

FULL LENGTH ARTICLE

Mutations in EZH2 are associated with poor prognosis for patients with myeloid neoplasms

Qi Zhang^a, Qi Han^a, Jie Zi^a, Jinlong Ma^a, Huihui Song^a,
Yulu Tian^a, Mary McGrath^b, Chunhua Song^b, Zheng Ge^{a,*}

^a Department of Hematology, Zhongda Hospital Southeast University, Institute of Hematology Southeast University, Nanjing 210009, China

^b Department of Pediatrics, Pennsylvania State University College of Medicine, Hershey, PA 17033, USA

Received 14 February 2019; received in revised form 18 April 2019; accepted 7 May 2019

Available online 16 May 2019

KEYWORDS

EZH2;
Meta-analysis;
Mutations;
Myeloid neoplasms;
Prognostic

Abstract EZH2 is a component of the polycomb repressive complex 2 (PRC2), which is a highly conserved histone methyltransferase that methylates lysine 27 of histone 3. EZH2 mutations are associated with oncogenesis and progression of cancers. However, the relationship between the clinical outcome of patients with myeloid malignancies and EZH2 mutations is controversial. Therefore, we performed a meta-analysis of 8 studies ($n = 2243$ patients) that evaluates the correlation between EZH2 mutations and overall survival (OS) in patients with myeloid neoplasms. EZH2 mutations were associated with significantly worse OS (hazard ratio [HR] = 2.37, 95% confidential interval (CI), 1.48–3.79). In a word, EZH2 mutations indicate a poor prognosis for patients with myeloid neoplasms.

Copyright © 2019, Chongqing Medical University. Production and hosting by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Introduction

Enhancer of zeste homolog 2 (EZH2), a functional enzymatic component of the polycomb repressive complex 2 (PRC2), participates in histone methylation and gene silencing by posttranslational histone modifications.¹ Moreover, EZH2 is involved with cell proliferation, differentiation, invasion, as well as metastasis.² EZH2 localizes to the long arm of chromosome 7 at position 7q35.³

* Corresponding author. Zhongda Hospital Southeast University, Department of Hematology, Nanjing 210009, China. Fax: +86 25 83262471.

E-mail address: Janeg879@hotmail.com (Z. Ge).

Peer review under responsibility of Chongqing Medical University.

Emerging data have shown that EZH2 is abnormally expressed in different human malignancies including breast cancer,⁴ prostate cancer,⁵ gastric cancer,⁶ colorectal cancer⁷ and lung cancer. Nevertheless, the prognostic role of EZH2 is unknown in myeloid neoplasms.

Myeloid malignancies are defined as clonal hematopoietic disorders originating from myeloid stem/progenitor cells, including acute myeloid leukemia (AML), myelodysplastic syndromes (MDSs), myeloproliferative neoplasms (MPNs), and myelodysplastic/myeloproliferative neoplasms (MDSs/MPNs). Altering different hematopoietic cell lineages at different stages of maturation leads to various clinical and prognostic features. With the advent of next-generation sequencing (NGS) in the last several years, certain recurrently mutated genes, such as SF3B1,⁸ TP53,⁹ ASXL1¹⁰ and so on, have been revealed to provide prognostic information. Although numerous studies have demonstrated that EZH2 is tightly connected with all kinds of human malignancies, the prognostic roles of EZH2 in myeloid neoplasms are still inconclusive and unclear. The aim of this study is to explore the association between EZH2 and prognosis in patients with myeloid neoplasms.

Materials and methods

Literature search

Relevant articles from PubMed, the Cochrane Library, Embase and Medline were included in this meta-analysis. An upper date limit of Dec 01, 2018 was applied, and there was no lower date limit. The keywords included the following terms: EZH2 or Enhancer of zeste homolog 2; acute myeloid leukemia or myelodysplastic syndromes or myeloproliferative neoplasms. Subsequently, we glanced over titles and abstracts to select available studies.

Literature selection

Studies were considered eligible if they met the following criteria: (1) the diagnosis was made according to the World Health Organization (WHO) 2008 criteria or French-American-British (FAB) criteria; (2) The correlation between EZH2 mutations and clinical outcome was analyzed; (3) Detailed survival information of patients with EZH2 mutations was provided, such as overall survival (OS) with a hazard ratio (HR) and 95% confidence interval (CI); (4) The

studies were published in English. The articles which did not directly offer HR and 95%CI were also kept if we could rebuild them using the P values and other data reported. OS was measured from the date of the first sample collection to the time of death from any cause, or the time at last follow-up.

Data extraction

Data retrieved from the reports included the name of the first author, publication year, region, subtype of myeloid neoplasms, sample size, number of EZH2 mutations, age and gender distribution of patients (Table 1). HR and 95% CI for OS were also extracted from the selected articles.

Statistical methods

HR and 95%CI for OS were used to assess the association between EZH2 mutations and the survival outcomes for patients with myeloid neoplasms. If HRs and 95%CIs were not directly offered, we calculated them on the basis of the reported Kaplan-Meier, P-value, or other statistical parameters given in the text using the method proposed by Parmar *et al.*¹¹ and Hotta *et al.*¹² By convention, an observed HR > 1 indicates worse survival for the group with EZH2 mutations. The impact of EZH2 on survival was considered to be statistically significant if the 95%CI did not overlap with 1. In the figures, 95%CIs are represented with horizontal lines. Each box indicates the HR point estimate and its area is proportional to the weight of the study. The overall summary estimate is represented by the diamond, with CI represented by its width. The unbroken vertical line is set at the null value (HR = 1.0).

We assessed the heterogeneity among the studies based on the Q value and I² statistical value (25%, 50%, and 75% correspond to the cut-off points for low, moderate, and high degrees of heterogeneity, respectively). Heterogeneity was considered to be statistically significant if I² was greater than 50%; otherwise, no significant heterogeneity was observed. When the heterogeneity across studies was identified as greater than 50%, the random effects model (the DerSimonian and Laird method) was used; otherwise, the fixed-effect model (the Mantel-Haenszel method) was used. Eventually, we adopted the random-effects model. To find the causes of heterogeneity, we performed subgroup analyses according to the region, sample size, and sequencing method. In addition, we conducted a sensitivity analysis in order to assess the

Table 1 Characteristics of the included studies.

First Author	Year	Region	Subtype	Number	Sex (M/F)	Age (range)	EZH2 mutations
Grossmann V	2011	Germany	AML	81	57/24	72.8 (40–85.5)	9 (11.1%)
Saygin C	2018	America	AML	100	52/48	58.5 (24–75)	5 (5.0%)
Wang XL	2013	China	MDS	714	396/318	43 (8–83)	13 (1.8%)
Wang JY	2013	China	MDS	153	106/47	51 (16–81)	8 (5.2%)
Bejar R	2012	America	MDS	288	203/85	69 (15–90)	23 (8.0%)
Tobiasson M	2016	Sweden and UK	MDS	134	Unknown	70.5 (35–88)	12 (9.0%)
SN Khan	2013	America	MDS, MDS/MPN, MPN, AML	469	Unknown	Unknown	38 (8.0%)
Wu LY	2015	China	MDS	304	162/142	57 (11–89)	11 (3.6%)

impact of each individual study on the strength and stability of the results. Evidence of publication bias was sought using the methods of Begg et al.¹³ and Egger et al.¹⁴ In this study, all calculations were performed by Stata version 12.0 (College Station, TX, USA). A two-tailed P-value of $P < 0.05$ was considered statistically significant.

Results

Characteristics of the selected studies

Eight studies published between 2011 and 2018, covering a total of 2243 patients were included in this meta-analysis (Fig. 1, Table 1). The main characteristics of the studies and patients are summarized in Table 1. Detailed information of the patients in the study published by Khan²¹ are not available. The sample size ranged from 81 to 714, and the frequency of EZH2 mutated cases from eight studies varied between 1.8% and 11.1%. Among the 2243 patients, 1040 patients had MDS, 942 had AML, 195 had MDS/MPN, and 66 had MPN. EZH2 mutations were frequently observed in MDS (2.59%), AML (0.94%) and MDS/MPN (0.80%), but were rare in the patients with MPN (0.09%). In total, 119 of 2243 patients were found to harbor EZH2 mutations.

Prognostic influence of EZH2 mutations in patients with myeloid neoplasms

After evaluating all available studies, the overall HR for the OS was 2.37 (95%CI 1.48–3.79), revealing that EZH2 mutations are associated with poor prognosis for myeloid neoplasms (Fig. 2). Furthermore, subgroup analysis was adopted in order to find the causes of heterogeneity in the analysis of OS. A subgroup analysis for region revealed the pooled HR for North America was 3.26 (95%CI 2.32–4.58, $I^2 = 0.0\%$, $P < 0.001$). However, no significant relationship was found for Europe or Asia (HR = 1.53, 95%CI 0.33–7.16, $I^2 = 84.3\%$, $P = 0.592$; HR = 2.19, 95%CI 0.82–5.85, $I^2 = 76.5\%$, $P = 0.117$, respectively) (Fig. 3). The pooled HR for sample size less than 200 was 2.52 (95%CI 0.94–6.75, $I^2 = 77.6\%$, $P = 0.066$), whereas the HR when more than 200 cases were enrolled in studies was 2.37 (95%CI 1.40–3.99, $I^2 = 62.4\%$, $P = 0.001$). For studies evaluating OS by using different methods for EZH2 mutations detection, the results indicate the pooled HR for Sanger

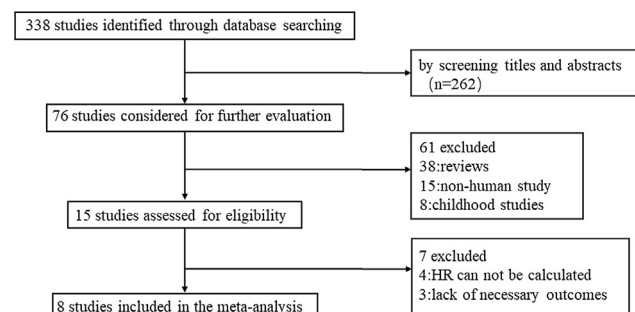


Figure 1 Flowchart of study search and selection strategy.

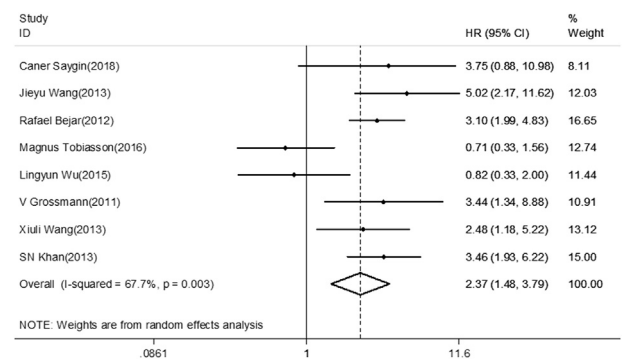


Figure 2 Forest plots of pooled HRs and 95% CI of association between EZH2 mutations and overall survival in the entire cohort of patients with myeloid neoplasms. The size of the blocks or diamonds represent the weight of the random-effect model in the meta-analysis.

sequence was 3.07 (95%CI 2.23–4.23, $I^2 = 0.0\%$, $P < 0.001$). There was no statistically significant association observed between detection by next-generation sequence (NGS) and OS (HR = 2.00, 95%CI 0.85–4.72, $I^2 = 76.9\%$, $P = 0.114$) (Fig. 3).

Sensitivity test and publication bias

A sensitivity analysis was conducted by calculating pooled HR again when omitting one study each time. As shown in Fig. 4, there is no obvious difference when any study was omitted, which implies that each individual study did not affect the stability of the association between EZH2 mutations and OS in patients with myeloid neoplasms.

The funnel plot and Egger's test were performed to detect the existence of publication bias, and the resulting figure indicates asymmetry (Fig. 5). Nevertheless, the P value for Egger's test was 0.595, which demonstrates that there is no publication bias.

Discussion

Polycomb group (PcG) proteins have two main families that are polycomb repressive complexes 1(PRC1), and PRC2.¹ The human PRC2 complex is composed of EZH2, EED, SUZ12, RbAp46/48 and AEBP2 (Fig. 6A).²⁴ The EZH2 gene is a catalytic subunit of the PRC2, and its C-terminal SET domain has methyltransferase activity (Fig. 6B). EZH2 spans about 40 kb and is composed of 20 exons.³ Furthermore, EZH2 has been reported to harbor mutations in some exons (Fig. 6C).^{25–29} EZH2 plays an important role in oncogenesis and cancer progression by epigenetic gene silencing and chromatin remodeling.⁵ Moreover, cancer progression may be affected by EZH2 mutations. For instance, the mutation of tyrosine 641(Y641) with the C-terminal catalytic SET domain of EZH2 increases the level of trimethylated H3K27 (H3K27me3) and thus represses the expression of Polycomb targets.²³ Although the prognostic function of EZH2 mutations in myeloid neoplasms have been assessed by several studies, the results are disputable.^{15–19} Therefore, we performed a meta-analysis of published studies to explore

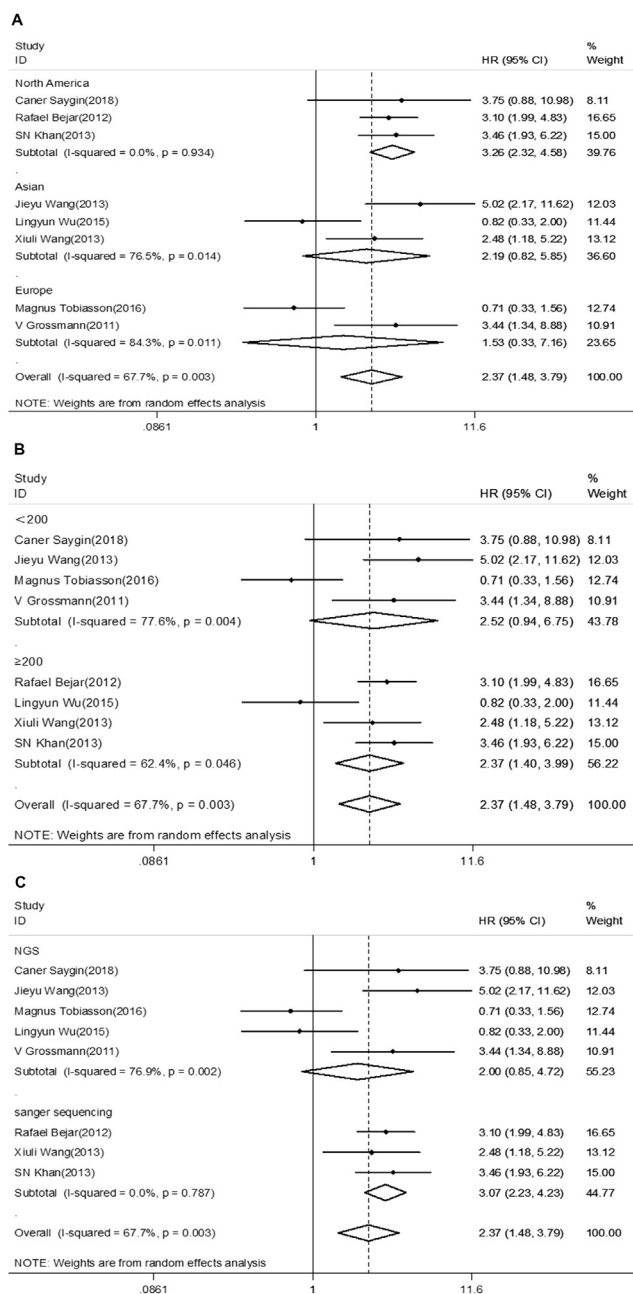


Figure 3 Forest plot of a different region, sample size, and mutation detection method included in the subgroup analysis for OS. (A) Subgroup analysis of OS by region. (B) Subgroup analysis of OS by sample size. (C) Subgroup analysis of OS by the mutation detection method.

the prognostic impact of EZH2 mutations in patients with myeloid neoplasms. This is the first time that a review of relevant literature has been performed to evaluate the correlation between EZH2 mutations and overall survival in patients with myeloid neoplasms.

Our meta-analysis combined 8 different publications, and included 2243 patients with myeloid malignancies. Combined hazard ratios suggest that the HR for the OS was 2.37 (95%CI = 1.48–3.79), which indicates that patients carrying an EZH2 mutation have shorter OS in myeloid

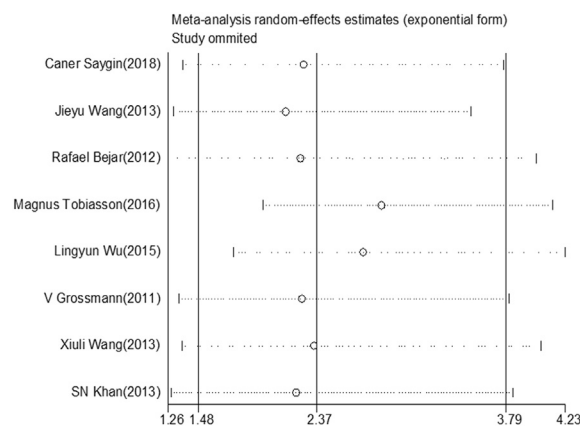


Figure 4 Sensitivity analysis. The middle vertical axis represents the pooled HR, and the two vertical axes indicate the corresponding 95% CI. Each hollow circle represents the pooled HR when the left study was omitted in this meta-analysis, and the two ends of every broken line indicate the 95% CI.

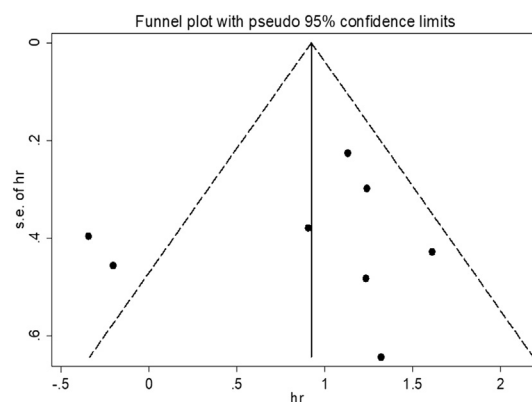


Figure 5 Funnel plot for publication bias in terms of the association of EZH2 mutation with OS.

neoplasms. These results were consistent with most studies previously done.^{15,16,19–22} Although the heterogeneity was large ($I^2 = 67.6\%$), the sensitivity analysis demonstrates that the results of this meta-analysis were stable and reliable. Subgroup analyses showed that North American cohorts (HR = 3.26, 95%CI 2.32–4.58), Sanger sequencing (HR = 3.07, 95%CI 2.23–4.23), and sample size over 200 individuals (HR = 2.37, 95%CI 1.40–3.99) were significantly associated with OS, suggesting the influence of ethnic variations on EZH2 mutations.

Our meta-analysis has its own limitations. First, it is inevitable that relevant studies are left out, in spite of using a comprehensive search strategy. In particular studies published in languages other than English have been omitted. In addition, the first-hand data of a few studies could not be acquired, and they therefore were not included in the analysis. Second, all the enrolled studies were retrospective studies rather than prospective studies. Finally, this analysis covers only a small subset of patients with myeloid neoplasms.

In conclusion, this meta-analysis showed that EZH2 mutations have significant influence in OS for myeloid neoplasms, which suggests that EZH2 mutations may be useful

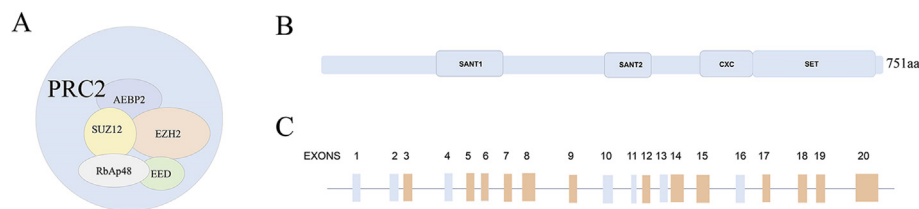


Figure 6 Schematic representation of Polycomb complex PRC2, EZH2 and distribution of EZH2 mutations. (A) The five core subunits of human PRC2. (B) The domain architecture of human EZH2. (C) Localization of EZH2 mutations. Orange bars correspond to mutations that have been reported before.

in predicting prognosis and guiding treatment in clinical practice. At the same time, prospective randomized controlled studies with large numbers and different types of myeloid neoplasms are needed.

Conflicts of interest

All authors have none to declare.

Acknowledgments

This work is supported in part by The National Natural Science Foundation of China, China (81770172, 81270613); Key Research & Technology Projects in Jiangsu Province, China (BE2017747); Milstein Medical Asian American Partnership (MMAAP) Foundation Research Project Award, USA in Hematology (2017).

References

- Margueron R, Reinberg D. The Polycomb complex PRC2 and its mark in life. *Nature*. 2011;469(7330):343–349.
- Lu C, Han HD, Mangala LS, et al. Regulation of tumor angiogenesis by EZH2. *Cancer Cell*. 2010;18(2):185–197.
- Cardoso C, Mignon C, Hetet G, Grandchamps B, Fontes M, Colleaux L. The human EZH2 gene: genomic organisation and revised mapping in 7q35 within the critical region for malignant myeloid disorders. *Eur J Hum Genet*. 2000;8(3):174–180.
- Kleer CG, Cao Q, Varambally S, et al. EZH2 is a marker of aggressive breast cancer and promotes neoplastic transformation of breast epithelial cells. *Proc Natl Acad Sci USA*. 2003;100(20):11606–11611.
- Varambally S, Dhanasekaran SM, Zhou M, et al. The polycomb group protein EZH2 is involved in progression of prostate cancer. *Nature*. 2002;419(6907):624–629.
- Matsukawa Y, Semba S, Kato H, Ito A, Yanagihara K, Yokozaki H. Expression of the enhancer of zeste homolog 2 is correlated with poor prognosis in human gastric cancer. *Cancer Sci*. 2006;97(6):484–491.
- Wang CG, Ye YJ, Yuan J, Liu FF, Zhang H, Wang S. EZH2 and STAT6 expression profiles are correlated with colorectal cancer stage and prognosis. *World J Gastroenterol*. 2010;16(19):2421–2427.
- Yoshida K, Sanada M, Shiraishi Y, et al. Frequent pathway mutations of splicing machinery in myelodysplasia. *Nature*. 2011;478(7367):64–69.
- Rücker FG, Schlenk RF, Bullinger L, et al. TP53 alterations in acute myeloid leukemia with complex karyotype correlate with specific copy number alterations, monosomal karyotype, and dismal outcome. *Blood*. 2012;119(9):2114–2121.
- Metzeler KH, Becker H, Maharry K, et al. ASXL1 mutations identify a high-risk subgroup of older patients with primary cytogenetically normal AML within the ELN Favorable genetic category. *Blood*. 2011;118(26):6920–6929.
- Parmar MK, Torri V, Stewart L. Extracting summary statistics to perform meta-analyses of the published literature for survival endpoints. *Stat Med*. 1998;17(24):2815–2834.
- Hotta K, Matsuo K, Ueoka H, Kiura K, Tabata M, Tanimoto M. Meta-analysis of randomized clinical trials comparing cisplatin to carboplatin in patients with advanced non-small-cell lung cancer. *J Clin Oncol*. 2004;22(19):3852–3859.
- Begg CB, Mazumdar M. Operating characteristics of a rank correlation test for publication bias. *Biometrics*. 1994;50(4):1088–1101.
- Egger M, Davey SG, Schneider M, Minder C. Bias in meta-analysis detected by a simple, graphical test. *BMJ*. 1997;315(7109):629–634.
- Grossmann V, Kohlmann A, Eder C, et al. Molecular profiling of chronic myelomonocytic leukemia reveals diverse mutations in >80% of patients with TET2 and EZH2 being of high prognostic relevance. *Leukemia*. 2011;25(5):877–879.
- Saygin C, Hirsch C, Przychodzen B, et al. Mutations in DNMT3A, U2AF1, and EZH2 identify intermediate-risk acute myeloid leukemia patients with poor outcome after CR1. *Blood Canc J*. 2018;8(1).
- Wang X, Dai H, Wang Q, et al. EZH2 mutations are related to low blast percentage in bone marrow and -7/del(7q) in de novo acute myeloid leukemia. *PLoS One*. 2013;8(4):e61341.
- Wang J, Ai X, Gale RP, et al. TET2, ASXL1 and EZH2 mutations in Chinese with myelodysplastic syndromes. *Leuk Res*. 2013;37(3):305–311.
- Bejar R, Stevenson KE, Caghey BA, et al. Validation of a prognostic model and the impact of mutations in patients with lower-risk myelodysplastic syndromes. *J Clin Oncol*. 2012;30(27):3376–3382.
- Tobiasson M, McLornan DP, Karimi M, et al. Mutations in histone modulators are associated with prolonged survival during azacitidine therapy. *Oncotarget*. 2016;7(16):22103–22115.
- Khan SN, Jankowska AM, Mahfouz R, et al. Multiple mechanisms deregulate EZH2 and histone H3 lysine 27 epigenetic changes in myeloid malignancies. *Leukemia*. 2013;27(6):1301–1309.
- Wu L, Song L, Xu L, et al. Genetic landscape of recurrent ASXL1, U2AF1, SF3B1, SRSF2, and EZH2 mutations in 304 Chinese patients with myelodysplastic syndromes. *Tumor Biol*. 2016;37(4):4633–4640.
- Chang WS, Liao CH, Tsai CW, et al. Association of enhancer of zeste 2 (EZH2) genotypes with bladder cancer risk in Taiwan. *Anticancer Res*. 2016;36(9):4509–4514.
- Tan JZ, Yan Y, Wang XX, Jiang Y, Xu HE. EZH2: biology, disease, and structure-based drug discovery. *Acta Pharmacol Sin*. 2014;35(2):161–174.
- Morin RD, Johnson NA, Severson TM, et al. Somatic mutations altering EZH2 (Tyr641) in follicular and diffuse large B-cell

- lymphomas of germinal-center origin. *Nat Genet.* 2010;42(2):181–185.
26. Ernst T, Pflug A, Rinke J, et al. A somatic EZH2 mutation in childhood acute myeloid leukemia. *Leukemia.* 2012;26(7):1701–1703.
27. Grossmann V, Bacher U, Kohlmann A, et al. EZH2 mutations and their association with PICALM-MLLT10 positive acute leukemia. *Br J Haematol.* 2012;157(3):387–390.
28. Guglielmelli P, Biamonte F, Score J, et al. EZH2 mutational status predicts poor survival in myelofibrosis. *Blood.* 2011;118(19):5227–5234.
29. Nikoloski G, Langemeijer SMC, Kuiper RP, et al. Somatic mutations of the histone methyltransferase gene EZH2 in myelodysplastic syndromes. *Nat Genet.* 2010;42(8):665–667.