

CHANGES OF CATHEPSIN D AND α_2 -MACROGLOBULIN IN RATS AFTER ACUTE TOTAL BODY IRRADIATION*

Jin Weiqiao (金为翘), Chen Shijie (陈识杰), Mo Suzhen (莫素珍),

Fan Peifang (范佩芳), Le Min (乐敏), Zhang Hao, (张皓),

Li Yonghe (李永河), and He Jiewei (何介薇)

(Institute of Radiation Medicine, Shanghai Medical University, Shanghai 200032, China)

(Received February 1989)

ABSTRACT

α_2 -macroglobulin (α_2 M) could stimulate the regeneration of thymic and bone marrow cells in rats received γ -irradiation, but there was very few reports concerning its mechanism. Wistar rats were irradiated by ^{16}Co at 7 Gy, 8.5 Gy, 15 Gy total body doses. Blood plasma and some tissue's extracts were collected α_2 M level. α M activity and cathepsin D activity, malonaldehyde level were determined by radioimmunoassay, modified Schidlow's method, Barrett's method and Ohkawa's method respectively.

Key words: Alpha-2-macroglobulin Cathepsin D Malonaldehyde
Radiation Rat

1. INTRODUCTION

There was increase of plasma α_2 M levels as well as plasma α M activities after 7 Gy, 8.5 Gy, 15 Gy irradiation. Both of them were in parallel with the increase of radiation doses. The changes of plasma malonaldehyde also correlated with the elevation of plasma α_2 M concentration. There was marked elevation of cathepsin D activities in splenic tissue but no increase of α_2 M level in spleen in the first 24 h, after 8.5 Gy exposure. It seems that the ability of α_2 M synthesis by splenic lymphatic cells was reduced during severe radiation injuries.

Burger et al. found that sheep splenic extracts and human serum α_2 M could stimulate the regeneration of thymic and bone marrow cells in rats received γ -irradiation^[1]. In 1974 I. Berenblum et al. further proved that α_2 M, just like splenic extracts, could reduce the incidence of lymphatic leukemia in irradiated rats^[2], but there was very few reports concerning its mechanism.

α_2 M is a protease inhibitor of wide spectrum. It regulates the level of intracellular and extracellular proteolytic enzymes^[3]. Recently it was reported that combined with protease α_2 M was a free radical scavenger as well^[4].

During irradiation there is an increase of free radical flux in the organism. Free

* The Project Supported by National Natural Science Foundation of China

radicals accelerated proteolysis of tissue proteins^[5]. And proteases were involved in the production of superoxide radical in polymorphonuclear neutrophils too^[6], so it was enhanced the radiation injury, and was speculated that the beneficial effects of $\alpha_2\text{M}$ might be both binding with proteases and scavenging of free radicals.

Cathepsin D, being one of the important proteolytic enzymes in lysosomes, presents widely in mammalian cells with tissue proteolytic effects, for which there are very few natural inhibitors^[7]. Normally, cathepsin D could be trapped by $\alpha_2\text{M}$ in $\text{pH} \geq 6$ ^[8]. In this study changes of cathepsin D, $\alpha_2\text{M}$ and malonaldehyde (MDA) (one kind of lipid peroxidation products in free radical chain reaction) were investigated in irradiated rats.

II. MATERIALS AND METHODS

240 male Wistar rats (Shanghai laboratory animal permission. No. 32* — 34*) with body weight 211 ± 31 g ($\bar{X} \pm \text{SD}$) were divided into several groups with 5—8 rats each. The total body irradiated groups received 7 Gy, 8.5 Gy and 15 Gy separately from ^{60}Co source with dose rates 0.03483—0.04257 C/(kg.min). The distance between ^{60}Co source and rats was 80cm. Blood and tissue samples from each group were collected at 1/4, 1, 3, 7 days after irradiation, some even longer. Plasma was obtained from heparinized blood after centrifugation. Tissue samples were obtained after bleeding and irrigating with 0.9% NaCl solution through abdominal vessels. Then it was homogenized and centrifuged, and supernatants were used for examination.

By using bovine hemoglobin as the substrate the activities of cathepsin D were determined by Barrett's method^[9].

Rat's acute phase plasma $\alpha_2\text{M}$ was purified through Sephacryl S-300 gel filtration and affinity chromatography coupled with anti-normal rat's plasma. Immunopure $\alpha_2\text{M}$ showed only one band in polyacrylamide gel electrophoresis was obtained^[10]. Anti- $\alpha_2\text{M}$ antiserum was collected by immunizing rabbits as usual.

Plasma concentrations of $\alpha_2\text{M}$ were determined by rocket electrophoresis^[11]. Estimation of $\alpha_2\text{M}$ contents in liver, spleen, kidney, parotid gland and skin tissues was done by radioimmunoassay^[12]. Changes of $\alpha_2\text{M}$ concentration in rats after irradiation varied remarkably individually, so statistic analysis on logarithm was performed.

Plasma $\alpha_2\text{M}$ activities were tested by modified Schidlow's method using BAPNA as its substrate^[13].

Plasma concentration of MDA was measured according to Leu Shizhong's method by using thiobarbituric acid as chief reagent^[14]. MDA in tissues was estimated by modified Ohkawa's method^[15] and protein concentration in tissue's extracts by Lowry's method^[16].

III. RESULTS

Total body irradiated rats with 7 Gy were all alive at the 28th day (8/8). 12/16 irradiated rats with 8.5 Gy were still living 7 days after exposure. 4/8 15 Gy irradiated rats were alive at the 3rd day.

There was no significant changes of plasma cathepsin D activities in the first few days after 7 Gy, 8.5 Gy and 15 Gy doses of ^{60}Co irradiation. But a tendency of increase in cathepsin D activities was observed in spleen 7 days after 7 Gy exposure (irradiated: control = $40.8 \pm 10.2^*$: 34.5 ± 5.8), and a pronounced rise was observed 6 h after 8.5 Gy irradiation and persisted even more than 7 days in spleen (Tab.1). An increase of cathepsin D activities was also detected in the liver at 7th day after 8.5 Gy irradiation (Tab.1).

Table 1

Cathepsin D activities in plasma (mu/ml) and tissue extracts (mu/mg protein) of rats after 8.5 Gy ^{60}Co total body irradiation (TBI) ($X \pm SD$) ($n = 8$)

Groups		Plasma	Spleen	Kidney	Liver	Skin
Control		22.5 ± 3.0	31.7 ± 3.6	15.5 ± 8.4	5.7 ± 1.4	27.4 ± 7.8
	1/4d	23.4 ± 2.8	53.7 ± 9.6* * *	9.5 ± 2.4	4.9 ± 1.9	21.9 ± 3.8
Postirrad.	1d	21.8 ± 0.9	48.7 ± 3.9* * *	17.2 ± 3.8	3.6 ± 0.7	27.6 ± 4.7
	3d	19.7 ± 3.0	49.4 ± 11.2* * *	18.9 ± 4.6	6.4 ± 1.8	35.9 ± 7.6
	7d	21.6 ± 6.9	53.0 ± 7.4* * *	13.8 ± 2.9	11.5 ± 3.6* * *	31.6 ± 5.5

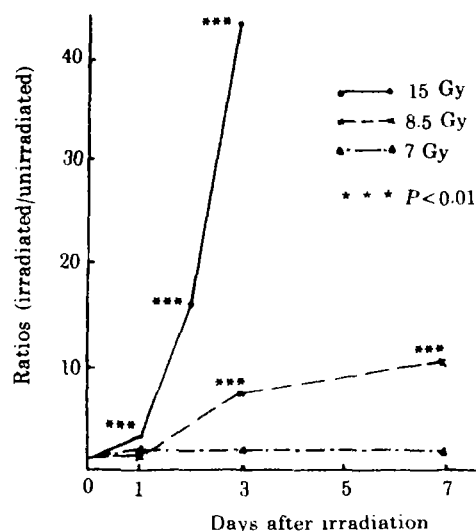


Fig. 1 Changes of plasma α_2 M levels in rats after irradiation with different doses

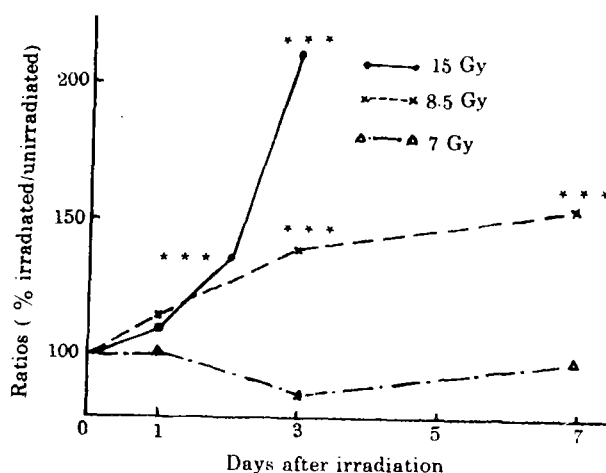


Fig. 2 Changes of plasma α M activities in rats after irradiation with different doses

Plasma α_2M levels increased 28 days after 7 Gy irradiation (irradiated: control = $66.5 \pm 2.8^{***}$: 25.0 ± 1.3), but without much changes in αM activities. Both plasma α_2M levels and activities were increased evidently 3 days after 8.5 Gy exposure and 1 day after 15 Gy irradiation (Fig.1, 2). 3 days after exposure both plasma α_2M contents and activities increased in parallel with increases of radiation doses ($r=0.6739$, $P=0.003$ & $r=0.8566$, $P=0$ respectively). And there was correlation between plasma α_2M levels and αM activities too ($r=0.7535$, $P=0.0005$).

In 8.5 Gy irradiated rats, α_2M levels in liver, kidney, parotid gland, skin and spleen rose and correlated with plasma α_2M levels ($P=0$, $P=0$, $P=0$, $P=0$ & $P=0.0019$ respectively). But there was no increase of α_2M in the spleen in the first 24h and less increase of α_2M in spleen than other tissues in 7 days after irradiation (Tab. 2).

Table 2

α_2M levels in tissue extracts (ng/mg protein) in rats after 8.5 Gy ^{60}Co TBI

[geometric mean $\sqrt{(\log X) \pm (\log X)}$]

Sample	Control	Post - irradiation			
	(n=8)	1/4d (n=7)	1d (n=8)	3d (n=7)	7d (n=7)
Spleen	12.59 1.1 ± 0.06	11.75 1.07 ± 0.08	10.96 1.04 ± 0.08	52.48 $1.72 \pm 0.20^{***}$	81.28 $1.91 \pm 0.13^{***}$
Skin	33.88 ⁺ 1.53 ± 0.05	41.69 1.62 ± 0.05	53.70 ⁺ 1.73 ± 0.03	104.71 2.02 ± 0.21	380.19 $2.58 \pm 0.19^{***}$
Parotid gland	9.12 0.96 ± 0.06	13.80 1.14 ± 0.06	13.49 1.13 ± 0.03	40.74 $1.61 \pm 0.19^{**}$	102.33 $2.01 \pm 0.25^{***}$
Kidney	5.25 0.72 ± 0.06	10.23 [#] 1.01 ± 0.11	8.71 0.94 ± 0.07	56.23 ⁺ $1.75 \pm 0.06^{***}$	77.62 $1.89 \pm 0.26^{***}$
Liver	1.32 0.12 ± 0.13	4.90 0.69 ± 0.16	1.70 0.23 ± 0.11	15.14 $1.18 \pm 0.25^{***}$	33.88 $1.53 \pm 0.29^{***}$

$^{**} P < 0.05$ $^{***} P < 0.01$ $^{+} n = 7$ $^{#} n = 8$ $^{+} n = 6$

The increase of plasma αM activities was positively correlated with increase of cathepsin D activities in spleen ($r=0.4285$, $P=0.0065$).

Plasma concentration of MDA increased on the 3rd and 28th day after 7 Gy radiation, they were $7.71 \pm 0.23^{***}$ and $7.18 \pm 1.71^{**}$ nmol/ml respectively with 5.77 ± 1.06 nmol/ml in control groups. MDA concentration in plasma, liver, kidney and testis of 8.5 Gy irradiated rats all increased one day after exposure, and still higher than control in plasma, kidney and testis 7 days after exposure (Tab.3). There was correlation between plasma MDA and plasma α_2M level with $r=0.7077$ and $P=0$. In 15 Gy group, plasma levels of MDA increased 1,2 and 3 days after irradiation ($9.2 \pm 0.7^{***}$, $9.3 \pm 1.4^{**}$, 26.8 respectively with control 7.8 ± 0.8 nmol/ml), and it correlated with the increase of α_2M contents in plasma too ($r=0.7676$, $P=0$).

Table 3
MDA in plasma (nmol/ml) and tissue extracts (n mol/mg protein) of rats
after 8.5 Gy ^{60}Co TBI ($\bar{X} \pm \text{SE}$) (n=8)

Sample	Control	Post- irradiation		
		1d	3d	7d
Liver	339.6 \pm 33.8	457.3 \pm 38.8* *	370.3 \pm 19.5	306.0 \pm 37.4
Kidney	97.1 \pm 7.2	130.5 \pm 9.0* * *	217.5 \pm 21.1* * *	146.2 \pm 10.8* * *
Testis	127.4 \pm 7.3	210.3 \pm 31.9* *	152.9 \pm 28.6	282.6 \pm 12.3* * *
Submax. gland	65.0 \pm 6.2	67.6 \pm 4.3	89.5 \pm 19.4	51.0 \pm 6.2
Plasma	6.81 \pm 0.17	7.98 \pm 0.18* * *	8.62 \pm 0.65	7.73 \pm 0.37* *

IV. DISCUSSION

It is well known that there is an increase of free radical flux in organism after high dose ionizing radiation. Free radicals induces lipid peroxidation with increase levels of MDA in bone marrow, spleen, kidney, plasma etc^[17].

Hectepchk reported that in rats 3 days after 12 Gy ^{60}Co TBI the cathepsin D activities increased 3—5 times in small intestinal extracts than control group^[18]. Our observation revealed similar findings as the MDA increased in plasma, liver, kidney, testis and cathepsin D activities also increased in spleen in 8.5 Gy ^{60}Co irradiated rats.

Weimer et al.^[19] found that 7 days after 0.0516, 0.1032, 0.1548 and 0.2064 C/kg ^{60}Co irradiation α_2 M appeared in rat's plasma, and was even detected 24h after 800 r irradiation. Rat's α_2 M is one kind of acute phase proteins. Konnova et al.^[20] considered that rat plasma α_2 M activities reduced to almost 58% in the first 3 days after 6 Gy exposure. Our results showed there were increases both of α_2 M levels and α M activities with increase of radiation dosage. It seemed that rat's plasma α_2 M was a kind of acute phase proteins, it increased significantly after radiation stimulation to compensate the requirement in microenvironment for binding of proteases, such as cathepsin D etc, and scavenging free radicals.

α_2 M is synthesized by liver parenchymal cells, Kupffer's cells and other reticulo-endothelial cell system as well as fibroblasts^[21]. In acute phase there is increase of α_2 M synthesis by lymphatic cells too^[22]. In our study after 8.5 Gy exposure the α_2 M level in spleen increased gradually and less than other tissues. It seemed that the ability of α_2 M synthesis by splenic lymphatic cells was reduced with severe radiation injuries.

V. SUMMARY

There was marked elevation of cathepsin D activities in splenic tissues and no

increase of α_2 M levels in spleen in the first 24h after 8.5 Gy TBI by ^{60}Co in male Wistar rats. The increase of splenic cathepsin D activities persisted even 7 days after exposure, while splenic α_2 M levels increased less than other tissues (liver, kidney, skin etc).

There was increase of plasma α_2 M levels as well as α M activities after 7 Gy, 8.5 Gy and 15 Gy irradiation. Both of them were in parallel with the increase of radiation doses.

There was increase of MDA levels in plasma, liver, kidney, testis etc. The changes of plasma MDA also correlated with the elevation of plasma α_2 M concentration.

REFERENCES

- [1] M. Burger et al., *Radiat Res.*, **40** (1969), 193.
- [2] I. Berenblum et al., *Radiat Res.*, **60** (1974), 501.
- [3] D.H.Bing ed., *The chemistry and physiology of the human plasma proteins*, Pergamon, N. Y, 1979, p.385.
- [4] M.Hoffman et al., *Biochem. Biophys. Acta*, **760** (1983), 421.
- [5] R.T.Dean et al., *Biochem. Biophys. Res. Commun.*, **126** (1985), 3:1082.
- [6] S.Kitagawa et al., *FEBS Letter*, **99** (1979), 275.
- [7] V.Kostka ed., *Aspartic proteinases and their inhibitors*, Walter de Gruyter, Berlin, 1985, p.1.
- [8] A.J.Barrett, *Proteases in mammalian cells and tissues*, Amsterdam, North- Holland, 1977, p.226.
- [9] J.T.Dingle ed. *Lysosomes—a laboratory hand book*, 2nd ed, Amsterdam, North- Holland, 1977, p.124.
- [10] Mo Suzhen et al., *Acta Academiae Medicinae Shanghai*, 1989, 16 (in press).
- [11] Mo Suzhen et al., *Chinese J. Radiol Med. & Prot.*, **8** (1988), 4:233.
- [12] Li Yonghe et al., *Nuclear Techniques* (in press).
- [13] D.A.Schidlow et al., *Amer. Rev. Respira Dis.*, **121** (1980), 1:31.
- [14] Leu Shizhong et al., *Acta. Academiae Medicinae Pharmaceutics Collegiate Guangdong*, **1** (1985), 18.
- [15] H.Ohkawa et al., *Anal. Biochem.*, **95** (1979), 2:351.
- [16] O.H.Lowry et al., *J. Biol. Chem.*, **193** (1951), 1:265.
- [17] J.F.Kergonou et al., *Biochimie*, **63** (1981), 6:555.
- [18] B.C.HectepeHko et al., *Radiobiologiya*, **23** (1983), 3:379.
- [19] H.E.Weimer et al., *Amer. J. Physiology*, **209** (1965), 736.
- [20] L.A.Konnova et al., *Patol. fiziol. eksp. ter.*, **5** (1984), 17.
- [21] D.F.Mosher, *Annals of the N.Y. academy of sciences*, **421** (1983), 327.
- [22] D.E.Panrucker, *Annals of the N.Y. academy of sciences*, **417** (1983), 117.