# <sup>3</sup>H-LABELLING OF BELLADONNA ALKALOIDS BY CATALYSED EXCHANGE WITH MICROWAVE EXCITATION\*

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#### ABSTRACT

The <sup>3</sup>H-labelled belladonna alkaloids obtained by catalysed exchange method with microwave excitation was investigated. The specific activities of the labelled products were 16— 32 TBq/mol. More than 90% labelled positions of these <sup>3</sup>H-tracers were on phenyl rings. The radiochemical purity and chemical purity of crude products were both in 75— 80%.

Keywords: ${}^{3}H-NMR$ Microwavecatalysedexchange ${}^{3}H-$  anisodaminehydrobromide ${}^{3}H-$  anisodinehydrobromide ${}^{3}H-$  scopolaminehydrobromide ${}^{3}H-$  labelling

## **1** INTRODUCTION

In recent times, numerous tritiated alkaloids are used as radioactive ligands in studies of the neurotransmitter of the central nervous system<sup>[1-2]</sup>. However, belladonna alkaloids are the heat or light-sensitive compounds which cannot stand the vigorous experiment conditions. This work exhibits the preparation of belladonna alkaloids, such as <sup>3</sup>H- anisodine hydrobromide, <sup>3</sup>H- anisodamine hydrobromide, <sup>3</sup>H- scopolamine hydrobromide and <sup>3</sup>H- atropine sulfate by catalysed exchange method combined with microwave excitation. The specific activity of the tracers obtained by this method was 16— 32 TBq/mol, which are three orders higher in comparison with the results from only microwave excitation or only catalysed exchange<sup>[3]</sup>.

The following scheme outlines the process of this labelling:

- a. A molecule of belladonna alkaloids <u>Pt</u> Pt.A\* unstable ligand
- b. Tritium water Microwave excitation <sup>3</sup>H·+ OH·
- c.  ${}^{\circ}H + OH + Pt.A^*$   ${}^{\circ}H {}^{\circ}H + exchange$   $Pt + H_2O + {}^{\circ}H A^*$

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The labelled positions of  ${}^{3}\text{H}$ -belladonna alkaloids determined by  ${}^{3}\text{H}$ -NMR were mostly on the phenyl rings of the molecules.  ${}^{3}\text{H}$ -NMR signal integral area comparison on  ${}^{3}\text{H}$ -NMR spectrum exhibited the radiochemical purity was 75— 80%, and  ${}^{1}\text{H}$ -NMR peaks integral area comparison with the impurity peaks on  ${}^{1}\text{H}$ -NMR spectrum showed the chemical purity of the labelled compounds was 75— 80% <sup>[4]</sup>.

## 2 EXPERIMENTAL

## 2.1 Preparation of <sup>3</sup>H – labelled belladonna alkaloids

The samples under investigation were anisodine hydrobromide, anisodamine hydrobromide, scopolamine hydrobromide and atropine sulfate. Their purities were checked by  ${}^{1}H-NMR$  spectroscopy.

Isotope exchange procedure: The materials to be labelled (15 mg) together with a known amount of catalyst (40— 50 mg) that had been freshly reduced with NaBH<sub>4</sub> (400 — 500 mg) and 50  $\mu$ l of HTO (0.93 TBq/ml) were placed in a narrow tube, which was then frozen (liquid N<sub>2</sub>) and evacuated, after that, the tube was sealed, then the tube was put into the microwave oven (2450 MHz) for 4 min. After being cooled, the tube was opened and the content was mixed with 1 ml of methanol; the catalyst was filtered off and the solution was washed with 1 ml methanol two times for removing any possible labile tritium, after drying over IR lamp, the substrate was right the <sup>3</sup>H-labelled belladonna tracers.

## 2.2 <sup>1</sup>H- NMR and <sup>3</sup>H- NMR experimental conditions

Triton and proton nuclear magnetic resonance spectra were obtained using a JEOL FX-100 spectrometer,  ${}^{3}$ H- NMR spectra (with  ${}^{1}$ H decoupling) were recorded at 106.19 MHz and  ${}^{1}$ H spectra were recorded at 99.55 MHz. Each sample prepared by above method was redissolved in deuteriated methanol (to provide for field frequency locking) and a trace of tetramethyl silicane (TMS) was added for  ${}^{1}$ H reference. The solution to be determined by  ${}^{1}$ H- and  ${}^{3}$ H- NMR were sealed in a cylindrical microcelles (~40 µl) under lyophilization (liquid N<sub>2</sub>) and inserted into standard NMR tubes (5mm) which were capped and mounted in a spinner adapter for a 5 mm probe.  ${}^{1}$ H- NMR parameters: rf 99.55 MHz, spectral width 1000 Hz, obset 51.95 kHz, pulse width 31.2 µs (90°), dual pulse order; pulse delay 5 s, pulse interval 1.8 s, data points 8 k.  ${}^{3}$ H- NMR parameters: rf 106.19 MHz, spectral width 1060 Hz, obset 49.40 kHz, pulse width 5 s (5°), pulse interval 1 s, data points 4 k.

## **3 RESULTS AND DISCUSSION**

Specific activity obtained by the three methods and  ${}^{3}H$ -NMR results of our products are listed in Table 1. The chemical shift and distribution of labelling on phenyl ring for some  ${}^{3}H$ - belladonna alkaloids are listed in Table 2. The structure

#### formulas of belladonna alkaloids are showed in Fig.1.

#### Table 1

## Specific activity and <sup>3</sup>H NMR analysis of <sup>3</sup>H – belladonna alkaloids

Compound	This method	Assignment	Chemical shift	'Η (° <sub>o</sub> )	Catalysed method	Microwave method
	(TBq/mol)		(ppm)		(GBq/mol)	(GBq/mol)
<sup>3</sup> H– Anisodine hydrobromide	16.0	phenyl	7.27-7.36	75	10.8	<1
<sup>9</sup> H- Anisodamine hydrobromide	18.4	phenyl	7.20- 7.26	75	3.3	<1
'H– Scopolamine hydrobromide	17.5	phenyl	7.17— 7.37	75	3.4	<1
'H– Atropine sulfate	32.8	phenyl	7.23 7.30	80	0.8	< 1

Table 2

#### Chemical shifts and distribution of labelling on phenyl ring for some ${}^{3}H$ – belladonna alkaloids

Compound	Chemical shift (ppm)	Assignment	Distribution of tritium (° <sub>0</sub> )
	7.27	Ortho	20
'H–Anisodine hydrobromide	7.29	Para	20
	7.36	Meta	60
	7.20	Ortho	32
ð <sup>a</sup> H–Anisodamine hydrobromide	7.24	Para	-48
	7.26	Meta	20
	7.17	Ortho	26
'H-Scopolamine hydrobromide	7.25	Para	51
	7.31	Meta	23
	7.23	Ortho	37
'H– Atropine sulfate	7.28	Para	33
	7.30	Meta	30

<sup>3</sup>H-NMR analyses: the solutions, which usually contained 0.185— 1.11GBq of radioactivity, were separately loaded into about 3 mm diameter combination tubes. Referencing was to a ghost reference generated from the <sup>1</sup>H resonance frequency of the internal standard (measured at 99.55 MHz) by multiplying by the larmor ratio 1.06663974.<sup>[5]</sup> <sup>3</sup>H-<sup>3</sup>H lines coupling were noticed for the higher radioactivity. Its accurate <sup>3</sup>H labelling positions on phenyl ring were assigned by the coupling constants of ortho positions, para positions and meta positions, usually, coupling constants, J Hz, in the monosubstituted benzenes were ortho positions of 7— 10 Hz, para position of 0— 1 Hz and meta positions on <sup>3</sup>H-phenyl ring were ortho < para < meta. According to the coupling constants and the chemical shifts and integral area of each chemical shift, the distribution of labelling on phenyl ring were obtained

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in Table 2. It can be seen that ring was not uniformly labelled with tritium, the integrated line intensities have given a quantitative tritium distribution on the each site of phenyl ring. All these results showed <sup>3</sup>H NMR spectroscopy is ar especially attractive technique for determination of structure and position of H labelled compounds.



I R=OH. <sup>3</sup>H-Anisodamine hydrobromide II R'=OH: <sup>3</sup>H-Anisodine hydrobromide III R=H: <sup>3</sup>H-Atropine sulfate IV R'=H: <sup>3</sup>H-Scopolamine hydrobromide \* = <sup>3</sup>H

Fig.1 The structure formulas of belladonna alkaloids

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