149.
37. di Leonardo A, Nasi S, Pulciani S. Cancer: we should not forget the past. *J Cancer*. 2015;6(1):29–39.
38. Papac RJ. Origins of cancer therapy. *Yale J Biol Med*. 2001;74(6):391–398.
39. Morrison WB. Cancer chemotherapy: an annotated history. *J Vet Intern Med*. 2010;24(6):1249–1262.
40. Riddle JM. Ancient and medieval chemotherapy for cancer. *Isis*. 1985;76(3):319–330.
41. Lee CT, Huang YW, Yang CH, Huang KS. Drug delivery systems and combination therapy by using Vinca alkaloids. *Curr Top Med Chem*. 2015;15(15):1491–1500.
42. Lau YC, Lip GYH. New advances in the treatment of atrial fibrillation: focus on stroke prevention. *Expert Opin Pharmacother*. 2014;15(15):2193–2204.
43. DeVita VT, Hellman S, Rosenberg SA. *Cancer, Principles & Practice of Oncology*. 11th ed. Philadelphia, PA: Lippincott Williams & Wilkins; 2018.
44. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144(5):646–674.
45. Caon I, Bartolini B, Parnigoni A, et al. Revisiting the hallmarks of cancer: the role of hyaluronan. *Semin Cancer Biol*. 2020;62:9–19.
46. Dieci MV, Arnedos M, Andre F, Soria JC. Fibroblast growth factor receptor inhibitors as a cancer treatment: from a biologic rationale to medical perspectives. *Cancer Discov*. 2013;3(3):264–279.
47. Holderfield M, Deuker MM, McCormick F, McMahon M. Targeting RAF kinases for cancer therapy: BRAF-mutated melanoma and beyond. *Nat Rev Cancer*. 2014;14(7):455–467.

48. Kalra RS, Chaudhary A, Yoon AR, et al. CARF enrichment promotes epithelial-mesenchymal transition via Wnt/ β -catenin signaling: its clinical relevance and potential as a therapeutic target. *Oncogenesis*. 2018;7(5):39.
49. Chua TC, Yan TD, Saxena A, Morris DL. Should the treatment of peritoneal carcinomatosis by cytoreductive surgery and hyperthermic intraperitoneal chemotherapy still be regarded as a highly morbid procedure? : a systematic review of morbidity and mortality. *Ann Surg*. 2009;249(6):900–907.
50. Jin JF, Zhu LL, Chen M, et al. The optimal choice of medication administration route regarding intravenous, intramuscular, and subcutaneous injection. *Patient Prefer Adherence*. 2015;9:923–942.
51. Giger-Pabst U, Bucur P, Roger S, et al. Comparison of tissue and blood concentrations of oxaliplatin administered by different modalities of intraperitoneal chemotherapy. *Ann Surg Oncol*. 2019;26(13):4445–4451.
52. Lichota A, Gwozdziński K. Anticancer activity of natural compounds from plant and marine environment. *Int J Mol Sci*. 2018;19(11):3533.
53. Weaver BA. How Taxol/paclitaxel kills cancer cells. *Mol Biol Cell*. 2014;25(18):2677–2681.
54. Omar A, Kalra RS, Putri J, Elwakeel A, Kaul SC, Wadhwa R. Soyasapogenol-A targets CARF and results in suppression of tumor growth and metastasis in p53 compromised cancer cells. *Sci Rep*. 2020;10(1):6323.
55. Sun G, Rong D, Li Z, et al. Role of small molecule targeted compounds in cancer: progress, opportunities, and challenges. *Front Cell Dev Biol*. 2021;9:694363.
56. Apetoh L, Ladoire S, Coukos S, Ghiringhelli F. Combining immunotherapy and anticancer agents: the right path to achieve cancer cure? *Ann Oncol*. 2015;26(9):1813–1823.
57. Piktel E, Niemirówicz K, Wątek M, Wollny T, Deptuła P, Bucki R. Recent insights in nanotechnology-based drugs and formulations designed for effective anti-cancer therapy. *J Nanobiotechnol*. 2016;14(1):39.
58. Anand U, Carpena M, Kowalska-Górska M, et al. Safer plant-based nanoparticles for combating antibiotic resistance in bacteria: a comprehensive review on its potential applications, recent advances, and future perspective. *Sci Total Environ*. 2022:153472.
59. Kaur K, Reddy S, Barathe P, et al. Combating drug-resistant bacteria using photothermally active nanomaterials: a perspective review. *Front Microbiol*. 2021;12:747019.
60. Mitra AK, Agrahari V, Mandal A, et al. Novel delivery approaches for cancer therapeutics. *J Contr Release*. 2015;219:248–268.
61. Large DE, Soucy JR, Hebert J, Auguste DT. Advances in receptor-mediated, tumor-targeted drug delivery. *Adv Ther*. 2019;2(1):1800091.
62. Yarchoan M, Agarwal P, Villanueva A, et al. Recent developments and therapeutic strategies against hepatocellular carcinoma. *Cancer Res*. 2019;79(17):4326–4330.
63. Vaidyanathan R, Soon RH, Zhang P, Jiang K, Lim CT. Cancer diagnosis: from tumor to liquid biopsy and beyond. *Lab Chip*. 2018;19(1):11–34.
64. Bera K, Schalper KA, Rimm DL, Velcheti V, Madabhushi A. Artificial intelligence in digital pathology - new tools for diagnosis and precision oncology. *Nat Rev Clin Oncol*. 2019;16(11):703–715.
65. Charmsaz S, Prencipe M, Kiely M, Pidgeon GP, Collins DM. Innovative technologies changing cancer treatment. *Cancers*. 2018;10(6):208.
66. Markham MJ, Wachter K, Agarwal N, et al. Clinical cancer advances 2020: annual report on progress against cancer from the American society of clinical oncology. *J Clin Oncol*. 2020;38(10):1081.
67. Jaffee EM, van Dang C, Agus DB, et al. Future cancer research priorities in the USA: a lancet oncology commission. *Lancet Oncol*. 2017;18(11):e653–e706.
68. Zhao J, Dean DC, Hornicek FJ, Yu X, Duan Z. Emerging next-generation sequencing-based discoveries for targeted osteosarcoma therapy. *Cancer Lett*. 2020;474:158–167.
69. Coccia M. Deep learning technology for improving cancer care in society: new directions in cancer imaging driven by artificial intelligence. *Technol Soc*. 2020;60:101198.
70. Patel S. Emerging adjuvant therapy for cancer: propolis and its constituents. *J Diet Suppl*. 2016;13(3):245–268.
71. Grob JJ, Garbe C, Ascierto P, Larkin J, Dummer R, Schadendorf D. Adjuvant melanoma therapy with new drugs: should physicians continue to focus on metastatic disease or use it earlier in primary melanoma? *Lancet Oncol*. 2018;19(12):e720–e725.
72. Benson 3rd AB, Schrag D, Somerfield MR, et al. American Society of Clinical Oncology recommendations on adjuvant chemotherapy for stage II colon cancer. *J Clin Oncol*. 2004;22(16):3408–3419.
73. Matei D, Filiaci V, Randall ME, et al. Adjuvant chemotherapy plus radiation for locally advanced endometrial cancer. *N Engl J Med*. 2019;380(24):2317–2326.
74. Cao L, Xu C, Cai G, et al. How does the interval between completion of adjuvant chemotherapy and initiation of radiotherapy impact clinical outcomes in operable breast cancer patients? *Ann Surg Oncol*. 2021;28(4):2155–2168.
75. Salazar MC, Rosen JE, Wang Z, et al. Association of delayed adjuvant chemotherapy with survival after lung cancer surgery. *JAMA Oncol*. 2017;3(5):610–619.
76. Kalyanaraman B. Teaching the basics of cancer metabolism: developing antitumor strategies by exploiting the differences between normal and cancer cell metabolism. *Redox Biol*. 2017;12:833–842.
77. Moore A. Metabolic cycles in cancer cells? *Bioessays*. 2020;42(4):e2000048.
78. Anand U, Jacobo-Herrera N, Altemimi A, Lakhssassi N. A comprehensive review on medicinal plants as antimicrobial therapeutics: potential avenues of biocompatible drug discovery. *Metabolites*. 2019;9(11):258.
79. Alibek K, Bekmurzayeva A, Mussabekova A, Sultankulov B. Using antimicrobial adjuvant therapy in cancer treatment: a review. *Infect Agents Cancer*. 2012;7(1):33.
80. Shoshan-Barmatz V, Anand U, Nahon-Crystal E, di Carlo M, Shteinifer-Kuzmine A. Adverse effects of metformin from diabetes to COVID-19, cancer, neurodegenerative diseases, and aging: is VDAC1 a common target? *Front Physiol*. 2021;12:730048.
81. Nicholson LB. The immune system. *Essays Biochem*. 2016;60(3):275–301.
82. Gonzalez H, Hagerling C, Werb Z. Roles of the immune system in cancer: from tumor initiation to metastatic progression. *Genes Dev*. 2018;32(19–20):1267–1284.
83. Whiteside TL. Immune responses to cancer: are they potential biomarkers of prognosis? *Front Oncol*. 2013;3:107.
84. Kreuzaler P, Panina Y, Segal J, Yuneva M. Adapt and conquer: metabolic flexibility in cancer growth, invasion and evasion. *Mol Metabol*. 2020;33:83–101.
85. Yoshida GJ. Metabolic reprogramming: the emerging concept and associated therapeutic strategies. *J Exp Clin Cancer Res*. 2015;34:111.
86. Wadhwa R, Kalra RS, Kaul SC. CARF is a multi-module regulator of cell proliferation and a molecular bridge between cellular senescence and carcinogenesis. *Mech Ageing Dev*. 2017;166:64–68.
87. Mroz EA, Rocco JW. The challenges of tumor genetic diversity. *Cancer*. 2017;123(6):917–927.
88. Gambara G, Gaebler M, Keilholz U, Regenbrecht CRA, Silvestri A. From chemotherapy to combined targeted

- therapeutics: *in vitro* and *in vivo* models to decipher intra-tumor heterogeneity. *Front Pharmacol.* 2018;9:77.
89. Palmer AC, Sorger PK. Combination cancer therapy can confer benefit via patient-to-patient variability without drug additivity or synergy. *Cell.* 2017;171(7):1678–1691.e13.
 90. Ke X, Shen L. Molecular targeted therapy of cancer: the progress and future prospect. *Front Lab Med.* 2017;1(2):69–75.
 91. Kondo N, Takahashi A, Ono K, Ohnishi T. DNA damage induced by alkylating agents and repair pathways. *J Nucleic Acids.* 2010;2010:543531.
 92. Qiu H, Wang Y. Exploring DNA-binding proteins with *in vivo* chemical cross-linking and mass spectrometry. *J Proteome Res.* 2009;8(4):1983–1991.
 93. Mehrmohamadi M, Jeong SH, Locasale JW. Molecular features that predict the response to antimetabolite chemotherapies. *Cancer Metabol.* 2017;5:8.
 94. Katzung BG. *Basic and Clinical Pharmacology*. 10th ed. Singapore: McGraw-Hill; 2008.
 95. Lambie DG, Johnson RH. Drugs and folate metabolism. *Drugs.* 1985;30(2):145–155.
 96. Raimondi MV, Randazzo O, la Franca M, et al. DHFR inhibitors: reading the past for discovering novel anticancer agents. *Molecules.* 2019;24(6):1140.
 97. Löwenberg B, Pabst T, Vellenga E, et al. Cytarabine dose for acute myeloid leukemia. *N Engl J Med.* 2011;364(11):1027–1036.
 98. Grem JL. 5-Fluorouracil: forty-plus and still ticking. A review of its preclinical and clinical development. *Invest N Drugs.* 2000;18(4):299–313.
 99. Masuda N, Lee SJ, Ohtani S, et al. Adjuvant capecitabine for breast cancer after preoperative chemotherapy. *N Engl J Med.* 2017;376(22):2147–2159.
 100. Dyawanapelly S, Kumar A, Chourasia MK. Lessons learned from gemcitabine: impact of therapeutic carrier systems and gemcitabine's drug conjugates on cancer therapy. *Crit Rev Ther Drug Carrier Syst.* 2017;34(1):63–96.
 101. Malik P, Cashen AF. Decitabine in the treatment of acute myeloid leukemia in elderly patients. *Cancer Manag Res.* 2014;6:53–61.
 102. Liu Y, Wu W, Hong W, Sun X, Wu J, Huang Q. Raltitrexed-based chemotherapy for advanced colorectal cancer. *Clin Res Hepatol Gastroenterol.* 2014;38(2):219–225.
 103. Hasegawa K, Saiura A, Takayama T, et al. Oral adjuvant chemotherapy using uracil-tegafur (UFT) with leucovorin (LV) after resection of colorectal cancer liver metastases: long-term survival results of the phase III UFT/LV study. *J Clin Oncol.* 2017;35:672.
 104. Moudi M, Go R, Yien CY, Nazre M. Vinca alkaloids. *Int J Prev Med.* 2013;4(11):1231–1235.
 105. Bates D, Eastman A. Microtubule destabilising agents: far more than just antimetabolic anticancer drugs. *Br J Clin Pharmacol.* 2017;83(2):255–268.
 106. Habli Z, Toumeh G, Fatfat M, Rahal ON, Gali-Muhtasib H. Emerging cytotoxic alkaloids in the battle against cancer: overview of molecular mechanisms. *Molecules.* 2017;22(2):250.
 107. Falzone L, Salomone S, Libra M. Evolution of cancer pharmacological treatments at the turn of the third millennium. *Front Pharmacol.* 2018;9:1300.
 108. Schneider F, Pan L, Ottenbruch M, List T, Gaich T. The chemistry of nonclassical taxane diterpene. *Acc Chem Res.* 2021;54(10):2347–2360.
 109. Fanale D, Bronte G, Passiglia F, et al. Stabilizing versus destabilizing the microtubules: a double-edge sword for an effective cancer treatment option? *Anal Cell Pathol.* 2015;2015:690916.
 110. Leung JC, Cassimeris L. Reorganization of paclitaxel-stabilized microtubule arrays at mitotic entry: roles of depolymerizing kinesins and severing proteins. *Cancer Biol Ther.* 2019;20(10):1337–1347.
 111. Schwab CL, English DP, Roque DM, Santin AD. Taxanes: their impact on gynecologic malignancy. *Anti Cancer Drugs.* 2014;25(5):522–535.
 112. Abu Samaan TM, Samec M, Liskova A, Kubatka P, Büsselberg D. Paclitaxel's mechanistic and clinical effects on breast cancer. *Biomolecules.* 2019;9(12):789.
 113. Zhu L, Chen L. Progress in research on paclitaxel and tumor immunotherapy. *Cell Mol Biol Lett.* 2019;24:40.
 114. Mahalaxmi I, Devi SM, Kaavya J, Arul N, Balachandrar V, Santhy KS. New insight into NANOG: a novel therapeutic target for ovarian cancer (OC). *Eur J Pharmacol.* 2019;852:51–57.
 115. Postma TJ, Vermorken JB, Liefing AJ, Pinedo HM, Heimans JJ. Paclitaxel-induced neuropathy. *Ann Oncol.* 1995;6(5):489–494.
 116. Das T, Anand U, Pandey SK, et al. Therapeutic strategies to overcome taxane resistance in cancer. *Drug Resist Updates.* 2021;55:100754.
 117. Stähelin HF, von Wartburg A. The chemical and biological route from podophyllotoxin glucoside to etoposide: ninth Cain memorial Award lecture. *Cancer Res.* 1991;51(1):5–15.
 118. Green JA, Tarpey AW, Warenius HM. Pharmacokinetic study of high dose etoposide infusion in patients with small cell lung cancer. *Acta Oncol.* 1988;27(6b):819–822.
 119. Infante Lara L, Fenner S, Ratcliffe S, et al. Coupling the core of the anticancer drug etoposide to an oligonucleotide induces topoisomerase II-mediated cleavage at specific DNA sequences. *Nucleic Acids Res.* 2018;46(5):2218–2233.
 120. Pommier Y, Leo E, Zhang H, Marchand C. DNA topoisomerases and their poisoning by anticancer and antibacterial drugs. *Chem Biol.* 2010;17(5):421–433.
 121. Bruni E, Reichle A, Scimeca M, Bonanno E, Ghibelli L. Lowering etoposide doses shifts cell demise from caspase-dependent to differentiation and caspase-3-independent apoptosis via DNA damage response, inducing AML culture extinction. *Front Pharmacol.* 2018;9:1307.
 122. Hande KR. Etoposide: four decades of development of a topoisomerase II inhibitor. *Eur J Cancer.* 1998;34(10):1514–1521.
 123. Wall ME. Camptothecin and taxol: discovery to clinic. *Med Res Rev.* 1998;18(5):299–314.
 124. Martino E, Della Volpe S, Terribile E, et al. The long story of camptothecin: from traditional medicine to drugs. *Bioorg Med Chem Lett.* 2017;27(4):701–707.
 125. Yu S, Huang QQ, Luo Y, Lu W. Total synthesis of camptothecin and SN-38. *J Org Chem.* 2012;77(1):713–717.
 126. Li F, Jiang T, Li Q, Ling X. Camptothecin (CPT) and its derivatives are known to target topoisomerase I (Top1) as their mechanism of action: did we miss something in CPT analogue molecular targets for treating human disease such as cancer? *Am J Cancer Res.* 2017;7(12):2350–2394.
 127. Guo Y, Shi M, Shen X, Yang C, Yang L, Zhang J. Capecitabine plus irinotecan versus 5-FU/leucovorin plus irinotecan in the treatment of colorectal cancer: a meta-analysis. *Clin Colorectal Cancer.* 2014;13(2):110–118.
 128. Herben VM, ten Bokkel Huinink WW, Beijnen JH. Clinical pharmacokinetics of topotecan. *Clin Pharmacokinet.* 1996;31(2):85–102.
 129. Pommier Y. Topoisomerase I inhibitors: camptothecins and beyond. *Nat Rev Cancer.* 2006;6(10):789–802.
 130. Tomicic MT, Christmann M, Kaina B. Topotecan-triggered degradation of topoisomerase I is p53-dependent and impacts cell survival. *Cancer Res.* 2005;65(19):8920–8926.
 131. Bailly C. Irinotecan: 25 years of cancer treatment. *Pharmacol Res.* 2019;148:104398.

132. Sawada S, Yokokura T, Miyasaka T. Synthesis of CPT-11 (irinotecan hydrochloride trihydrate). *Ann N Y Acad Sci*. 1996; 803:13–28.
133. Sobell HM. Actinomycin and DNA transcription. *Proc Natl Acad Sci USA*. 1985;82(16):5328–5331.
134. Avendaño C, Menéndez JC. *Medicinal Chemistry of Anti-cancer Drugs*. Amsterdam: Elsevier; 2008.
135. Arndt C, Hawkins D, Anderson JR, Breitfeld P, Womer R, Meyer W. Age is a risk factor for chemotherapy-induced hepatopathy with vincristine, dactinomycin, and cyclophosphamide. *J Clin Oncol*. 2004;22(10):1894–1901.
136. Al-Hazmi A, Zwaan J, Awad A, Al-Mesfer S, Mullaney PB, Wheeler DT. Effectiveness and complications of mitomycin C use during pediatric glaucoma surgery. *Ophthalmology*. 1998; 105(10):1915–1920.
137. Crooke ST, Bradner WT. Mitomycin C: a review. *Cancer Treat Rev*. 1976;3(3):121–139.
138. Tomasz M. Mitomycin C: small, fast and deadly (but very selective). *Chem Biol*. 1995;2(9):575–579.
139. Noll DM, Mason TM, Miller PS. Formation and repair of inter-strand cross-links in DNA. *Chem Rev*. 2006;106(2):277–301.
140. Haffty BG, Wilson LD, Son YH, et al. Concurrent chemoradiotherapy with mitomycin C compared with porfiromycin in squamous cell cancer of the head and neck: final results of a randomized clinical trial. *Int J Radiat Oncol Biol Phys*. 2005; 61(1):119–128.
141. Murphy T, Yee KWL. Cytarabine and daunorubicin for the treatment of acute myeloid leukemia. *Expert Opin Pharmacother*. 2017;18(16):1765–1780.
142. Tan C, Tasaka H, Yu KP, Murphy ML, Karnofsky DA. Daunomycin, an antitumor antibiotic, in the treatment of neoplastic disease. Clinical evaluation with special reference to childhood leukemia. *Cancer*. 1967;20(3):333–353.
143. Fornari FA, Randolph JK, Yalowich JC, Ritke MK, Gewirtz DA. Interference by doxorubicin with DNA unwinding in MCF-7 breast tumor cells. *Mol Pharmacol*. 1994; 45(4):649–656.
144. Bodley A, Liu LF, Israel M, et al. DNA topoisomerase II-mediated interaction of doxorubicin and daunorubicin congeners with DNA. *Cancer Res*. 1989;49(21):5969–5978.
145. Zhang G, Fang L, Zhu L, et al. Syntheses and biological activities of disaccharide daunorubicins. *J Med Chem*. 2005; 48(16):5269–5278.
146. Yu Z, Yan B, Gao L, et al. Targeted delivery of bleomycin: a comprehensive anticancer review. *Curr Cancer Drug Targets*. 2016;16(6):509–521.
147. Lown JW, Sim SK. The mechanism of the bleomycin-induced cleavage of DNA. *Biochem Biophys Res Commun*. 1977;77(4): 1150–1157.
148. Weber GF. DNA damaging drugs. In: *Molecular Therapies of Cancer*. Cham.: Springer; 2015.
149. Meldrum C, Doyle MA, Tothill RW. Next-generation sequencing for cancer diagnostics: a practical perspective. *Clin Biochem Rev*. 2011;32(4):177–195.
150. Kalra RS, Cheung CT, Chaudhary A, Prakash J, Kaul SC, Wadhwa R. CARF (Collaborator of ARF) overexpression in p53-deficient cells promotes carcinogenesis. *Mol Oncol*. 2015; 9(9):1877–1889.
151. Joshi V, Upadhyay A, Prajapati VK, Mishra A. How autophagy can restore proteostasis defects in multiple diseases? *Med Res Rev*. 2020;40(4):1385–1439.
152. Mishra R, Upadhyay A, Prajapati VK, Mishra A. Proteasome-mediated proteostasis: novel medicinal and pharmacological strategies for diseases. *Med Res Rev*. 2018;38(6):1916–1973.
153. Upadhyay A, Amanullah A, Chhangani D, Joshi V, Mishra R, Mishra A. Ibuprofen induces mitochondrial-mediated apoptosis through proteasomal dysfunction. *Mol Neurobiol*. 2016;53(10):6968–6981.
154. Na Y, Kaul SC, Ryu J, et al. Stress chaperone mortalin contributes to epithelial-mesenchymal transition and cancer metastasis. *Cancer Res*. 2016;76(9):2754–2765.
155. Bapat SA, Krishnan A, Ghanate AD, Kusumbe AP, Kalra RS. Gene expression: protein interaction systems network modeling identifies transformation-associated molecules and pathways in ovarian cancer. *Cancer Res*. 2010;70(12): 4809–4819.
156. Stringer AM, Gibson RJ, Logan RM, et al. Irinotecan-induced mucositis is associated with changes in intestinal mucins. *Cancer Chemother Pharmacol*. 2009;64(1):123–132.
157. Thorpe D, Stringer A, Butler R. Chemotherapy-induced mucositis: the role of mucin secretion and regulation, and the enteric nervous system. *Neurotoxicology*. 2013;38: 101–105.
158. Mitchison TJ. The proliferation rate paradox in antimitotic chemotherapy. *Mol Biol Cell*. 2012;23(1):1–6.
159. McQuade RM, Stojanovska V, Abalo R, Bornstein JC, Nurgali K. Chemotherapy-induced constipation and diarrhea: pathophysiology, current and emerging treatments. *Front Pharmacol*. 2016;7:414.
160. Bower JE. Cancer-related fatigue: mechanisms, risk factors, and treatments. *Nat Rev Clin Oncol*. 2014;11(10):597–609.
161. Wang Y, Probin V, Zhou D. Cancer therapy-induced residual bone marrow injury-Mechanisms of induction and implication for therapy. *Curr Cancer Ther Rev*. 2006;2(3):271–279.
162. Kuter DJ. Managing thrombocytopenia associated with cancer chemotherapy. *Oncology*. 2015;29(4):282–294.
163. Manikandan R, Kumar S, Dorairajan LN. Hemorrhagic cystitis: a challenge to the urologist. *Indian J Urol*. 2010;26(2): 159–166.
164. Horie S, Oya M, Nangaku M, et al. Guidelines for treatment of renal injury during cancer chemotherapy 2016. *Clin Exp Nephrol*. 2018;22(1):210–244.
165. Maor Y, Malnick S. Liver injury induced by anticancer chemotherapy and radiation therapy. *Int J Hepatol*. 2013; 2013:815105.
166. Yeh ET, Tong AT, Lenihan DJ, et al. Cardiovascular complications of cancer therapy: diagnosis, pathogenesis, and management. *Circulation*. 2004;109(25):3122–3131.
167. Schover LR, van der Kaaij M, van Dorst E, Creutzberg C, Huyghe E, Kiserud CE. Sexual dysfunction and infertility as late effects of cancer treatment. *EJC Suppl*. 2014;12(1): 41–53.
168. Voznesensky M, Annam K, Kreder KJ. Understanding and managing erectile dysfunction in patients treated for cancer. *J Oncol Pract*. 2016;12(4):297–304.
169. Mitchell T, Turton P. 'Chemobrain': concentration and memory effects in people receiving chemotherapy - a descriptive phenomenological study. *Eur J Cancer Care*. 2011;20(4): 539–548.
170. Moore HC. An overview of chemotherapy-related cognitive dysfunction, or 'chemobrain'. *Oncology*. 2014;28(9):797–804.
171. Dutta V. Chemotherapy, neurotoxicity, and cognitive changes in breast cancer. *J Cancer Res Therapeut*. 2011;7(3):264–269.
172. Omoti A, Omoti C. Ocular toxicity of systemic anticancer chemotherapy. *Pharm Pract*. 2006;4:55–59.
173. Al-Tweigeri T, Nabholz JM, MacKey JR. Ocular toxicity and cancer chemotherapy. A review. *Cancer*. 1996;78(7): 1359–1373.
174. Curigliano G, Cardinale D, Suter T, et al. Cardiovascular toxicity induced by chemotherapy, targeted agents and radiotherapy: ESMO Clinical Practice Guidelines. *Ann Oncol*. 2012;23(Suppl 7):vii155–vii166.
175. Cohen JD, Li L, Wang Y, et al. Detection and localization of surgically resectable cancers with a multi-analyte blood test. *Science*. 2018;359(6378):926–930.

176. Alix-Panabières C, Pantel K. Clinical applications of circulating tumor cells and circulating tumor DNA as liquid biopsy. *Cancer Discov.* 2016;6(5):479–491.
177. Li L, Gao R, Yu Y, et al. Tumor suppressor activity of miR-451: identification of CARF as a new target. *Sci Rep.* 2018; 8(1):375.
178. Shohdy KS, West HJ. Circulating tumor DNA testing-liquid biopsy of a cancer. *JAMA Oncol.* 2020;6(5):792.
179. Vivek R, Thangam R, NipunBabu V, et al. Multifunctional HER2-antibody conjugated polymeric nanocarrier-based drug delivery system for multi-drug-resistant breast cancer therapy. *ACS Appl Mater Interfaces.* 2014;6(9):6469–6480.
180. Alley SC, Okeley NM, Senter PD. Antibody-drug conjugates: targeted drug delivery for cancer. *Curr Opin Chem Biol.* 2010; 14(4):529–537.
181. Wang X, Wang Y, Chen ZG, Shin DM. Advances of cancer therapy by nanotechnology. *Cancer Res Treat.* 2009;41(1): 1–11.
182. Firer MA, Gellerman G. Targeted drug delivery for cancer therapy: the other side of antibodies. *J Hematol Oncol.* 2012; 5:70.
183. Kutova OM, Guryev EL, Sokolova EA, Alzeibak R, Balalaeva IV. Targeted delivery to tumors: multidirectional strategies to improve treatment efficiency. *Cancers.* 2019;11(1):68.
184. Cho K, Wang X, Nie S, Chen ZG, Shin DM. Therapeutic nanoparticles for drug delivery in cancer. *Clin Cancer Res.* 2008; 14(5):1310–1316.
185. Rosenblum D, Joshi N, Tao W, Karp JM, Peer D. Progress and challenges towards targeted delivery of cancer therapeutics. *Nat Commun.* 2018;9(1):1410.
186. Sutradhar KB, Amin ML. Nanotechnology in cancer drug delivery and selective targeting. *ISRN Nanotechnol.* 2014;2014: 939378.
187. Lammers T, Kiessling F, Ashford M, Hennink W, Crommelin D, Storm G. Cancer nanomedicine: is targeting our target? *Nat Rev Mater.* 2016;1(9):16069.
188. El-Readi MZ, Althubiti MA. Cancer nanomedicine: a new era of successful targeted therapy. *J Nanomater.* 2019;2019: 4927312.
189. Dias-Santagata D, Akhavanfard S, David SS, et al. Rapid targeted mutational analysis of human tumours: a clinical platform to guide personalized cancer medicine. *EMBO Mol Med.* 2010;2(5):146–158.
190. Dong L, Wang W, Li A, et al. Clinical next generation sequencing for precision medicine in cancer. *Curr Genom.* 2015;16(4):253–263.
191. Fedorenko IV, Gibney GT, Sondak VK, Smalley KSM. Beyond BRAF: where next for melanoma therapy? *Br J Cancer.* 2015; 112(2):217–226.
192. Zaman A, Wu W, Bivona TG. Targeting oncogenic BRAF: past, present, and future. *Cancers.* 2019;11(8):1197.
193. André F, Ciruelos E, Rubovszky G, et al. Alpelisib for PIK3CA-mutated, hormone receptor-positive advanced breast cancer. *N Engl J Med.* 2019;380(20):1929–1940.
194. Kalra RS, Bapat SA. *Proteomics to Predict Loss of RXR-γ during Progression of Epithelial Ovarian Cancer. Methods in Molecular Biology.* New York, NY: New York: Springer; 2019: 1–14.
195. Soda M, Choi YL, Enomoto M, et al. Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. *Nature.* 2007;448(7153):561–566.
196. Hyman DM, Puzanov I, Subbiah V, et al. Vemurafenib in multiple nonmelanoma cancers with BRAF V600 mutations. *N Engl J Med.* 2015;373(8):726–736.
197. Babina IS, Turner NC. Advances and challenges in targeting FGFR signalling in cancer. *Nat Rev Cancer.* 2017;17(5):318–332.
198. Tutt A, Robson M, Garber JE, et al. Oral poly(ADP-ribose) polymerase inhibitor olaparib in patients with BRCA1 or BRCA2 mutations and advanced breast cancer: a proof-of-concept trial. *Lancet.* 2010;376(9737):235–244.
199. Fong PC, Boss DS, Yap TA, et al. Inhibition of poly(ADP-ribose) polymerase in tumors from BRCA mutation carriers. *N Engl J Med.* 2009;361(2):123–134.
200. Chaudhary A, Kalra RS, Huang C, Prakash J, Kaul SC, Wadhwa R. 2, 3-dihydro-3β-methoxy withaferin-A protects normal cells against stress: molecular evidence of its potent cytoprotective activity. *J Nat Prod.* 2017;80(10):2756–2760.
201. Friedman AA, Letai A, Fisher DE, Flaherty KT. Precision medicine for cancer with next-generation functional diagnostics. *Nat Rev Cancer.* 2015;15(12):747–756.
202. de la Fuente-Núñez C, Silva ON, Lu TK, Franco OL. Antimicrobial peptides: role in human disease and potential as immunotherapies. *Pharmacol Ther.* 2017;178:132–140.
203. Felício MR, Silva ON, Gonçalves S, Santos NC, Franco OL. Peptides with dual antimicrobial and anticancer activities. *Front Chem.* 2017;5:5.
204. Hilchie AL, Hoskin DW, Power Coombs MR. Anticancer activities of natural and synthetic peptides. *Adv Exp Med Biol.* 2019;1117:131–147.
205. Mwangi J, Hao X, Lai R, Zhang ZY. Antimicrobial peptides: new hope in the war against multidrug resistance. *Zool Res.* 2019;40(6):488–505.
206. Qin Y, Qin ZD, Chen J, et al. From antimicrobial to anticancer peptides: the transformation of peptides. *Recent Pat Anti-Cancer Drug Discov.* 2019;14(1):70–84.
207. Lei J, Sun L, Huang S, et al. The antimicrobial peptides and their potential clinical applications. *Am J Transl Res.* 2019; 11(7):3919–3931.
208. Roudi R, Syn NL, Roudbary M. Antimicrobial peptides as biologic and immunotherapeutic agents against cancer: a comprehensive overview. *Front Immunol.* 2017;8:1320.
209. Tornesello AL, Borrelli A, Buonaguro L, Buonaguro FM, Tornesello ML. Antimicrobial peptides as anticancer agents: functional properties and biological activities. *Molecules.* 2020;25(12):2850.
210. Khare T, Anand U, Dey A, et al. Exploring phytochemicals for combating antibiotic resistance in microbial pathogens. *Front Pharmacol.* 2021;12:720726.
211. van Harten RM, van Woudenberg E, van Dijk A, Haagsman HP. Cathelicidins: immunomodulatory antimicrobials. *Vaccines.* 2018;6(3):63.
212. Wang G, Narayana JL, Mishra B, et al. *Design of Antimicrobial Peptides: Progress Made with Human Cathelicidin LL-37. Advances in Experimental Medicine and Biology.* Singapore: Springer Singapore; 2019:215–240.
213. Fabisiak A, Murawska N, Fichna J. LL-37: cathelicidin-related antimicrobial peptide with pleiotropic activity. *Pharmacol Rep.* 2016;68(4):802–808.
214. Chen X, Ji S, Si J, et al. Human cathelicidin antimicrobial peptide suppresses proliferation, migration and invasion of oral carcinoma HSC-3 cells via a novel mechanism involving caspase-3 mediated apoptosis. *Mol Med Rep.* 2020;22(6): 5243–5250.
215. Porter RJ, Murray GI, Alnabulsi A, et al. Colonic epithelial cathelicidin (LL-37) expression intensity is associated with progression of colorectal cancer and presence of CD8⁺ T cell infiltrate. *J Pathol Clin Res.* 2021;7(5):495–506.
216. Wang J, Cheng M, Law IKM, Ortiz C, Sun M, Koon HW. Cathelicidin suppresses colon cancer metastasis via a P2RX7-dependent mechanism. *Mol Ther Oncolytics.* 2019;12: 195–203.
217. Biswalo LS, da Costa Sousa MG, Rezende TMB, Dias SC, Franco OL. Antimicrobial peptides and nanotechnology, recent advances and challenges. *Front Microbiol.* 2018;9:855.
218. Niemirowicz K, Prokop I, Wilczewska AZ, et al. Magnetic nanoparticles enhance the anticancer activity of cathelicidin

- LL-37 peptide against colon cancer cells. *Int J Nanomed.* 2015;10:3843–3853.
219. Fan R, Tong A, Li X, et al. Enhanced antitumor effects by docetaxel/LL37-loaded thermosensitive hydrogel nanoparticles in peritoneal carcinomatosis of colorectal cancer. *Int J Nanomed.* 2015;10:7291–7305.
 220. Chauhan S, Dhawan DK, Saini A, Preet S. Antimicrobial peptides against colorectal cancer—a focused review. *Pharmacol Res.* 2021;167:105529.
 221. Dolkar T, Trinidad CM, Nelson KC, et al. Dermatologic toxicity from novel therapy using antimicrobial peptide LL-37 in melanoma: a detailed examination of the clinicopathologic features. *J Cutan Pathol.* 2018;45(7):539–544.
 222. Divyashree M, Mani MK, Reddy D, et al. Clinical applications of antimicrobial peptides (AMPs): where do we stand now? *Protein Pept Lett.* 2020;27(2):120–134.
 223. Karlitepe A, Ozalp O, Avci CB. New approaches for cancer immunotherapy. *Tumour Biol.* 2015;36(6):4075–4078.
 224. Weiner LM, Murray JC, Shuptrine CW. Antibody-based immunotherapy of cancer. *Cell.* 2012;148(6):1081–1084.
 225. Vigneron N. Human tumor antigens and cancer immunotherapy. *BioMed Res Int.* 2015;2015:948501.
 226. Kim I, Sanchez K, McArthur HL, Page D. Immunotherapy in triple-negative breast cancer: present and future. *Curr Breast Cancer Rep.* 2019;11(4):259–271.
 227. Marra A, Viale G, Curigliano G. Recent advances in triple negative breast cancer: the immunotherapy era. *BMC Med.* 2019;17(1):90.
 228. Sami M, Bagheri L, Szwczuk MR. Current challenges in cancer immunotherapy: multimodal approaches to improve efficacy and patient response rates. *JAMA Oncol.* 2019;2019:4508794.
 229. Paz-Ares L, Luft A, Vicente D, et al. Pembrolizumab plus chemotherapy for squamous non-small-cell lung cancer. *N Engl J Med.* 2018;379(21):2040–2051.
 230. Finn OJ. Vaccines for cancer prevention: a practical and feasible approach to the cancer epidemic. *Cancer Immunol Res.* 2014;2(8):708–713.
 231. Pérez-García J, Soberino J, Racca F, Gion M, Stradella A, Cortés J. Atezolizumab in the treatment of metastatic triple-negative breast cancer. *Expert Opin Biol Ther.* 2020;20(9):981–989.
 232. Reddy SM, Carroll E, Nanda R. Atezolizumab for the treatment of breast cancer. *Expert Rev Anticancer Ther.* 2020;20(3):151–158.
 233. Cyprian FS, Akhtar S, Gatalica Z, Vranic S. Targeted immunotherapy with a checkpoint inhibitor in combination with chemotherapy: a new clinical paradigm in the treatment of triple-negative breast cancer. *Bosn J Basic Med Sci.* 2019;19(3):227–233.
 234. Ledford H. Engineered cell therapy for cancer gets thumbs up from FDA advisers. *Nature.* 2017;547(7663):270.
 235. Schirrmacher V. From chemotherapy to biological therapy: a review of novel concepts to reduce the side effects of systemic cancer treatment (Review). *Int J Oncol.* 2019;54(2):407–419.
 236. Vanneman M, Dranoff G. Combining immunotherapy and targeted therapies in cancer treatment. *Nat Rev Cancer.* 2012;12(4):237–251.
 237. Hughes PE, Caenepeel S, Wu LC. Targeted therapy and checkpoint immunotherapy combinations for the treatment of cancer. *Trends Immunol.* 2016;37(7):462–476.
 238. Grosser R, Cherkassky L, Chintala N, Adusumilli PS. Combination immunotherapy with CAR T cells and checkpoint blockade for the treatment of solid tumors. *Cancer Cell.* 2019;36(5):471–482.
 239. Chiossone L, Dumas PY, Vienne M, Vivier E. Natural killer cells and other innate lymphoid cells in cancer. *Nat Rev Immunol.* 2018;18(11):671–688.
 240. Anyaegbu CC, Lake RA, Heel K, Robinson BW, Fisher SA. Chemotherapy enhances cross-presentation of nuclear tumor antigens. *PLoS One.* 2014;9(9):e107894.
 241. Konstantinopoulos PA, Cheng SC, Wahner Hendrickson AE, et al. Berzosertib plus gemcitabine versus gemcitabine alone in platinum-resistant high-grade serous ovarian cancer: a multicentre, open-label, randomised, phase 2 trial. *Lancet Oncol.* 2020;21(7):957–968.
 242. Reck M. Pembrolizumab as first-line therapy for metastatic non-small-cell lung cancer. *Immunotherapy.* 2018;10(2):93–105.
 243. Kang C, Syed YY. Atezolizumab (in combination with nab-paclitaxel): a review in advanced triple-negative breast cancer. *Drugs.* 2020;80(6):601–607.
 244. Allard B, Allard D, Buisseret L, Stagg J. The adenosine pathway in immuno-oncology. *Nat Rev Clin Oncol.* 2020;17(10):611–629.
 245. Pasquini S, Contri C, Borea PA, Vincenzi F, Varani K. Adenosine and inflammation: here, there and everywhere. *Int J Mol Sci.* 2021;22(14):7685.
 246. Franco R, Rivas-Santisteban R, Navarro G, Reyes-Resina I. Adenosine receptor antagonists to combat cancer and to boost anti-cancer chemotherapy and immunotherapy. *Cells.* 2021;10(11):2831.
 247. Helms RS, Powell JD. Rethinking the adenosine-A_{2A}R checkpoint: implications for enhancing anti-tumor immunotherapy. *Curr Opin Pharmacol.* 2020;53:77–83.
 248. Yu F, Zhu C, Xie Q, Wang Y. Adenosine A_{2A} receptor antagonists for cancer immunotherapy. *J Med Chem.* 2020;63(21):12196–12212.
 249. Yan W, Ling L, Wu Y, et al. Structure-based design of dual-acting compounds targeting adenosine A_{2A} receptor and histone deacetylase as novel tumor immunotherapeutic agents. *J Med Chem.* 2021;64(22):16573–16597.
 250. Churov A, Zhulai G. Targeting adenosine and regulatory T cells in cancer immunotherapy. *Hum Immunol.* 2021;82(4):270–278.
 251. Marks DL, Olson RL, Fernandez-Zapico ME. Epigenetic control of the tumor microenvironment. *Epigenomics.* 2016;8(12):1671–1687.
 252. Lodewijk I, Nunes SP, Henrique R, Jerónimo C, Dueñas M, Paramio JM. Tackling tumor microenvironment through epigenetic tools to improve cancer immunotherapy. *Clin Epigenet.* 2021;13(1):63.
 253. Pan X, Zheng L. Epigenetics in modulating immune functions of stromal and immune cells in the tumor microenvironment. *Cell Mol Immunol.* 2020;17(9):940–953.
 254. Duan Q, Zhang H, Zheng J, Zhang L. Turning cold into hot: firing up the tumor microenvironment. *Trends Cancer.* 2020;6(7):605–618.
 255. Lim WA, June CH. The principles of engineering immune cells to treat cancer. *Cell.* 2017;168(4):724–740.
 256. Murciano-Goroff YR, Warner AB, Wolchok JD. The future of cancer immunotherapy: microenvironment-targeting combinations. *Cell Res.* 2020;30(6):507–519.
 257. Hulen TM, Chamberlain CA, Svane IM, Met Ö. ACT up TIL now: the evolution of tumor-infiltrating lymphocytes in adoptive cell therapy for the treatment of solid tumors. *Immunology.* 2021;1(3):194–211.
 258. Newick K, O'Brien S, Moon E, Albelda SM. CAR T cell therapy for solid tumors. *Annu Rev Med.* 2017;68:139–152.
 259. Singh AK, McGuirk JP. CAR T cells: continuation in a revolution of immunotherapy. *Lancet Oncol.* 2020;21(3):e168–e178.
 260. Jazirehi AR. Molecular analysis of elements of melanoma insensitivity to TCR-engineered adoptive cell therapy. *Int J Mol Sci.* 2021;22(21):11726.

261. Tan E, Gakhar N, Kirtane K. TCR gene-engineered cell therapy for solid tumors. *Best Pract Res Clin Haematol*. 2021;34(3): 101285.
262. Ö Met, Jensen KM, Chamberlain CA, Donia M, Svane IM. Principles of adoptive T cell therapy in cancer. *Semin Immunopathol*. 2019;41(1):49–58.
263. Wolf B, Zimmermann S, Arber C, Irving M, Trueb L, Coukos G. Safety and tolerability of adoptive cell therapy in cancer. *Drug Saf*. 2019;42(2):315–334.
264. Kaufman HL, Maciorowski D. Advancing oncolytic virus therapy by understanding the biology. *Nat Rev Clin Oncol*. 2021; 18(4):197–198.
265. Ferrucci PF, Pala L, Conforti F, Cocorocchio E. Talimogene laherparepvec (T-VEC):an intralesional cancer immunotherapy for advanced melanoma. *Cancers*. 2021;13(6): 1383.
266. Lu L, Jiang J, Zhan M, et al. Targeting tumor-associated antigens in hepatocellular carcinoma for immunotherapy: past pitfalls and future strategies. *Hepatology*. 2021;73(2): 821–832.
267. Liu Z, Han C, Fu YX. Targeting innate sensing in the tumor microenvironment to improve immunotherapy. *Cell Mol Immunol*. 2020;17(1):13–26.
268. Bejarano L, Jordão MJC, Joyce JA. Therapeutic targeting of the tumor microenvironment. *Cancer Discov*. 2021;11(4): 933–959.
269. Anand U, Tudu CK, Nandy S, et al. Ethnodermatological use of medicinal plants in India: from ayurvedic formulations to clinical perspectives - a review. *J Ethnopharmacol*. 2022;284: 114744.
270. Paul S, Chakraborty S, Anand U, et al. Withania somnifera (L.) Dunal (Ashwagandha):a comprehensive review on ethnopharmacology, pharmacotherapeutics, biomedical and toxicological aspects. *Biomed Pharmacother*. 2021;143: 112175.
271. Lin SR, Chang CH, Hsu CF, et al. Natural compounds as potential adjuvants to cancer therapy: preclinical evidence. *Br J Pharmacol*. 2020;177(6):1409–1423.
272. Nobili S, Lippi D, Witort E, et al. Natural compounds for cancer treatment and prevention. *Pharmacol Res*. 2009;59(6): 365–378.
273. Mitra S, Anand U, Jha NK, et al. Anticancer applications and pharmacological properties of piperidine and piperine: a comprehensive review on molecular mechanisms and therapeutic perspectives. *Front Pharmacol*. 2021;12: 772418.
274. Mitra S, Anand U, Sanyal R, et al. Neoechinulins: molecular, cellular, and functional attributes as promising therapeutics against cancer and other human diseases. *Biomed Pharmacother*. 2022;145:112378.
275. Bandyopadhyay S, Anand U, Gadekar VS, et al. Dioscin: a review on pharmacological properties and therapeutic values. *Biofactors*. 2022;48(1):22–55.
276. Datta S, Ramamurthy PC, Anand U, et al. Wonder or evil? : multifaceted health hazards and health benefits of *Cannabis sativa* and its phytochemicals. *Saudi J Biol Sci*. 2021;28(12): 7290–7313.
277. Dutta T, Anand U, Saha SC, et al. Advancing urban ethnopharmacology: a modern concept of sustainability, conservation and cross-cultural adaptations of medicinal plant lore in the urban environment. *Conserv Physiol*. 2021;9(1): coab073.
278. Tournilhac O, Cazin B, Leprêtre S, et al. Impact of frontline fludarabine and cyclophosphamide combined treatment on peripheral blood stem cell mobilization in B-cell chronic lymphocytic leukemia. *Blood*. 2004;103(1):363–365.
279. Chu E, DeVita VT. *Physician's Cancer Chemotherapy Drug Manual* 2015. 15th ed. Burlington, MA: Jones & Bartlett Learning; 2015.
280. Yerram P, Reiss SN, Modelevsky L, Gavrilovic IT, Kaley T. Evaluation of toxicity of carmustine with or without bevacizumab in patients with recurrent or progressive high grade gliomas. *J Neuro Oncol*. 2019;145(1):57–63.
281. Karahoca M, Momparler RL. Pharmacokinetic and pharmacodynamic analysis of 5-aza-2'-deoxycytidine (decitabine) in the design of its dose-schedule for cancer therapy. *Clin Epigenet*. 2013;5(1):3.
282. Vanneste M, Huang Q, Li M, et al. High content screening identifies monensin as an EMT-selective cytotoxic compound. *Sci Rep*. 2019;9(1):1200.
283. Rycenga HB, Long DT. The evolving role of DNA inter-strand crosslinks in chemotherapy. *Curr Opin Pharmacol*. 2018;41: 20–26.
284. Samuels BL, Bitran JD. High-dose intravenous melphalan: a review. *J Clin Oncol*. 1995;13(7):1786–1799.
285. Ma MT, He M, Wang Y, et al. miR-487a resensitizes mitoxantrone (MX)-resistant breast cancer cells (MCF-7/MX) to MX by targeting breast cancer resistance protein (BCRP/ABCG2). *Cancer Lett*. 2013;339(1):107–115.
286. Graham J, Muhsin M, Kirkpatrick P. Oxaliplatin. *Nat Rev Drug Discov*. 2004;3(1):11–12.
287. Jacinto FV, Esteller M. MGMT hypermethylation: a prognostic foe, a predictive friend. *DNA Repair*. 2007;6(8):1155–1160.
288. Arrieta O, Lara-Mejía L, Zatarain-Barrón ZL. Carboplatin plus etoposide or topotecan for small-cell lung cancer. *Lancet Oncol*. 2020;21(9):1132–1134.
289. Hwang DY, Kim SJ, Cheong JW, et al. High pre-transplant serum ferritin and busulfan-thiotepa conditioning regimen as risk factors for hepatic sinusoidal obstructive syndrome after autologous stem cell transplantation in patients with malignant lymphoma. *Leuk Lymphoma*. 2016;57(1):51–57.
290. D'Incalci M, Badri N, Galmarini CM, Allavena P. Trabectedin, a drug acting on both cancer cells and the tumour microenvironment. *Br J Cancer*. 2014;111(4):646–650.
291. Maltzman JS, Koretzky GA. Azathioprine: old drug, new actions. *J Clin Invest*. 2003;111(8):1122–1124.
292. Hasanali ZS, Saroya BS, Stuart A, et al. Epigenetic therapy overcomes treatment resistance in T cell prolymphocytic leukemia. *Sci Transl Med*. 2015;7(293):293ra102.
293. Bonate PL, Arthaud L, Cantrell WR, Stephenson K, Secrist JA, Weitman S. Discovery and development of clofarabine: a nucleoside analogue for treating cancer. *Nat Rev Drug Discov*. 2006;5(10):855–863.
294. Ricci F, Tedeschi A, Morra E, Montillo M. Fludarabine in the treatment of chronic lymphocytic leukemia: a review. *Therapeut Clin Risk Manag*. 2009;5(1):187–207.
295. Mini E, Nobili S, Caciagli B, Landini I, Mazzei T. Cellular pharmacology of gemcitabine. *Ann Oncol*. 2006;17(Suppl 5): v7–v12.
296. Engels FK, Sparreboom A, Mathot RAA, Verweij J. Potential for improvement of docetaxel-based chemotherapy: a pharmacological review. *Br J Cancer*. 2005;93(2):173–177.
297. Cohen MH, Johnson JR, Massie T, et al. Approval summary: nelarabine for the treatment of T-cell lymphoblastic leukemia/lymphoma. *Clin Cancer Res*. 2006;12(18):5329–5335.
298. Cannon T, Mobarek D, Wegge J, Tabbara IA. Hairy cell leukemia: current concepts. *Cancer Invest*. 2008;26(8):860–865.
299. Wang H, Wang Y. LC-MS/MS coupled with stable isotope dilution method for the quantification of 6-thioguanine and S6-methylthioguanine in genomic DNA of human cancer cells treated with 6-thioguanine. *Anal Chem*. 2010;82(13): 5797–5803.

300. Sertel S, Tome M, Briehl MM, et al. Factors determining sensitivity and resistance of tumor cells to arsenic trioxide. *PLoS One*. 2012;7(5):e35584.
301. Madaan K, Kaushik D, Verma T. Hydroxyurea: a key player in cancer chemotherapy. *Expert Rev Anticancer Ther*. 2012;12(1):19–29.
302. Scripture CD, Szebeni J, Loos WJ, Figg WD, Sparreboom A. Comparative *in vitro* properties and clinical pharmacokinetics of Paclitaxel following the administration of taxol(r) and paxene(r). *Cancer Biol Ther*. 2005;4(5):555–560.
303. Alken S, Kelly CM. Benefit risk assessment and update on the use of docetaxel in the management of breast cancer. *Cancer Manag Res*. 2013;5:357–365.
304. Chan ES, Cronstein BN. Mechanisms of action of methotrexate. *Bull Hosp Jt Dis*. 2013;71(Suppl 1):S5–S8.
305. Adjei AA. Pharmacology and mechanism of action of pemetrexed. *Clin Lung Cancer*. 2004;5(Suppl 2):S51–S55.
306. Weatherstone K, Meyer T. Streptozocin-based chemotherapy is not history in neuroendocrine tumours. *Targeted Oncol*. 2012;7(3):161–168.
307. Haque A, Rahman MA, Faizi MSH, Khan MS. Next generation antineoplastic agents: a review on structurally modified vinblastine (VBL) analogues. *Curr Med Chem*. 2018;25(14):1650–1662.
308. Beaver C. Vincristine minibag administration: a quality improvement project to minimize medical errors. *Clin J Oncol Nurs*. 2018;22(6):669–672.
309. Mehta-Shah N, Ratner L, Horwitz SM. Adult T-cell leukemia/lymphoma. *J Oncol Pract*. 2017;13(8):487–492.
310. Altinoz MA, Ozpinar A, Alturfan EE, Elmali I. Vinorelbine's anti-tumor actions may depend on the mitotic apoptosis, autophagy and inflammation: hypotheses with implications for chemo-immunotherapy of advanced cancers and pediatric gliomas. *J Chemother*. 2018;30(4):203–212.
311. Jäger U, Barcellini W, Broome CM, et al. Diagnosis and treatment of autoimmune hemolytic Anemia in adults: recommendations from the first international consensus meeting. *Blood Rev*. 2020;41:100648.
312. Kreitman RJ, Arons E. Update on hairy cell leukemia. *Clin Adv Hematol Oncol*. 2018;16(3):205–215.
313. Begna K, Abdelatif A, Schwager S, Hanson C, Pardanani A, Tefferi A. Busulfan for the treatment of myeloproliferative neoplasms: the Mayo Clinic experience. *Blood Cancer J*. 2016;6(5):e427.
314. Palmer J, McCune JS, Perales MA, et al. Personalizing busulfan-based conditioning: considerations from the American society for blood and marrow transplantation practice guidelines committee. *Biol Blood Marrow Transplant*. 2016;22(11):1915–1925.
315. Ho GY, Woodward N, Coward JIG. Cisplatin versus carboplatin: comparative review of therapeutic management in solid malignancies. *Crit Rev Oncol Hematol*. 2016;102:37–46.
316. Lee HZ, Miller BW, Kwitkowski VE, et al. U.S. food and drug administration approval: obinutuzumab in combination with chlorambucil for the treatment of previously untreated chronic lymphocytic leukemia. *Clin Cancer Res*. 2014;20(15):3902–3907.
317. Hew J, Pham D, Matthews Hew T, Minocha V. A novel treatment with obinutuzumab-chlorambucil in a patient with B-cell prolymphocytic leukemia: a case report and review of the literature. *J Investig Med High Impact Case Rep*. 2018;6::2324709618788674.
318. Makovec T. Cisplatin and beyond: molecular mechanisms of action and drug resistance development in cancer chemotherapy. *Radiol Oncol*. 2019;53(2):148–158.
319. Raudenska M, Balvan J, Fojtu M, Gumulec J, Masarik M. Unexpected therapeutic effects of cisplatin. *Metallomics*. 2019;11(7):1182–1199.
320. Casak SJ, Lemery SJ, Shen YL, et al. U.S. Food and drug administration approval: rituximab in combination with fludarabine and cyclophosphamide for the treatment of patients with chronic lymphocytic leukemia. *Oncol*. 2011;16(1):97–104.
321. Jiang G, Li RH, Sun C, Liu YQ, Zheng JN. Dacarbazine combined targeted therapy versus dacarbazine alone in patients with malignant melanoma: a meta-analysis. *PLoS One*. 2014;9(12):e111920.
322. Velho TR. Metastatic melanoma - a review of current and future drugs. *Drugs Context*. 2012;2012:212242.
323. Krauss AC, Gao X, Li L, et al. FDA approval summary: (daunorubicin and cytarabine) liposome for injection for the treatment of adults with high-risk acute myeloid leukemia. *Clin Cancer Res*. 2019;25(9):2685–2690.
324. Blair HA. Daunorubicin/cytarabine liposome: a review in acute myeloid leukaemia. *Drugs*. 2018;78(18):1903–1910.
325. Dhillon S. Decitabine/cedazuridine: first approval. *Drugs*. 2020;80(13):1373–1378.
326. Kanwal U, Irfan Bukhari N, Ovais M, Abass N, Hussain K, Raza A. Advances in nano-delivery systems for doxorubicin: an updated insight. *J Drug Target*. 2018;26(4):296–310.
327. Cagel M, Grotz E, Bernabeu E, Moretton MA, Chiappetta DA. Doxorubicin: nanotechnological overviews from bench to bedside. *Drug Discov Today*. 2017;22(2):270–281.
328. Buzdar AU, Suman VJ, Meric-Bernstam F, et al. Fluorouracil, epirubicin, and cyclophosphamide (FEC-75) followed by paclitaxel plus trastuzumab versus paclitaxel plus trastuzumab followed by FEC-75 plus trastuzumab as neoadjuvant treatment for patients with HER2-positive breast cancer (Z1041): a randomised, controlled, phase 3 trial. *Lancet Oncol*. 2013;14(13):1317–1325.
329. Hollingshead LM, Faulds D. Idarubicin. A review of its pharmacodynamic and pharmacokinetic properties, and therapeutic potential in the chemotherapy of cancer. *Drugs*. 1991;42(4):690–719.
330. Matz EL, Hsieh MH. Review of advances in uroprotective agents for cyclophosphamide- and ifosfamide-induced hemorrhagic cystitis. *Urology*. 2017;100:16–19.
331. de Man FM, Goey AKL, van Schaik RHN, Mathijssen RHJ, Bins S. Individualization of irinotecan treatment: a review of pharmacokinetics, pharmacodynamics, and pharmacogenetics. *Clin Pharmacokinet*. 2018;57(10):1229–1254.
332. KuKanich B, Warner M, Hahn K. Analysis of lomustine drug content in FDA-approved and compounded lomustine capsules. *J Am Vet Med Assoc*. 2017;250(3):322–326.
333. Sun J, Wei Q, Zhou Y, Wang J, Liu Q, Xu H. A systematic analysis of FDA-approved anticancer drugs. *BMC Syst Biol*. 2017;11(Suppl 5):87.
334. Fox EJ. Mechanism of action of mitoxantrone. *Neurology*. 2004;63(12 Suppl 6):S15–S18.
335. Ibrahim A, Hirschfeld S, Cohen MH, Griebel DJ, Williams GA, Pazdur R. FDA drug approval summaries: oxaliplatin. *Oncol*. 2004;9(1):8–12.
336. Stein A, Arnold D. Oxaliplatin: a review of approved uses. *Expert Opin Pharmacother*. 2012;13(1):125–137.
337. Mutter N, Stupp R. Temozolomide: a milestone in neuro-oncology and beyond? *Expert Rev Anticancer Ther*. 2006;6(8):1187–1204.
338. Ardzizoni A, Hansen H, Dombernowsky P, et al. Topotecan, a new active drug in the second-line treatment of small-cell lung cancer: a phase II study in patients with refractory and sensitive disease. The European organization for research and treatment of cancer early clinical studies group and new drug development office, and the lung cancer cooperative group. *J Clin Oncol*. 1997;15(5):2090–2096.
339. Garst J. Topotecan: an evolving option in the treatment of relapsed small cell lung cancer. *Therapeut Clin Risk Manag*. 2007;3(6):1087–1095.

340. Eckardt JR, von Pawel J, Pujol JL, et al. Phase III study of oral compared with intravenous topotecan as second-line therapy in small-cell lung cancer. *J Clin Oncol*. 2007; 25(15):2086–2092.
341. Kokolo MB, Fergusson D, O'Neill J, et al. Effectiveness and safety of thiotepea as conditioning treatment prior to stem cell transplant in patients with central nervous system lymphoma. *Leuk Lymphoma*. 2014;55(12):2712–2720.
342. Barone A, Chi DC, Theoret MR, et al. FDA approval summary: trabectedin for unresectable or metastatic liposarcoma or leiomyosarcoma following an anthracycline-containing regimen. *Clin Cancer Res*. 2017;23(24):7448–7453.
343. Rosenberg JD, Burian C, Waalen J, Saven A. Clinical characteristics and long-term outcome of young hairy cell leukemia patients treated with cladribine: a single-institution series. *Blood*. 2014;123(2):177–183.
344. Zhenchuk A, Lotfi K, Juliusson G, Albertioni F. Mechanisms of anti-cancer action and pharmacology of clofarabine. *Biochem Pharmacol*. 2009;78(11):1351–1359.
345. Schmiegelow K, Al-Modhwahy I, Andersen MK, et al. Methotrexate/6-mercaptopurine maintenance therapy influences the risk of a second malignant neoplasm after childhood acute lymphoblastic leukemia: results from the NOPHO ALL-92 study. *Blood*. 2009;113(24):6077–6084.
346. Dillman RO. Pentostatin (Nipent®) in the treatment of chronic lymphocyte leukemia and hairy cell leukemia. *Expert Rev Anticancer Ther*. 2004;4(1):27–36.
347. Munshi PN, Lubin M, Bertino JR. 6-thioguanine: a drug with unrealized potential for cancer therapy. *Oncol*. 2014;19(7):760–765.
348. Dent S, Messersmith H, Trudeau M. Gemcitabine in the management of metastatic breast cancer: a systematic review. *Breast Cancer Res Treat*. 2008;108(3):319–331.
349. Moysan E, Bastiat G, Benoit JP. Gemcitabine versus Modified Gemcitabine: a review of several promising chemical modifications. *Mol Pharm*. 2013;10(2):430–444.
350. Matt P, van Zwieten-Boot B, Calvo Rojas G, et al. The European Medicines Agency review of Tegafur/Gimeracil/Oteracil (Teysuno™) for the treatment of advanced gastric cancer when given in combination with cisplatin: summary of the Scientific Assessment of the Committee for medicinal products for human use (CHMP). *Oncol*. 2011;16(10):1451–1457.
351. Alimoghaddam K. A review of arsenic trioxide and acute promyelocytic leukemia. *Int J Hematol Oncol Stem Cell Res*. 2014;8(3):44–54.
352. Emadi A, Gore SD. Arsenic trioxide - an old drug rediscovered. *Blood Rev*. 2010;24(4–5):191–199.
353. Menzin AW, King SA, Aikins JK, Mikuta JJ, Rubin SC. Taxol (paclitaxel) was approved by FDA for the treatment of patients with recurrent ovarian cancer. *Gynecol Oncol*. 1994;54(1):103.
354. Li J, Ren J, Sun W. Systematic review of ixabepilone for treating metastatic breast cancer. *Breast Cancer*. 2017;24(2):171–179.
355. Abolmaali SS, Tamaddon AM, Dinarvand R. A review of therapeutic challenges and achievements of methotrexate delivery systems for treatment of cancer and rheumatoid arthritis. *Cancer Chemother Pharmacol*. 2013;71(5):1115–1130.
356. Moore AS, Nelson RW, Henry CJ, et al. Streptozocin for treatment of pancreatic islet cell tumors in dogs: 17 cases (1989-1999). *J Am Vet Med Assoc*. 2002;221(6):811–818.
357. Said R, Tsimberidou AM. Pharmacokinetic evaluation of vincristine for the treatment of lymphoid malignancies. *Expert Opin Drug Metabol Toxicol*. 2014;10(3):483–494.
358. Martino E, Casamassima G, Castiglione S, et al. Vinca alkaloids and analogues as anti-cancer agents: looking back, peering ahead. *Bioorg Med Chem Lett*. 2018;28(17):2816–2826.
359. Smith GA. Current status of vinorelbine for breast cancer. *Oncology (Williston Park)*. 1995;9(8):767–773; discussion 774, 776, 779.
360. Gasparini G, Caffo O, Barni S, et al. Vinorelbine is an active antiproliferative agent in pretreated advanced breast cancer patients: a phase II study. *J Clin Oncol*. 1994;12(10):2094–2101.